Amended Final Report on the Safety Assessment of Glyceryl Dilaurate, Glyceryl Diarachidate, Glyceryl Dibehenate, Glyceryl Dierucate, Glyceryl Dihydroxystearate, Glyceryl Diisopalmitate, Glyceryl Diisostearate, Glyceryl Dilinoleate, Glyceryl Dimyristate, Glyceryl Dioleate, Glyceryl Dipalmitate, Glyceryl Dipalmitate, Glyceryl Dipalmitate, Glyceryl Dipalmitate Lactate, Glyceryl Stearate Citrate, Glyceryl Stearate Lactate, and Glyceryl Stearate Succinate¹

Glyceryl Dilaurate, Glyceryl Diarachidate, Glyceryl Dibehenate, Glyceryl Dierucate, Glyceryl Dihydroxystearate, Glyceryl Diisopalmitate, Glyceryl Diisostearate, Glyceryl Dilinoleate, Glyceryl Dimyristate, Glyceryl Dioleate, Glyceryl Diricinoleate, Glyceryl Dipalmitate, Glyceryl Dipalmitoleate, Glyceryl Distearate, Glyceryl Palmitate Lactate, Glyceryl Stearate Citrate, Glyceryl Stearate Lactate, and Glyceryl Stearate Succinate are diacylglycerols (also known as diglycerides or glyceryl diesters) that function as skin conditioning agents-emollients in cosmetics. Only Glyceryl Dilaurate (up to 5%), Glyceryl Diisostearate (up to 43%), Glyceryl Dioleate (up to 2%), Glyceryl Distearate (up to 7%), and Glyceryl Stearate Lactate (up to 5%) are reported to be in current use. Production proceeds from fully refined vegetable oils, which are further processed using hydrogenation and fractionation techniques, and the end products are produced by reacting selected mixtures of the partly hydrogenated, partly fractionated oils and fats with vegetable-derived glycerine to yield partial glycerides. In the final stage of the production process, the products are purified by deodorization, which effectively removes pesticide residues and lower boiling residues such as residues of halogenated solvents and aromatic solvents. Diglycerides have been approved by the Food and Drug Administration (FDA) for use as indirect food additives. Nominally, these ingredients are 1,3-diglycerides, but are easily isomerized to the 1,2-diglycerides form. The 1,3-diglyceride isomer is not a significant toxicant in acute, short-term, subchronic, or chronic animal tests. Glyceryl Dilaurate was a mild primary irritant in albino rabbits, but not a skin sensitizer in guinea pig maximization tests. Diacylglycerol Oil was not genotoxic in the Ames test, in mammalian Chinese hamster lung cells, or in a rodent bone marrow micronucleus assay. An eye shadow containing 1.5% Glyceryl Dilaurate did not induce skin irritation in a single insult patch

test, but mild skin irritation reactions to a foundation containing the same concentration were observed. A trade mixture containing an unspecified concentration of Glyceryl Dibehenate did not induce irritation or significant cutaneous intolerance in a 48-h occlusive patch test. In maximization tests, neither an eye shadow nor a foundation containing 1.5% Glyceryl Dilaurate was a skin sensitizer. Sensitization was not induced in subjects patch tested with 50% w/w Glyceryl Dioleate in a repeated insult, occlusive patch test. Glyceryl Palmitate Lactate (50% w/v) did not induce skin irritation or sensitization in subjects patch tested in a repeat-insult patch test. Phototoxicity or photoallergenicity was not induced in healthy volunteers tested with a lipstick containing 1.0% Glyceryl Rosinate. Two diacylglycerols, 1-oleoyl-2-acetoyl-sn-glycerol and 1,2-dipalmitoyl-sn-glycerol, did not alter cell proliferation (as determined by DNA synthesis) in normal human dermal fibroblasts in vitro at doses up to 10 μ g/ml. In the absence of initiation, Glyceryl Distearate induced a moderate hyperplastic response in randomly bred mice of a tumor-resistant strain, and with 9,10-dimethyl-1,2benzanthracene (DMBA) initiation, an increase in the total cell count was observed. In a glyceryl monoester study, a single application of DMBA to the skin followed by 5% Glyceryl Stearate twice weekly produced no tumors, but slight epidermal hyperplasia at the site of application. Glyceryl Dioleate induced transformation in 3-methylcholanthrene-initiated BALB/3T3 A31-1-1 cloned cells in vitro. A tumor-promoting dosing regimen that consisted of multiple applications of 10 μ mol of a 1,2-diacylglycerol (sn-1,2didecanoylglycerol) to female mice twice daily for 1 week caused more than a 60% decrease in protein kinase C (PKC) activity and marked epidermal hyperplasia. Applications of 10 μ mol sn-1,2didecanoylglycerol twice weekly for 1 week caused a decrease in cytosolic PKC activity, an increase in particulate PKC activity, and no epidermal hyperplasia. In studies of the tumor-promoting activity of 1,2-diacylglycerols, dose and the exposure regimen by which the dose is delivered play a role in tumor promotion. The 1,2-diacylglycerol-induced activation of PKC may also relate to the saturation of the fatty acid in the 1 or 2 position; 1,2-Diacylglycerols with two saturated fatty acids are less effective. Also, the activity of 1,2-diacylglycerols may be reduced when the fatty acid moiety in the structure is a long-chain fatty acid. A histological evaluation

was performed on human skin from female volunteers (18 to 56

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¹Reviewed by the Cosmetic Ingredient Review Expert Panel.

years old) who had applied a prototype lotion or placebo formulation, both containing 0.5% Glyceryl Dilaurate, consecutively for 16 weeks or 21 weeks. Skin irritation was not observed in any of the subjects tested. Biopsies (2 mm) taken from both legs of five subjects indicated no recognizable abnormalities of the skin; the epidermis was normal in thickness, and there was no evidence of scaling, inflammation, or neoplasms in any of the tissues that were evaluated. The Cosmetic Ingredient Review (CIR) Expert Panel considered that the available safety test data indicate that diglycerides in the 1,3-diester form do not present any significant acute toxicity risk, nor are these ingredients irritating, sensitizing, or photosensitizing. Whereas no data are available regarding reproductive or developmental toxicity, there is no reason to suspect any such toxicity because the dermal absorption of these chemicals is negligible. The Panel noted that these nominally 1,3-diglycerides contain 1,2diglycerides, raising the concern that 1,2-diglycerides could potentially induce hyperplasia. Data regarding the induction of PKC and the tumor promotion potential of 1,2-diacylglycerols increased the level of concern. Most of the diglycerides considered in this safety assessment, however, have fatty acid chains longer than 14 carbons and none have mixed saturated/unsaturated fatty acid moieties. The Panel considered it particularly important that a 21-week use study of a prototype lotion containing 0.5% Glyceryl Dilaurate (a 14-carbon chain fatty acid) indicated no evidence of scaling, inflammation, or neoplasms in biopsy specimens. Also, DNA synthesis assays on Glyceryl Dilaurate and Glyceryl Distearate indicated that neither chemical altered cell proliferation (as determined by DNA synthesis) in normal human dermal fibroblasts in vitro at doses up to 10 μ g/ml. The Panel understands that use testing is a common practice in industry and, if histopathology data are collected, the Panel believes that such an approach can demonstrate an absence of epidermal hyperplasia. Because the concentration of these ingredients can vary (up to 43% for Glyceryl Diisostearate in lipstick), the frequency of application can be several times daily, and the proportion of diglycerides that are inactive 1,3 isomers versus potentially biologically active 1,2 isomers is unknown, the Panel believes that each use should be examined to ensure the absence of epidermal hyperplasia during product development and testing. In the absence of inhalation toxicity data on the Glyceryl Diesters in this safety assessment, the Panel determined that these ingredients can be used safely in aerosolized products because they are not respirable. The Panel recognizes that certain ingredients in this group are reportedly used in a given product category, but the concentration of use is not available. For other ingredients in this group, information regarding use concentration for specific product categories is provided, but the number of such products is not known. In still other cases, an ingredient is not in current use, but may be used in the future. Although there are gaps in knowledge about product use, the overall information available on the types of products in which these ingredients are used and at what concentration indicate a pattern of use. Within this overall pattern of use, the CIR Expert Panel considers all ingredients in this group to be safe.

INTRODUCTION

This report addresses the safety of the following cosmetic ingredients:

Glyceryl Dilaurate, Glyceryl Dioleate, Glyceryl Diarachidate, Glyceryl Diricinoleate,

Glyceryl Dibehenate,

Glyceryl Dipalmitate,

Glyceryl Dierucate,

Glyceryl Dipalmitoleate,

Glyceryl Dihydroxystearate,

Glyceryl Distearate,

Glyceryl Diisopalmitate,

Glyceryl Palmitate Lactate,

Glyceryl Diisostearate,

Glyceryl Stearate Citrate,

Glyceryl Dilinoleate,

Glyceryl Stearate Lactate,

Glyceryl Dimyristate, and

Glyceryl Stearate Succinate.

These ingredients are commonly referred to as diacylglycerols, diglycerides, or glyceryl diesters, and are used in cosmetics as skin-conditioning agents—emollients. Not all are in current use. Of those in current use, safety test data are available on: Glyceryl Diisostearate, Glyceryl Dilaurate, Glyceryl Dioleate, Glyceryl Distearate, and Glyceryl Palmitate Lactate. Glyceryl Stearate Lactate is in current use, but no safety test data are available.

Other related assessments of safety have been done. For example, glycerin and glycerides are classified as generally recognized as safe (GRAS) food ingredients (Informatics, Inc. 1973; Federation Of American Societies For Experimental Biology 1975).

Related Cosmetic Ingredient Safety Assessments

The Cosmetic Ingredient Review (CIR) found Isostearic Acid safe in the present practices of use (Elder 1983). Oleic, Lauric, Palmitic, Myristic, and Stearic Acids were found safe in the present practices of use and concentration (Elder 1987). The available data were insufficient to support the safety of Arachidonic Acid (Elder 1993). Lactic Acid was found safe for use with qualifications (Andersen 1998). Glyceryl Stearate and Stearate SE were found safe for topical application in the present practices of use and concentration (Elder 1982).

CIR found the following glyceryl monoesters safe in the present practices of use and concentration, except that the available data were insufficient to support the safety of Glyceryl Arachidonate (CIR 2001):

Glyceryl Laurate,

Glyceryl Lanolate,

Glyceryl Laurate SE,

Glyceryl Linoleate,

Glyceryl Laurate/Oleate,

Glyceryl Linolenate,

Glyceryl Adipate,

Glyceryl Montanate,

Glyceryl Alginate,

Glyceryl Myristate,

Glyceryl Arachidate,

Glyceryl Isotridecanoate/Stearate/Adipate,

Glyceryl Behenate,

Glyceryl Caprate,

Glyceryl Oleate SE,

Glyceryl Caprylate,

Glyceryl Oleate/Elaidate,

Glyceryl Caprylate/Caprate,

Glyceryl Palmitate,

Glyceryl Citrate/Lactate/Linoleate/Oleate,

Glyceryl Palmitate/Stearate,

Glyceryl Palmitoleate,

Glyceryl Cocoate,

Gyceryl Pentadecanoate,

Glyceryl Collagenate,

Glyceryl Polyacrylate,

Glyceryl Erucate,

Glyceryl Rosinate,

Glyceryl Hydrogenated Rosinate,

Glyceryl Sesquioleate,

Glyceryl Hydrogenated Soyate,

Glyceryl/Sorbitol Oleate/Hydroxystearate,

Glyceryl Hydroxystearate,

Glyceryl Isopalmitate,

Glyceryl Stearate/Acetate,

Glyceryl Isostearate,

Glyceryl Stearate/Maleate,

Glyceryl Isostearate/Myristate,

Glyceryl Tallowate,

Glyceryl Isostearates,

Glyceryl Thiopropionate, and

Glyceryl Undecylenate.

CIR found the following glyceryl triesters to be safe as used (Andersen 2001):

Trilaurin.

Trimyristin,

Triarachidin,

Trioctanoin,

Tribehenin,

Triolein,

Tricaprin,

Tripalmitin,

Tricaprylin,

Tripalmitolein,

Trierucin,

Triricinolein,

Triheptanoin,

Tristearin,

Triheptylundecanoin,

Triundecanoin,

Triisononanoin.

Glyceryl Triacetyl Hydroxystearate,

Triisopalmitin,

Glyceryl Triacetyl Ricinoleate,

Triisostearin,

Glyceryl Stearate Diacetate, and

Trilinolein.

CIR has also found the following ingredients to be safe in the practices of use and concentration: Ricinus Communis (Castor) Seed Oil, Hydrogenated Castor Oil, Glyceryl Ricinoleate, Glyceryl Ricinoleate SE, Ricinoleic Acid, Potassium Ricinoleate, Sodium Ricinoleate, Zinc Ricinoleate, Cetyl Ricinoleate, Ethyl Ricinoleate, Glycol Ricinoleate, Isopropyl Ricinoleate, Methyl Ricinoleate, and Octyldodecyl Ricinoleate (CIR 2004).

CHEMISTRY

Chemical and Physical Properties

Properties of glyceryl diesters (Glyceryl Dipalmitate, Glyceryl Distearate, and Glyceryl Diisostearate) are included in Table 1.

Glyceryl Dilaurate (CAS No. 27638-00-2) is the diester of glycerin and lauric acid that conforms generally to the formula shown in Figure 1 (Pepe et al. 2002). According to the Scientific and Technical Information Network (STN) International (1997a) and Pepe et al. (2002), other names for Glyceryl Dilaurate include:

- Dilaurin:
- Dodecanoic Acid, Diester with 1,2,3-Propanetriol;
- Didodecanovl Glyceride;
- Dilauroyl Glyceride;
- Glycerine Dilaurate; and
- Glycerol Dilaurate.

Glyceryl Diarachidate (CAS Nos. 89648-24-8 and 127039-55-8) is the diester of glycerin and arachidic acid (q.v.) that generally conforms to the formula shown in Figure 2 (Pepe et al. 2002).

According to Pepe et al. (2002), other names for Glyceryl Diarachidate include:

$$\begin{array}{c} CH_{2}O \longrightarrow C(CH_{2})_{10}CH_{3} \\ \\ \\ \\ \\ C \longrightarrow OH \\ \\ \\ \\ CH_{2}O \longrightarrow C(CH_{2})_{10}CH_{3} \\ \\ \\ \\ \\ \\ \\ \\ \\ \end{array}$$

FIGURE 1 Glyceryl Dilaurate.

TABLE 1
Chemical and physical properties of glyceryl diesters.

	Ingredient			
Property	Glyceryl dioleate (Aarhus Oliefabrik 1997)	Glyceryl dipalmitate (Lide and Frederikse 1993)	Glyceryl distearate (Lide and Frederikse 1993)	Glyceryl diisostearate [Cosmetic, Toiletry, and Fragrance Association (CTFA) 1998]
Form	Beige, waxy solid (at 20°C) with bland	Crystals obtained from ethanol and	Needles or plates obtained from ether,	Pale yellow, oily liquid with faint,
	odor	chloroform as solvents	chloroform, and ligroin as solvents	characteristic odor
Molecular weight	_	568.92	625.03	
Density at 50°C	0.89-0.92 g/ml	_	_	_
Solubility	Insoluble in water	Soluble in ether	Soluble in ether	_
Vapor pressure at 200°C	<0.01 mm Hg	_	_	
Melting point	_	72–74°C	79.1°C	_
Slip melting point	35–39°C	_	_	_
Flash point	>200°C	_	_	_
Acid value	2	_	_	Not more than 10
Saponification value	172–183	_	_	175–195
Iodine value	55–66	_	_	
Hydroxyl value	140-190	_	_	50-130
Peroxide value (mEq/kg)	2	_	_	_
Loss on drying	_	_	_	Not more than 1%
Residue on ignition	_	_	_	Not more than 0.5%

- Eicosanoic Acid, Diester with 1,2,3-Propanetriol;
- Eicosanoic Acid, 1-(Hydroxymethyl)-1,2-Ethanediyl Ester;
- Glyceryl Diarachate; and
- 1-(Hydroxymethyl)-1,2-Ethanediyl Eicosanoate.

Glyceryl Dibehenate (CAS No. 99880-64-5) is the diester of glycerin and behenic acid (q.v.) that generally conforms to the formula shown in Figure 3 (Pepe et al. 2002).

Docosanoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

According to Pepe et al. (2002), Glyceryl Dibehenate is present in a trade mixture, Compritol 888 ATO, that also contains Tribehenin and Glyceryl Behenate. The International Nomenclature Cosmetic Ingredient (INCI) name for this mixture is Glyceryl Dibehenate/Tribehenin/Glyceryl Behenate. Data on the chemical and physical properties of Compritol 888 ATO are included in Table 2.

Compritol 888 ATO reportedly consists of 1-mono- glycerides (12% to 18%), free glycerol (<1.0%), water (<0.5%), and behenic acid (>80%) (Gattefossé 1998a, 1998b).

$$\begin{array}{c} CH_{2}O \longrightarrow C(CH_{2})_{18}CH_{3} \\ \\ \\ C \longrightarrow OH \\ \\ \\ CH_{2}O \longrightarrow C(CH_{2})_{18}CH_{3} \\ \\ \\ \end{array}$$

FIGURE 2 Glyceryl Diarachidate.

FIGURE 3 Glyceryl Dibehenate.

et al. 2002).

TABLE 2
Properties of compritol 888 ATO (Gattefossé 1998a, 1998b).

Property	Specification/typical values
Form	powder
Color	white-yellow (<5 on the Gardner scale)
Odor	faint
Solubility (at 20°C)	soluble in chloroform and methylene
•	chloride when heated;
	insoluble in water, n-hexane, ethanol,
	and mineral oils
Melting point	69–74°C
Drop point	69–74°C
Boiling point	>150°C
Flash point	>150°C
Acid value	<4.0 mg KOH/g
Iodine value	$< 3 \text{ g I}_2/100 \text{ g}$
Saponification value	145–165 mg KOH/g
Peroxide value	$< 6.0 \text{ mEq O}_2/\text{kg}$

Glyceryl Dierucate is the diester of glycerin and erucic acid (q.v.) that generally conforms to the formula shown in Figure 4. Docosenoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

Glyceryl Dihydroxystearate (CAS No. 122546-20-7) is the diester of glycerin and hydroxystearic acid (q.v.) that generally conforms to the formula shown in Figure 5 (Pepe et al. 2002).

According to Pepe et al. (2002), other names for this chemical include:

- 12-Hydroxyoctadecanoic Acid, 2-Hydroxy-1,3-Propanediyl;
- 2-Hydroxy-1,3-Propanediyl 12-Hydroxyoctadecanoate:
- Hydroxystearic Acid, Diester with 1,2,3-Propanetriol;
- Octadecanoic Acid, 12-Hydroxy-,2-Hydroxy-1,3-; and
- Propanediyl Ester.

Glyceryl Diisopalmitate is the diester of glycerin and a branched chain 16-carbon aliphatic acid that generally conforms to the formula shown in Figure 6. Isohexadecanoic Acid, Diester

$$\begin{array}{c} \text{O} \\ \parallel \\ \text{CH}_2\text{O} \longrightarrow \text{C} \text{ (CH}_2)_{11}\text{CH} \Longrightarrow \text{CH(CH}_2)_7\text{CH}_3 \\ \mid \\ \text{HCOH} \\ \mid \\ \text{CH}_2\text{O} \longrightarrow \text{C} \text{ (CH}_2)_{11}\text{CH} \Longrightarrow \text{CH(CH}_2)_7\text{CH}_3 \\ \mid \\ \text{O} \end{array}$$

FIGURE 4 Glyceryl Dierucate.

with 1,2,3-Propanetriol is another name for this chemical (Pepe

Glyceryl Diisostearate (CAS No. 68958-48-5) is the diester of glycerin and isostearic acid that generally conforms to the formula shown in Figure 7. Isooctadecanoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002)

Glyceryl Dilinoleate (CAS No. 30606-27-0) is the diester of glycerin and linoleic acid that generally conforms to the formula shown in Figure 8. 9,12-Octadecadienoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

Glyceryl Dimyristate (CAS No. 53563-63-6) is the diester of glycerin and myristic acid that generally conforms to the formula shown in Figure 9. Tetradecanoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

Glyceryl Dioleate (CAS No. 25637-84-7) is the diester of glycerin and oleic acid that generally conforms to the formula shown in Figure 10. 9-Octadecenoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

According to a chemical supplier, Glyceryl Dioleate is a mixture consisting of diglycerides (49%), monoglycerides (36%), and triglycerides (15%) (Aarhus Oliefabrik 1997).

Glyceryl Diricinoleate (CAS No. 27902-24-5) is the diester of glycerin and Ricinoleic Acid that generally conforms to the formula shown in Figure 11. Other names for this chemical include: Glycerol Diricinoleate; 12-Hydroxy-9-Octadecenoic

FIGURE 6Glyceryl Diisopalmitate.

$$\begin{array}{c} & & \bigcirc \\ & & \bigcirc \\$$

FIGURE 7 Glyceryl Diisostearate.

Acid, Diester with 1,2,3-Propanetriol; 9-Octadecenoic Acid, 12-Hydroxy-, Diester with 1,2,3-Propanetriol; and Ricinolein, di-(Pepe et al. 2002).

Glyceryl Dipalmitate (CAS No. 26657-95-4) is the diester of glycerin and palmitic acid that generally conforms to the formula shown in Figure 12. Hexadecanoic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

Glyceryl Dipalmitoleate is the diester of glycerin and palmitoleic acid that generally conforms to the formula shown in Figure 13. Palmitoleic Acid, Diester with 1,2,3-Propanetriol is another name for this chemical (Pepe et al. 2002).

Glyceryl Palmitate Lactate is the lactic acid ester of glyceryl palmitate that conforms to the formula shown in Figure 14 (Pepe et al. 2002). According to a chemical supplier, Glyceryl Palmitate Lactate is a lactic acid ester of mono-diglyceride made from edible, refined palmitic acid (Danisco Ingredients 1999a).

Glyceryl Distearate (CAS No. 1323-83-7) is the diester of glycerin and stearic acid that conforms to the formula shown in Figure 15. Octadecanoic Acid, Diester with 1,2,3-Propanetriol is another name for Glyceryl Distearate (Pepe et al. 2002).

Glyceryl Stearate Citrate (CAS No. 39175-72-9) is the citric acid ester of glyceryl stearate (q.v.) that conforms to the formula shown in Figure 16 (Pepe et al. 2002). Huls America, Inc. (no date) characterized Glyceryl Stearate Citrate as a partially neutralized ester of monoglycerides and diglycerides of saturated, edible fatty acids with citric acid. It contains 1-monoglyceride (10% to 30%) and free glycerol (maximum 3%).

$$\begin{array}{c} \text{CH}_2\text{O} \longrightarrow \text{C} \text{ (CH}_2)_7\text{CH} \longrightarrow \text{CHCH}_2\text{CH} \longrightarrow \text{CH(CH}_2)_4\text{CH}_3\\ | \\ \text{HCOH} \\ | \\ \text{CH}_2\text{O} \longrightarrow \text{C} \text{ (CH}_2)_7\text{CH} \longrightarrow \text{CHCH}_2\text{CH} \longrightarrow \text{CH(CH}_2)_4\text{CH}_3\\ | \\ \text{O} \end{array}$$

FIGURE 8
Glyceryl Dilinoleate.

$$\begin{array}{c} \text{O} \\ \text{CH}_2\text{O} \longrightarrow \text{C} \ (\text{CH}_2)_{12}\text{CH}_3 \\ \\ \text{HCOH} \\ \\ \text{CH}_2\text{O} \longrightarrow \text{C} \ (\text{CH}_2)_{12}\text{CH}_3 \\ \\ \text{O} \end{array}$$

FIGURE 9
Glyceryl Dimyristate

According to Pepe et al. (2002) two other names for Glyceryl Stearate Citrate are:

- 2-Hydroxy-1,2,3-Propanetricarboxylic Acid, Monoester with 1,2,3-Propanetriol Monooctadecanoate; and
- 1,2,3-Propanetricarboxylic Acid, 2-Hydroxy-, Monoester with 1,2,3-Propanetriol Monooctadecanoate.

Glyceryl Stearate Lactate is the lactic acid ester of glyceryl stearate (q.v.) that conforms to the formula shown in Figure 17 (Pepe et al. 2002).

Glyceryl Stearate Succinate is the succinic acid ester of glyceryl stearate (q.v.) (Pepe et al. 2002). No structure was given in the *International Cosmetic Ingredient Dictionary and Handbook*, but using the same convention as in the previous formulae, it likely conforms to that shown in Figure 18.

Analytical Methods

Capillary supercritical fluid chromatography has been used in the analysis of Glyceryl Dilaurate (Giron et al. 1992).

Methods of Production

Methods of production for most glyceryl diesters were not available. Aarhus Oliefabrik (1997) stated that the raw materials for the production of Glyceryl Dioleate are fully refined nonlauric vegetable oils. Glyceryl Dioleate is said to consist of C16–C18 glycerides manufactured from food grade raw materials. The oils are further processed using special hydrogenation

$$\begin{array}{c} O \\ \parallel \\ CH_2O \longrightarrow C \ (CH_2)_7CH \longrightarrow CH(CH_2)_7CH_3 \\ \parallel \\ CH_2O \longrightarrow C \ (CH_2)_7CH \longrightarrow CH(CH_2)_7CH_3 \\ \parallel \\ O \end{array}$$

FIGURE 10 Glyceryl Dioleate.

FIGURE 11 Glyceryl Diricinoleate.

CH₂O \longrightarrow C (CH₂)₅CH \Longrightarrow CH(CH₂)₇CH₃

HCOH

CH₂O \longrightarrow C (CH₂)₅CH \Longrightarrow CH(CH₂)₇CH₃

O

FIGURE 13

Glyceryl Dipalmitoleate.

and fractionation techniques. The end products are produced by reacting selected mixtures of the partly hydrogenated, partly fractionated oils and fats with vegetable-derived glycerine to yield partial glycerides. In the final stage of the production process, the products are purified by deodorization.

Composition

Diglyceride Isomers

DeGroot (1972) reported that mixtures of 1,2- and 1,3-diglycerides stored at temperatures below their melting points undergo a rather rapid isomerization of practically all of the 1,2-diglycerides to 1,3-isomers. Prior to storage, the mixtures consisted of 47% to 75% 1,3-diglycerides and 25% to 53% 1,2-diglycerides. Storage temperatures and the duration of storage for the individual mixtures ranged from 26°C to 65°C and 2 to 210 days, respectively. The resulting mixtures (after storage) contained 90% to 98% 1,3-diglycerides and 2% to 10% 1,2-diglycerides. This solid-phase isomerization was characterized as resulting in a substantial increase in the yield of saturated and unsaturated mono- and mixed-acid 1,3-diglycerides.

According to another source, 1,2- and 1,3-diglycerides are not stable and are easily isomerized under acidic, basic, or thermal conditions to an equilibrium of approximately 40% of the 1,2- and 60% of the 1,3-isomer (Serdarevich 1967).

Enkelund (1999) provided an interpretation of the above data. He stated that, at high temperature, the equilibrium mixture has a ratio of 1,3/1,2-diglycerides of approximately 75%:25%. At lower temperatures, the equilibrium will, depending on the constituent fatty acids, temperature, and time, favor the 1,3-diester. The 1,2-diester content will be lowered to the range of 2% to

10%. In practice, this means that during cooling, packaging, and storing of the ingredient, most of the 1,2-diglycerides are isomerized to the 1,3-isomer. The process is favored by the cosmetic producer's handling, as the material will normally be heated to 60° C to 75° C when mixing the material with other lipids and then cooled to room temperature.

More recent information was provided by International Specialty Products (ISP). Based on the industrial chemical processes that are used to produce the glyceryl fatty acid esters, some diesters will be present in any monoester material (on the order of 10%) and diesters will also be present in triester material (on the order of 5%). Likewise, monoesters and triesters will be present in any diester material (ISP 2001).

ISP (2001) estimated that between 30% and 50% of the diester composition will be as 1,2-diester, and the remainder, 1,3-diester. Aarhus Oliefabrik (2001), however, stated that the content of 1,2 isomers in glyceryl diester products such as Cremeol FR-36 (now Cegesoft FR 36) and Cremeol FR-57 (now Cegesoft FR 57) is unlikely to be more than a few percent and under no circumstances would it be 20% to 50%. Neither report indicated the presence of 2,3-diesters.

ISP (2002) performed analytical testing to determine the percentage of 1,2 diester fraction (1,2-dilauryl-s,n-glycerol) in Emulsynt GDL, glycerol laurate, material on an as-sold basis. They estimated that 10% to 15% 1,2 diester (characterized as optically and physiologically active isomer) is present in the product as sold, with 10% to 15% 2,3 diester (characterized as optically active, physiologically non-active isomer) and 70% to 80% 1,3 diester (characterized as not optically or physiologically active) comprising the remainder of the product.

$$\begin{array}{c} {\rm CH_2O} {\ \ } \\ {\rm CH_2O} {\ \ } \\ {\rm C} \ ({\rm CH_2})_{14} {\rm CH_3} \\ {\rm HCOH} \\ {\rm |} \\ {\rm CH_2O} {\ \ } {\ \ } \\ {\rm CH_2O} {\ \ } \\ {\rm CH_2O} {\ \ } \\ {\rm C} \end{array}$$

FIGURE 12
Glyceryl Dipalmitate.

FIGURE 14
Glyceryl Palmitate Lactate

$$\begin{array}{c} \text{O} \\ \text{CH}_2\text{O} & \stackrel{\parallel}{-} \text{C}(\text{CH}_2)_{16}\text{CH}_3 \\ | \\ \text{HCOH} \\ | \\ \text{CH}_2\text{O} & \stackrel{\parallel}{-} \text{C}(\text{CH}_2)_{16}\text{CH}_3 \\ | \\ \text{O} \end{array}$$

FIGURE 15 Glyceryl Distearate

Glyceryl Palmitate Lactate

According to a chemical supplier, the composition of Glyceryl Palmitate Lactate has been defined as follows: free glycerol (maximum 1%); lactic acid (15%); and palmitic acid content, of total fatty acids (>90%) (Danisco Ingredients 1996). In a more recent description, the specifications for Glyceryl Palmitate Lactate were: palmitic acid (minimum 90%), lactic acid content (13% to 16%), iodine value (maximum 1.50), saponification value (245 to 265), acid value (maximum 4), free glycerol (maximum 1%), dropping point ($\approx 50^{\circ}$ C), and form (pellets) (Danisco Ingredients 1999a). This source also stated that Glyceryl Palmitate Lactate contains 0.03% glycerol and 0.82% free fatty acid, and that the ratio of 1,2-(mono)glycerol diester to total (mono)glycerol diester is 30.6 (Danisco Ingredients 1999a, 1999b).

Glyceryl Stearate Lactate

According to Danisco Ingredients (1999a, 1999b), Glyceryl Stearate Lactate contains glycerol (0.24%) and free fatty acid (0.16%), and the ratio of 1,2-(mono)glycerol diester to total (mono)glycerol diester is 33:9.

Impurities

Glyceryl Dioleate

The final stage in the processing of oil, deodorization, effectively removes pesticide residues and lower boiling residues such as residues of halogenated solvents and aromatic solvents. Thus, the following impurities are not present at concentrations above the detection limits of the methodology used in any of the deodorized oils and fats produced by Aarhus Oliefabrik:

$$\begin{array}{c} \text{O} \\ \text{CH}_2\text{O} \longrightarrow \text{C} \, (\text{CH}_2)_{16}\text{CH}_3 \\ | \\ \text{HCOH} \\ | \\ \text{CH}_2\text{O} \longrightarrow \text{CCH}_2\text{CCH}_2\text{COOH} \\ | \\ \text{O} \end{array}$$

FIGURE 16 Glyceryl Stearate Citrate.

FIGURE 17 Glyceryl Stearate Lactate.

organochlorine pesticides; halogenated solvents and aromatic solvents; PCBs, organic solvents (e.g., ethanol and acetone), metals (heavy metals included), aflatoxin, ash, radioactive material, and microbial activities (Aarhus Oliefabrik 1998).

Glyceryl Dibehenate

Sulfated ashes (<0.10%) and heavy metals (<10 ppm) are impurities in a trade mixture (Compritol 888 ATO) containing Glyceryl Dibehenate (Gattefossé 1998a, 1998b).

Glyceryl Diisostearate

According to Cosmetic, Toiletry and Fragrance Association (CTFA) (1998), Glyceryl Diisostearate contains not more than 20 ppm heavy metals and not more than 2 ppm arsenic.

Glyceryl Palmitate Lactate

Specifications for heavy metal impurities in Glyceryl Palmitate Lactate are as follows: arsenic (As) (maximum 3 mg/kg), lead (Pb) (maximum 5 mg/kg), mercury (Hg) (maximum 1 mg/kg), cadmium (Cd) (maximum 1 mg/kg), and heavy metals (as Pb) (maximum 10 mg/kg) (Danisco Ingredients 1999a). Glyceryl Palmitate Lactate has been described as containing heavy metals (as Pb) at <10 mg/kg (Danisco Ingredients 1996).

Glyceryl Stearate Citrate

Huls America, Inc. (no date) stated that Glyceryl Stearate Citrate is free of polyethylene glycol and its monomers.

$$\begin{array}{c} \text{CH}_2\text{O} & \begin{array}{c} \text{O} \\ \\ \\ \\ \\ \\ \end{array} \\ \text{H} & \begin{array}{c} \text{C} \\ \\ \\ \\ \\ \end{array} \\ \text{CH}_2\text{O} & \begin{array}{c} \text{C}(\text{CH}_2)_{16}\text{CH}_3 \\ \\ \\ \\ \\ \\ \\ \end{array} \\ \begin{array}{c} \text{C}(\text{CH}_2)_{2}\text{COOH} \\ \\ \\ \\ \\ \\ \\ \\ \end{array}$$

FIGURE 18 Glyceryl Stearate Succinate.

Stability/Reactivity

Glyceryl Dibehenate

A trade mixture (Compritol 888 ATO) containing an unspecified concentration of Glyceryl Dibehenate reacts with strong acids and oxidizing agents (dangerous reactions) (Gattefossé 1998b).

Glyceryl Dioleate

According to Aarhus Oliefabrik (1997), Glyceryl Dioleate is neither explosive or autoflammable, but pro-oxidants and oxidation promoting conditions should be avoided. Glyceryl Dioleate does not have hazardous decomposition products; water and carbon dioxide are the thermal decomposition products.

USE

Cosmetic

These ingredients function as skin conditioning agents—emollients in cosmetic products (Pepe et al. 2002). Voluntary industry reports to the Food and Drug Administration (FDA) in 2002 indicate that only 3 of the 18 ingredients in this safety assessment were used, Glyceryl Dilaurate, Glyceryl Diisostearate, and Glyceryl Dioleate (FDA 2002). The reported uses as a function of product type are given in Table 3. Where uses in a product were reported to FDA, column 1 also gives the total number of such product types; e.g., Glyceryl Dilaurate is used in 1 of 324 foundations, based on industry reports to FDA.

As shown in Table 3, concentration of use data provided by CTFA in 1999, based on an industry survey, confirm that Glyceryl Dilaurate, Glyceryl Diisostearate, and Glyceryl Dioleate are in current use, but also indicate that Glyceryl Distearate and Glyceryl Stearate Lactate are currently used in cosmetics (CTFA 1999a).

As can be seen in Table 3, certain ingredients in this group are reportedly used in a given product category, but the concentration of use is not available. For other ingredients in this group, information regarding use concentration for specific product categories is provided, but the number of such products is not known. In still other cases, an ingredient is not in current use, but may be used in the future. Although there are gaps in knowledge about product use, the overall information available on the types of products in which these ingredients are used and at what concentration indicate a pattern of use.

Of the 18 ingredients reviewed in this report, none are included among the substances listed as prohibited from use in cosmetic products marketed in the European Union (European Economic Community 2001).

According to the Ministry or Health, Labor and Welfare (MHLW), Glyceryl Diesters reviewed in this report are not included on the list of ingredients that must not be combined in cosmetic products or on the list of restricted ingredients for cosmetic products that are marketed in Japan (MHLW 2001a, 2001b).

Cosmetic products containing Glyceryl Diesters are applied to most areas of the body, and could come in contact with the ocular and nasal mucosae. These products could be used on a daily basis, and could be applied frequently over a period of several years. Product categories in which Glyceryl Diesters are being used include potential aerosol applications. Bower (1999) characterized the typical diameter of anhydrous hair spray particles in the 60- to 80- μ m range (typically, <1% are below 10 μ m). Pump hair sprays, in contrast, have typical particle diameters of \geq 80 μ m. Johnsen (2004) reported that the mean particle diameter is around 38 μ m in a typical aerosol spray. In practice, he stated that aerosols should have at least 99% of particle diameters in the 10- to 110- μ m range. For comparison, Jensen and O'Brien (1993) reported a mean aerodynamic diameter of 4.25 \pm 1.5 μ m for respirable particles.

Noncosmetic

The safety of mono- and diglycerides in food has been reviewed by the Food Protection Committee of the National Academy of Sciences National Research Council Food and Nutrition Board (National Academy of Sciences 1960). The Food Protection Committee concluded that there appears to be no reason to question the safety of mono-, di-, or triglycerides of lauric acid (i.e., Glyceryl Laurate, Glyceryl Dilaurate, or Glyceryl Trilaurate [Trilaurin]) as food additives. This conclusion was based on the following: (1) Lauric acid glycerides are found in important foods, such as human and cow's milk, at concentrations of 3% to 6% and in large quantities in coconut oil, without recognized toxic effects; (2) Lauric acid glycerides undergo the usual metabolic changes of the higher fatty acids; (3) When lauric acid glycerides are fed in diets containing a variety of glycerides, there is no evidence of a specific toxic or harmful effect. Soni et al. (2001) reported that diacylglycerol oil has been approved for use in cooking oil in Japan.

Glyceryl Diesters have been approved for use as components of adhesives, paper and paperboard, and other materials that come in contact with food (i.e., indirect food additive uses): (21 CFR 175.105; 176.210, and 177.2800).

Glyceryl Dilaurate has been detected in pharmaceutical excipients (Giron et al. 1992).

BIOLOGICAL ACTIVITY

Metabolism

Glyceryl diesters (diglycerides) are intermediates in the metabolism of triglycerides to free fatty acids and glycerol. Triglyceride digestion begins in the intestinal tract. Initially, the triglyceride is hydrolyzed enzymatically to α,β -diglyceride, which is then hydrolyzed to β -monoglyceride in the lumen of the intestines. These hydrolytic reactions occur at an oil-water interface. Approximately 28% of the β -monoglyceride is isomerized to α -monoglyceride, and approximately 75% of the α -monoglyceride is further hydrolyzed to free glycerol. Free

TABLE 3 Product formulation data on diglycerides.

Product category (number of formulations reported) (FDA 2002)	Number of formulations containing ingredient (FDA 2002)	Current concentration of use (CTFA 1999a) (%)
Glyceryl Dilai	ırate	
Eye makeup		
Eye shadow	_	2
Makeup		
Foundations (324)	1	_
Lipsticks (962)	1	_
Makeup bases (141)	3	_
Other makeup preparations (201)	1	_
Bath preparations		
Bath soaps and detergents (421)	3	0.02
Skin care preparations	J	0.02
Cleansing creams, lotions, liquids, and pads (775)	3	_
Body and hand skin care preparations (840)	13	2–4
Moisturizers (905)	5	2
Night skin care preparations (200)	2	5
Paste masks [mud packs] (271)	$\frac{2}{2}$	5
Suntan Preparations	2	3
Suntan Fleparations Suntan gels, creams, and liquids (131)	1	
	35	0.02-5
2002 total uses/concentration range for Glyceryl Dilaurate	33	0.02–3
Glyceryl Diisosi	earate	
Fragrance preparations		
Perfumes (235)	1	18
Makeup		
Lipsticks (962)	93	43
Nail care products		
Nail polish and enamel (123)	1	_
Skin care preparations		
Body and hand skin care preparations (840)	1	_
Moisturizers (905)	1	_
2002 total uses/concentration range for Glyceryl Diisostearate	97	18–43
Glyceryl Diol	eate	
Makeup		
Foundations	_	2
Skin care preparations		
Face and neck skin care preparations	_	0.8
Moisturizers	_	2
Other skin care preparations (725)	1	<u>-</u>
2002 total uses/concentration range for Glyceryl Dioleate	1	0.8–2
	-	0.0 2
Glyceryl Dister	arate	
Eye makeup		0.02.05
Eye lotion	_	0.02–0.5
Mascara	_	0.003

TABLE 3 Product formulation data on diglycerides (*Continued*).

Product category (number of formulations reported) (FDA 2002)	Number of formulations containing ingredient (FDA 2002)	Current concentration of use (CTFA 1999a) (%)
Glyceryl Di.	stearate	
Noncoloring hair preparations		
Shampoos	_	6
Makeup		
Foundations	_	7
Lipstick	_	7
Nail care products		
Cuticle softeners	_	0.02
Nail creams and lotions	_	0.02
Skin care preparations		
Cleansing creams, lotions, liquids, and pads	_	0.0003
Face and neck skin care preparations	_	0.2 - 0.5
Body and hand skin care preparations	_	7
Moisturizers	_	0.2-7
Skin fresheners	_	0.00003
2002 total uses/concentration range for Glyceryl Distearate	_	0.00003-7
Glyceryl Stear	ate Lactate	
Personal hygiene products		
Underarm deodorants	_	5
Skin care preparations		
Face and neck skin care preparations	_	5
Moisturizers	_	5
2002 total uses/concentration range for Glyceryl Stearate Lacta	ite —	5

glycerol enters the intestinal wall independent of the lipids, and there is no further use of it in terms of lipid absorption. The free fatty acids and glycerol are available for the resynthesis of triglycerides. β -monoglycerides are not hydrolyzed because of their transfer to a water-soluble phase and, also, because of enzyme specificity. However, they can be acylated directly to triglyceride (Mattson and Volpenhein 1964).

Percutaneous Absorption

Information on the percutaneous absorption of Glyceryl Diesters included in this review was not found in the published literature. However, $C \log P$ (octanol/water partition coefficient) values that were calculated for Glyceryl Dilaurate (10.4) and Glyceryl Distearate (16.7) have been interpreted to indicate that these two Glyceryl Diesters would not be well-absorbed (FDA 2000).

Skin Penetration Enhancement

Glyceryl Dilaurate

Aungst et al. (1986) evaluated the effect of Glyceryl Dilaurate or Glyceryl Laurate on the penetration of Naloxone-HCl across cadaver skin using Franz diffusion cells. Naloxone is a

potent opioid antagonist used for the reversal of narcosis. Naloxone concentrations in the reservoir were determined by high-performance liquid chromatography (HPLC) using ultraviolet (UV) detection. Glyceryl Dilaurate was evaluated at a concentration of 10% in propylene glycol. The average flux through human cadaver skin (10 experiments) for Naloxone alone was $1.6 \pm 0.4~\mu g/cm^2 \cdot h^{-1}$. In the presence of Glyceryl Dilaurate, average Naloxone flux increased to $18.7 \pm 1.8~\mu g/cm^2 \cdot h^{-1}$ (3 experiments). In the presence of urea (10% in propylene glycol), the average Naloxone flux was $0.4 \pm 0.1~\mu g/cm^2 \cdot h^{-1}$ (3 experiments).

Effect on Enzyme Activity

Because of the established link between prostate cancer and high dietary fat intake, Niederpruem et al. (1995) evaluated the effect of Glyceryl Dilaurate, various fatty acids, and their derivatives on 5α -reductase activity in vitro. Prostate gland tissue specimens (human) were used. The reduction of testosterone to dihydrotestosterone, which controls cellular division in the prostate gland, is catalyzed by 5α -reductase. The authors reported that it has been suggested that the modulation/inhibition of this enzyme could prevent carcinogenesis in the prostate gland. Results

indicated that the inhibitory effect of lauric acid on 5α -reductase activity was totally lost by esterification to Glyceryl Dilaurate.

Effect on Signal Transduction

According to Lee and Severson (1994), the generation of intracellular second messengers is a common mechanism of signal transduction for external stimuli such as hormones, neurotransmitters, growth factors, and drugs (agonists) that interact with plasma membrane receptors. The established role of phospholipid turnover in signal transduction mechanisms is based on the observations that agonist-induced hydrolysis of a minor phospholipid in the plasma membrane, phosphatidylinositol 4,5-bisphosphate (PIP₂), in a reaction catalyzed by a phosphoinositide-specific phospholipase C (PI-PLC) enzyme generated the following two intracellular second messengers: (1) inositol 1,4,5-triphosphate (IP₃), responsible for the mobilization of Ca²⁺ from intracellular stores, and (2) diacylglycerol, responsible for the activation of PKC.

As stated by Lévy et al. (1994) these isoenzymes transduce mitotic signals induced by growth factors. PKC α , as well as other isoenzymes in this family, are activated by Ca²⁺, phosphatidylserine, and diacylglycerols. Diacylglycerols are considered to be hydrophobic anchors that may recruit PKC to the membrane, leading to an increase in the enzyme's membrane affinity and to the activation of PKC.

Sánchez-Piñera et al. (1999) studied the lipid activation of PKC α by comparing the activation capacity of different 1,2-diacylglycerols and 1,3-diacylglycerols incorporated into mixed micelles or vesicles. The authors noted that PKC isoenzymes are a large family of serine/threonine kinases that are involved in cellular signaling. PKC consists of a family of at least ten isozymes that phosphorylate serine and threonine residues. Classical PKC isozymes (α , β_I , β_{II} , γ) are dependent on Ca²⁺and phospholipids and are activated by diacylglycerol.

Unsaturated 1,2-diacylglycerols were more potent activators of PKC α than saturated 1,2-diacylglycerols when 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphoserine (POPS)/Triton X-100 mixed micelles and pure POPS vesicles were used. These differences were not observed when 1-palmitoyl -2-oleoyl-sn-glycero-3-phosphocholine (POPC)/POPS (4:1 molar ratio) vesicles were used. Additionally, 1,2-diacylglycerols had a considerably greater activating capacity than 1,3-diacylglycerols in POPS Triton X-100 mixed micelles and in POPC/POPS vesicles.

The difference between 1,2- and 1,3-diacylglycerols was smaller when pure POPS vesicles were used. That is, both were able to activate PKC α practically to the same extent. Nevertheless, saturated diacylglycerols induced significant activation of PKC α in Triton X-100 micelles and in pure POPS vesicles in this study (Sánchez-Piñera et al. 1999).

Unlike the preceding study, a very low capacity for 1,3-diacylglycerol-induced activation was demonstrated in an earlier study in which sonicated phosphatidylserine vesicles were used (Nomura et al. 1986). Unsonicated preparations of multilamellar vesicles were used in the Sánchez-Piñera et al. (1999) study.

PKC, Cell Growth, and Proliferation

Blobe et al. (1994) stated that studies have indicated that the sustained activation or inhibition of PKC (diacylglycerolactivated isoenzyme) activity in vivo may play a critical role in the regulation of long-term cellular events such as proliferation, differentiation, and tumorigenesis. Many of the signals transduced by PKC are mitogenic and the result of growth factors (e.g., platelet-derived growth factor [PDGF] and epidermal growth factor [EGF]). For example, PDGF binds to its high affinity receptor (PDGFR) and activates this receptor's intrinsic tyrosine kinase to mediate the initiation of DNA synthesis and other cellular effects. It is important to note that PKC has been linked directly to the pathogenesis of several human cancers, including skin, colon, and breast cancers. In the epithelium, long-term changes in PKC activity, either through the action of phorbol esters or by specific changes in PKC isoenzyme activities, leads to growth of melanocytes, the differentiation of keratinocytes, or cellular transformation. In colon cancer, PKC acts as a tumor suppressor. In breast cancer, an increase in PKC activity appears to correlate with enhanced oncogenicity. Thus, the authors concluded that decreasing the PKC activity can result in cellular transformation.

Wang and Smart (1999) stated that the overexpression of PKC α in the epidermis of transgenic mice increases the expression of specific proinflammatory mediators and induces cutaneous inflammation, but has little to no effect on epidermal differentiation, proliferation, or 12-O-tetradecanoylphorbol-13-acetate (TPA) tumor promotion.

Diacylglycerols and PKC

In a study reported by Hansen et al. (1990) involving female CD-1 mice, PKC was linked to epidermal hyperplasia and tumor promotion. In this study, a tumor-promoting dosing regimen consisting of multiple applications of 5 or 10 μ mol sn-1,2-didecanoylglycerol, a model sn-1,2-diacylglycerol and complete tumor promoter, twice daily for 1 week caused more than a 60% decrease in cytosolic and particulate PKC activity and marked epidermal hyperplasia. However, two applications of 10 μ mol sn-1,2-didecanoylglycerol for 1 week, a nonpromoting dosing rate, caused a decrease in cytosolic PKC activity, increased particulate PKC activity, but did not induce hyperplasia. These data, together with other study results, demonstrated to the authors that the down-regulation of epidermal PKC is associated with and may be a permissive event for epidermal hyperplasia and tumor promotion (Hansen et al. 1990).

Competitive Binding to PKC by Diacylglcerol Analogues

Sharkey et al. (1984) reported that the phorbol ester receptor in brain may be the same as the Ca²⁺-phospholipid-dependent kinase (PKC) receptor. Assays were performed using the mouse brain cytosolic phorbol ester aporeceptor, which requires phospholipids for activity. In the presence of phosphatidylserine, 1,2-diolein inhibited the specific binding of [³H]phorbol 12,

13-dibutyrate ([³H]PBt₂) in a dose-dependent fashion. Other short-chain saturated diacylglycerol derivatives, 1,3-dicaprylin and 1,3-dicaproin, also inhibited ([³H]PBt₂) binding. However, the long-chain saturated derivatives 1,2-dipalmitin and 1,2-distearin were much less active.

König et al. (1985) also reported that diacylglycerol stimulates PKC in a manner that is similar to that of the phorbol esters and inhibits phorbol ester binding.

Sharkey and Blumberg (1985) stated that the diacylglycerol 1,2-diolein inhibits binding of the phorbol ester [20-³H]phorbol 12,13-dibutyrate to PKC. These authors also reported the inhibition of phorbol ester binding to reconstituted PKC by the diglyceride glycerol 1-myristate 2-acetate (Sharkey and Blumberg 1986).

Wender et al. (1986) performed computer modeling of the diterpene diester phorbol 12-myristate 13-acetate and other structural classes of tumor promoters. The authors stated that the results indicated a marked similarity in the relative positions of certain heteroatoms and hydrophobic groups. For phorbol esters, this mapping consists of the C-4, C-9, and C-20 hydroxyl groups and a hydrophobic region that is filled by a long-chain acyl functional group attached to the C-12 or the C-13 positions. Diacylglycerols, thought to be the endogenous activators of the major phorbol ester receptor PKC, fit this model in a stereospecific fashion.

Lee et al. (1996a, 1996b) and Sharma et al. (1996) also stated that PKC interaction with diacylglycerol analogues is stereo specific.

Caloca et al. (1999) reported that the binding of diacylglycerol and related analogs occurs at C1 domains (also called cysteine-rich regions or zinc fingers) that are present in PKC isozymes.

According to Lee et al. (1993) and Bögi et al. (1998), phorbol esters, potent tumor promoters, function as ultrapotent analogues of 1,2-diacylglycerol.

Diacylglycerols and UV Light

Punnonen and Yuspa (1992) reported that the irradiation of cultured murine keratinocytes with UVB light (270 to 380 nm; 20 to 120 J/m²) induced a dose-dependent increase in cellular concentrations of diacylglycerols. The turnover of other phospholipids (e.g., phosphatidylcholine and phosphatidylethanolamine) was unaffected. These results were reported at 24 h post irradiation. The UV light-induced increase in cellular diacylglycerols (specific chemical names not stated) was accompanied by changes in diacylglycerol kinase (DAG-kinase). Cytosolic DAG-kinase activity and the activity of DAG-kinase in the membrane fraction were decreased and increased, respectively. The authors suggested that irradiation with UVB light increases DAG concentrations via changes in de novo metabolism that involve a DAG-kinase pathway. Furthermore, it was suggested that elevated diacylglycerol may influence signal transduction pathways that are mediated by cellular lipids and contributes to keratinocyte responses to UV light.

Agin et al. (1991) studied the effect of topically applied diacylglycerols, 1,2-dioctanoyl-sn-glycerol (DOG) and 1-oleyl-2acetyl-sn-glycerol (OAG), on melanogenesis using female Skh HR-2 pigmented, hairless mice (ages 4 to 6 weeks). Each diacylglycerol was dissolved in acetone and applied (120 μ l doses, using micropipette) to the total exposed area of dorsal skin, approximately 25 cm². Additionally, some of the groups were then irradiated with a 0.8 minimum erythemal dose of UVA/UVB light. The light source was a Kodacel 401 filtered FS-20 lamp bank that delivered 134 mJ/cm² of UVB light (290 to 320 nm) and 146 mJ/cm² of UVA light (320 to 400 nm). Four different test protocols (experiments) incorporating this application procedure and study results were conducted. At the end of each treatment, dorsal skin was removed and stained for melanin and 1-dopa. The epidermis was then separated and prepared for microscopic examination.

In the first experiment, four groups of 10 mice were treated with 0.18, 0.88, 1.75, and 3.5 μ mol DOG in acetone, respectively, 3 days per week (Mondays, Wednesdays, and Fridays) for 3 weeks. A fifth group was treated with acetone (vehicle) only. A marked, dose-dependent increase in dopa-positive areas in the whole epidermis was noted, having reached a maximum at a dose of 1.75 μ mol. There were no visual signs of skin irritation at either of the doses administered. Few highly stained dopa-positive melanocytes were observed in untreated control epidermal tissue. The analysis of sectioned skin for melanin deposition revealed that treatment with DOG induced negligible enhancement of melanin deposition.

In the second experiment, four groups of 10 mice were treated with either vehicle alone or 1.75 μ mol DOG in acetone twice weekly (Tuesdays and Thursdays). Additionally, the animals either were or were not irradiated three times weekly (Mondays, Wednesdays, and Fridays) for 3 weeks. Treatment with DOG alone did not significantly affect the dopa-positive area and melanin deposition. UV light alone induced a 5.4-fold increase in the area that was dopa-positive. However, compared to controls, when DOG treatment was combined with UV exposure, the dopa-positive area increased 10-fold. The sum of DOG treatment alone plus UV light treatment alone amounted to an increase in the dopa-positive area that was less than that observed following the combined treatment, suggesting a synergistic effect. For melanin deposition, a 7.2-fold increase over that observed in the presence of UV light alone was reported. Hematoxylin and eosin and Mowry's staining of skin sections treated with DOG or DOG plus UV light revealed no skin irritation or mucopolysaccharide deposition that was in excess of that anticipated from exposure to UV light alone. Additionally, DOG alone did not induce hyperplasia of the stratum corneum or epidermis.

In the third experiment, diacylglycerol or vehicle was applied to groups of 10 mice on 5 consecutive days (Monday through Friday). Beginning on the following Monday, three of

the groups were irradiated on Mondays, Wednesdays, and Fridays for 2 weeks. A fourth group of animals (no irradiation) served as diacylglycerol-treated controls. Mice in a fifth group (no irradiation) were euthanized 2 days after the last application of OAG. The topical application of OAG for 1 week, followed by a 2-week nontreatment period without UV irradiation, resulted in a sixfold increase in melanin deposition. The sixfold increase refers to increased melanin deposition over that noted after 1 week of OAG treatment only. Furthermore, the extent of melanin deposition was 50% of that noted after 2 weeks of UV irradiation alone.

When 1 week of OAG treatment was followed by 2 weeks of UV irradiation, a 37-fold increase in melanin deposition (compared to results for OAG only) with no lag time was observed. This increase in melanin deposition was threefold higher when compared to the results for UV irradiation alone, indicating that OAG pretreatment acts synergistically with UV light exposure. It was determined that this synergistic effect with UV light was more pronounced when melanization was used as the end point.

In the fourth experiment, five groups of 10 mice were treated as follows over a period of 5 consecutive days (Monday through Friday): group 1 (vehicle), group 2 (0.18 μ mol OAG), group 3 (0.58 μ mol OAG), group 4 (1.75 μ mol OAG), and group 5 (1.75 μ mol DOG). All groups were irradiated on Mondays, Wednesdays, and Fridays for 2 weeks. The effect of OAG in combination with UV irradiation was comparable to that of DOG + UV irradiation. However, it is important to note that OAG treatment increased the dopa-positive area and melanin deposition more effectively than did DOG. Specifically, 0.58 μ mol of OAG induced the same effect as 1.75 μ mol of DOG.

The authors reported that the results suggest that a PKC-dependent process is involved in melanogenesis, because diacylglycerols are known to activate PKC (Agin et al. 1991).

Allan et al. (1995) studied the effects of 1-oleyl-2-acetyl-sn-glycerol (OAG) and its analogues [OAG (sn) and OAG (racemic)], 1,2 dioctanoyl-sn-glycerol (diC₈), and 1,2-dipalmitoyl-sn-glycerol (diC₁₆) on melanin production in vivo using four groups of outbred, pigmented guinea pigs (12 to 20 weeks old; 4 to 6/group). Test substances were applied to skin (1 cm²) that had been shaved and treated with a depilatory. Sites were irradiated using two Sylvania FS40UVB bulbs. The irradiance (2.6 \times 10⁻⁴ W/cm²) was measured through a quartz filter using an IL700A research radiometer fitted with a UVB probe (filter UVB, 290 \pm 5 nm). The animals were exposed daily for ten days to 70 mJ/cm² for 3 min (12 s equals 0.6 minimal erythemal dose).

Animals in group 1 were treated with the vehicle (propylene glycol) alone and four separate concentrations of OAG (20, 30, 40, and 60 mg/ml) once daily for 5 days. Group 2 animals were treated with vehicle only and five separate concentrations of OAG (10, 20, 30, 40, and 60 mg/ml) either once or twice daily for 5 days. Animals in group 3 were treated with 50 mg/ml OAG(sn), OAG(rac), diC₈, diC₁₆, and vehicle alone twice daily for 5 days. In group 4, animals were treated with OAG (25

mg/ml) three times daily at one site and with the estimated 0.6 minimal erythemal dose daily for 10 days at a second site. Sites were evaluated weekly according to the following grading scale: 0 (no change from baseline color) to +4 (profound even darkening). Punch biopsies were obtained from test and control application sites and specimens examined microscopically. For statistical analysis of the data, analysis of variance (ANOVA) was performed.

In group 1, erythema and scale were observed in most, but not all, sites treated with OAG. However, pigmentation developed at all test sites, and was classified as dose dependent (p < 0.001). The following results are based on a computer-image analysis that was performed on sections obtained from untreated, vehicle-treated, and sites treated with 60 mg/ml OAG. Compared to untreated or vehicle control skin sites, a significant increase (280% increase; p < 0.00001) in melanin content was reported for OAG-treated skin sites. The difference in melanin content between untreated and vehicle-treated skin was not significant.

Cutaneous reactions in group 2 were similar to those observed in group 1, with the exception that the 10 mg/ml dose of OAG induced less inflammation. Mild erythema was observed in one of the four animals dosed with 10 mg/ml OAG. Additionally, the degree of pigmentation induced by OAG in group 2 was consistent with the dose-response pattern (p < 0.02) that was noted in group 1. The following results are based on a computerimage analysis of sections from biopsies obtained from each test site in a representative animal. A linear relationship between the concentration of OAG applied and the percent epidermal area covered by melanin was noted. These results indicated a dose-dependent increase in pigmentation (p < 0.00001). Additionally, compared to the vehicle control, 60 mg/ml OAG induced a 250% increase in epidermal melanin content.

In group 3, mild erythema was observed at some of the sites treated with OAG. Erythema was not observed at vehicle control sites or sites treated with $\mathrm{diC_{16}}$. On days 19 to 24, peak pigmentation (scores of +1 to +2) was noted in all five animals at sites treated with OAG(sn), OAG (racemic), and $\mathrm{diC_{8}}$. The increase in skin pigmentation at these sites was statistically significant (p < 0.005), compared to the complete lack of increased pigmentation at sites treated with vehicle or $\mathrm{diC_{16}}$. The following results are based on a computer-image analysis of sections from biopsies. Compared to the control, OAG(sn), OAG(rac), and $\mathrm{diC_{8}}$ induced a significant increase in pigmentation (p < 0.0005). $\mathrm{DiC_{16}}$ did not induce a significant change in pigmentation.

In group 4, even pigmentation (scores of +2 to +3) was observed in all six animals at sites treated with OAG as well as those treated with UVB, but not at vehicle-treated sites. The results of an image analysis of biopsy cross sections from a representative animal indicated a 367% increase in melanin content at the OAG-treated site (compared to vehicle control) and a 397% increase in melanin content at the UVB-irradiated site (compared to untreated control). These results indicated that simulation of

melanin production by OAG treatment and UVB irradiation was comparable.

The authors stated that results indicate that topically applied diacylglycerols can induce an increase in epidermal pigmentation, presumably via PKC activation. The increase in epidermal pigmentation in this study was long-lasting, and, both clinically and histologically, was comparable to UV-induced tanning. The effects of OAG treatment on skin histology were also evaluated in this study. Except for the presence of acanthosis, hematoxylin and eosin-stained sections of OAG-treated skin at 19 to 25 days after the final application were similar to vehicle or untreated control sections (Allan et al. 1995).

Antimicrobial Activity

Glyceryl Dilaurate

No antimicrobial activity was observed in a study in which various bacterial strains (*Streptococcus pyogenes* included) were incubated with Glyceryl Dilaurate (Conley and Kabara 1973).

Kabara et al. (1977) observed that Glyceryl Dilaurate had no effect on the antimicrobial activity of Glyceryl Laurate in the bacterial strain *Streptococcus pyogenes*. In this assay, bacterial cultures were treated with 5 μ g/ml Glyceryl Laurate + 5 μ g/ml Glyceryl Dilaurate, 10 μ g/ml Glyceryl Laurate + 10 μ g/ml Glyceryl Dilaurate, 5 μ g/ml Glyceryl Laurate, and 10 μ g/ml Glyceryl Dilaurate, respectively. Bacterial growth was monitored by changes in the optical density of the medium at 550 nm.

ANIMAL TOXICOLOGY

Acute Oral Toxicity

Glyceryl Dilaurate

MB Research Laboratories, Inc. (1991a) evaluated the acute oral toxicity of Glyceryl Dilaurate using ten male Wistar Albino rats (weights 211–279 g). The test substance was melted in a water bath and administered orally (single dose of 5 g/kg) to each animal using a syringe and dosing needle. The animals were observed at 3 to 4 h post dosing and then daily for 14 days. Necropsy was not performed. None of the animals died, and all were in good health throughout the study. It was concluded that the LD_{50} was greater than 5.0 g/kg.

Glyceryl Dibehenate

The Institut Français de Recherches et Essais Biologiques (1980) evaluated a trade mixture (Compritol 888 ATO) containing an unspecified concentration of Glyceryl Dibehenate in an acute oral toxicity study using five female and five male OFA SPF rats (Sprague-Dawley originated; body weights = 163 g). The mixture was diluted with hydroxycellulose gel to a concentration of 25%, and then administered to each animal by esophageal intubation. Each animal received a single dose of 5 g/kg (dose volume = 20 ml/kg). Dosing was followed by a 14-day observation period. No deaths were reported.

Glyceryl Stearate Citrate

An acute oral LD_{50} of >2 g/kg has been reported for Glyceryl Stearate Citrate. Neither the species of animals tested nor details concerning the test protocol were included (Huls America, Inc., no date).

Acute Dermal Toxicity

Glyceryl Stearate Citrate

An acute dermal LD_{50} of >2 g/kg has been reported for Glyceryl Stearate Citrate. Neither the species of animals tested nor details concerning the test protocol were included (Huls America, Inc., no date).

Chronic Oral Toxicity

Glyceryl Dilaurate

In a study by Fitzhugh et al. (1960), 24 albino rats (males and females, Osborne-Mendel strain) were fed a diet consisting of Trilaurin (8%), Glyceryl Dilaurate (45%), and Glyceryl Laurate (40% to 45%) at a concentration of 25% in the diet for 2 years. Thus, the individual glyceryl esters were fed at effective dietary concentrations of \approx 2% (Trilaurin), \approx 11.25% (Glyceryl Dilaurate), and \approx 10% to 11.25% (Glyceryl Laurate). The 24 control rats were fed hydrogenated cottonseed oil at a concentration of 25% in the diet.

After 26 or 52 weeks of feeding, no significant difference in weight gain between test and control animals was noted. However, due to the higher caloric intake, weight gain was increased in the test group. Additionally, no significant difference in the total number of deaths between test and control groups was noted at the end of the 2-year study. No gross pathology attributable to feeding of the mixture was observed at necropsy. The only effect that was attributable to feeding, slight excess of hepatic cell fatty change (compared to controls), was noted at microscopic examination. A lesser and questionably significant degree of intrahepatic bile duct proliferation was also noted (Fitzhugh et al. 1960).

Ocular Irritation

Glyceryl Dilaurate

MB Research Laboratories, Inc. (1991b) evaluated the ocular irritation potential of Glyceryl Dilaurate using six New Zealand albino rabbits. The undiluted test substance (0.1 ml) was instilled into the conjunctival sac of one eye of each animal. The eye lids were held together briefly after instillation. Untreated eyes served as controls. Reactions, recorded on days 1, 2, and 3, were scored according to the Draize scale (0 to 110). Slight conjunctival irritation was observed in two of the six treated eyes, and had cleared by day 2. Glyceryl Dilaurate was non-irritating to the eyes of rabbits.

Glyceryl Dibehenate

Pharmakon Europe (1995) evaluated a trade mixture (Compritol 888 ATO) containing an unspecified concentration of

Glyceryl Dibehenate in an ocular irritation study using three male New Zealand white rabbits. A 5% (w/w) suspension of the mixture in kernel oil (0.1 ml) was instilled into the inferior conjunctival sac of the right eye of each animal. After instillation, the upper and lower eyelids were held together for approximately 10 s to prevent loss of the test substance; eyes were not rinsed. The conjunctiva, iris, and cornea were examined at 1 h and 1, 2, and 3 days post instillation. Ocular lesions were evaluated according to the Draize scale and mean ocular irritation indices calculated for each examination period. The maximum ocular irritation index (MaOI) was the highest of the four mean ocular irritation indices recorded. Ocular lesions were observed at 1 h, but had cleared by day 3. Compritol 888 ATO was classified as a slight ocular irritant (MaOI = 8.67, at 1 h post instillation).

Glyceryl Stearate Citrate

Glyceryl Stearate Citrate did not induce ocular irritation in the Draize test. Details concerning the test protocol and study results were not included (Huls America, Inc., no date).

Skin Irritation

Glyceryl Dilaurate

Product Safety Labs (1987) evaluated the skin irritation potential of undiluted Glyceryl Dilaurate using six male New Zealand albino rabbits. The test substance (0.5 ml) was applied to one abraded site and one intact site (2-cm² area, clipped free of hair) within an area encompassing dorsal and ventral surfaces from the scapular to pelvic area. The sites were covered with an occlusive patch, and the entire trunk (patches included) was wrapped with an elastic cloth. At 24 h post application, the patches were removed and sites wiped to remove any residual test material. Reactions were scored according to the following scales: 0 (no erythema) to 4 (severe erythema to slight eschar formation) and 0 (no edema) to 4 (severe edema, raised more than 1 mm and extending beyond the area of exposure).

Well-defined erythema was observed at all test sites (24 and 72 h post application, abraded and intact skin), except for one intact site in which the reaction diminished to barely perceptible erythema at 72 h. The test substance was classified as a mild primary irritant with an average primary irritation score = 2 (Product Safety Labs 1987).

In another study, MB Research Laboratories, Inc. (1991c) evaluated the skin irritation potential of Glyceryl Dilaurate using six New Zealand Albino rabbits. The test substance was melted in a water bath and then applied (0.5 ml) to an abraded site and an intact site on the back of each animal. Test sites were covered with occlusive patches, secured with adhesive tape and wrapped with plastic, for 24 h. After patch removal, reactions were scored according to the following scales: 0 (no erythema) to 4 (severe erythema to slight eschar formation) and 0 (no edema) to 4 (severe edema, raised more than 1 mm and extending beyond the area of exposure). The primary irritation index (PII) was calculated by adding the mean values (six rabbits) for ery-

thema/eschar and edema on intact and abraded skin at 24 and 72 h (total of eight values) and dividing the sum by 4. Glyceryl Dilaurate was classified as a nonirritant (PII = 0).

Glyceryl Stearate Citrate

Glyceryl Stearate Citrate induced slight skin irritation in the Draize test. Details concerning the test protocol and study results were not included (Huls America, Inc., no date).

Skin Sensitization

Glyceryl Stearate Citrate

Safepharm Laboratories (1981) evaluated the skin sensitization potential of Glyceryl Stearate Citrate in a modification of the Magnusson-Kligman guinea pig sensitization test using 10 female albino guinea pigs of the Dunkin-Hartley strain. The test methodology involved a two-stage induction procedure followed by a topical challenge 2 weeks later. The test concentrations of Glyceryl Stearate were as follows: intradermal induction (5% in liquid paraffin), topical induction (25% in petroleum jelly), and topical challenge (25% in petroleum jelly). During the first phase of induction (day 0), three pairs of intradermal injections were made simultaneously to skin (in shoulder region) that had been clipped free of hair as follows: Freund's complete adjuvant (0.1 ml), 5% Glyceryl Stearate Citrate (0.1 ml), and 5% Glyceryl Stearate emulsified in Freund's complete adjuvant.

The second phase of induction (day 7) involved the topical application of 25% Glyceryl Stearate Citrate to the injection site (at 1 week post injection) using a patch made of filter paper. The patch was secured with an elastic adhesive bandage for 48 h. During the challenge phase (day 21), 25% Glyceryl Stearate Citrate was applied (under occlusive patch) to shaved skin of the right flank. Petroleum jelly (vehicle) was applied to shaved skin of the left flank according to the same procedure. Patches were secured with an elastic adhesive bandage, and another bandage of the same type was wrapped around the trunk of the animal. At 24 h post application, patches were removed. Challenge reactions were scored at 24 and 48 h according to the following scale: 0 (no reaction) to 3 (intense redness and swelling). An additional five guinea pigs served as the controls, and they were exposed to the test substance vehicles according to the same induction procedure indicated for test animals. The challenge phase (same procedure) consisted of the application of 25% Glyceryl Stearate to the flank.

Sensitization was not induced in any of the animals tested (Safepharm Laboratories 1981).

Glyceryl Diisostearate

The Shiseido Research Center (1977) evaluated the contact allergenicity of Glyceryl Diisostearate in a maximization test using nine Hartley guinea pigs. On day 1 of the induction phase, the animals were injected intradermally (shaved nuchal region) with the following: 0.1 ml of emulsified Freund's complete adjuvant (E-FCA), from equal volumes of FCA and distilled water; 0.1

ml of 25% Glyceryl Diisostearate; and 0.1 ml of 50% Glyceryl Diisostearate emulsified with FCA. At week 1 post injection, 50 mg of 10% sodium lauryl sulfate (in petrolatum) was applied to the nuchal region. On the following day, 0.2 ml of 100% Glyceryl Diisostearate was applied occlusively for 48 h. The five control guinea pigs were injected intradermally with distilled water (intradermal injection phase) and then treated with "four points" of 0.1 ml of E-FCA at the same time that test animals received occlusive patch applications.

During the challenge phase (3 weeks after the first induction), Glyceryl Diisostearate concentrations of 5%, 20%, 50%, and 100% in ethanol (0.1 ml each) were applied topically to the flank (clipped free of hair) of test and control animals. Reactions were scored at 24 and 48 h post application according to the following scale: 0 (no reaction) to 4 (severe erythema [beet redness] to slight eschar formation (injuries in depth) and severe edema [raised more than 1 mm and extending beyond area of exposure).

Glyceryl Diisostearate did not induce sensitization at any of the challenge concentrations tested (Shiseido Research Center 1977).

PHOTOTOXICITY/PHOTOALLERGENICITY

Data on the phototoxicity/photoallergenicity of glyceryl diesters were not available. As noted earlier, however, glyceryl triesters and glyceryl monoesters do contain glyceryl diesters. Therefore, data on the phototoxicity/photoallergenicity of a glyceryl triester, Triisostearin, and a glyceryl monoester, Glyceryl Isostearate, are included in this review.

Triisostearin

Unichema International (1997) evaluated the phototoxicity and photoallergenicity potential of Triisostearin using 20 albino guinea pigs. The back and sides of each animal were divided into the following six treatment areas: test material plus UVA; test material plus UVB; test material alone; positive control (8-methoxypsoralen) plus UVA; UVB alone; and UVA alone.

Doses of the test material and positive control (dose for each = 0.02 ml/cm²) were applied 30 min prior to irradiation. UV irradiations were performed using Philips tubes (TL 20W/09 for UVA and TL 20W/12 UV for UVB). Cutaneous reactions were evaluated at 24 h post treatment.

With or without UV irradiation, Triisostearin did not induce significant cutaneous reactions. The positive control (8-methoxypsoralen) induced major reactions.

Glyceryl Isostearate

Unichema International (1997) evaluated the phototoxicity and photoallergenicity potential of Glyceryl Isostearate using 20 albino guinea pigs. The back and sides of each animal were divided into the following six treatment areas: test material + UVA, test material + UVB, test material alone, positive control (8-methoxypsoralen) + UVA, UVB alone, and UVA alone. Doses of the test material and positive control (dose for

each = 0.02 ml/cm²) were applied 30 min prior to irradiation. UV irradiations were performed using Philips tubes (TL 20W/09 for UVA and TL 20W/12 UV for UVB). Cutaneous reactions were evaluated at 24 h post treatment. Glyceryl Isostearate did not induce significant cutaneous reactions with or without UV irradiation. The positive control (8-methoxypsoralen) induced severe reactions.

EFFECT ON DNA SYNTHESIS

Glyceryl Dilaurate and Glyceryl Distearate

CTFA (2004) provided a report in which the effect of two commercial glyceryl diesters (Glyceryl Dilaurate [tradename: Emulsynt GDL] and Glyceryl Distearate [tradename: Stepan GDS]) and two other diacylglycerols (1-oleoyl-2-acetoyl-sn-glycerol [OAG] and 1,2-dipalmitoyl-sn-glycerol [DPG]) on DNA synthesis was studied using normal human dermal fibroblasts from neonatal foreskin. The fibroblasts were treated at approximately 50% confluence in Dulbecco/Vogt modified Eagle's minimal essential medium (DMEM) with 1% bovine calf serum (BCS) and 1% penicillin-streptomycin.

Each of the four diacylglycerols was tested at doses of 0.1, 1.0, and 10.0 μ g/ml during a 24-h incubation period. The controls were as follows: 1% BCS, 10% BCS, 30 μ M adenosine monophosphate (AMP; positive control), and ethanol (vehicle control). [3 H]Thymidine incorporation was used to measure DNA synthesis. Because of the extreme increase in [3 H]thymidine incorporation that was induced by AMP, statistical comparisons involved results for the four test substances versus those for the ethanol vehicle and 1% BCS without the AMP positive control.

No treatment-related changes in fibroblast proliferation (as determined by DNA synthesis) were induced by Glyceryl Dilaurate, Glyceryl Distearate, OAG, or DPG. Compared to the ethanol vehicle, the % changes in DNA synthesis were as follows: 19.1% (0.1 μ g/ml Glyceryl Distearate). 20.0% (1.0 μ g/ml Glyceryl Distearate), 4.5% (10.0 μ g/ml Glyceryl Distearate), 16.7% (0.1 μ g/ml Glyceryl Dilaurate), 12.9% (1.0 μ g/ml Glyceryl Dilaurate), and 3.2% (10.0 μ g/ml Glyceryl Dilaurate).

It was concluded that the two commercial glyceryl diesters, Glyceryl Dilaurate and Glyceryl Distearate, and the two diacylglycerols, OAG and DPG, did not alter cell proliferation in normal human dermal fibroblasts in vitro (CTFA 2004).

GENOTOXICITY

Diacylglycerol Oil

Kasamatsu et al. (2005) evaluated the genotoxicity of Diacylglycerol Oil (edible oil enriched in Diacylglycerol [more than 80%]) using the bacterial reverse mutation assay (Ames test), chromosomal aberration assay (Chinese hamster lung cells [CHL/IU]), and a bone marrow micronucleus assay (ICR CD mice). Diacylglycerol Oil was synthesized by esterifying glycerol with fatty acids obtained from soybean and rapeseed oil

using a reverse reaction of immobilized lipase. Heated Diacylglycerol Oil was also evaluated in these tests; the heating conditions (deep-frying) caused excessive deterioration of the oil.

In the Ames test, Diacylglycerol Oil (heated or unheated, in DMSO) was tested at concentrations up to $5000 \,\mu g$ /plate using the following bacterial strains with and without metabolic activation: *Salmonella typhimurium* strains TA98, TA100, TA1535, and TA1537, and *Escherichia coli* strain WP2*uvrA*. DMSO served as the vehicle control, and the positive controls were as follows: 2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide (AF-2), 2-aminoanthracene (2AA), and sodium azide (NaN₃).

With heated or unheated Diacylglycerol Oil, no bacterial toxicity was observed over the range of doses tested. Compared to the vehicle control, both unheated and heated Diacylglycerol Oil did not cause more than a twofold increase in the mean number of revertants per plate in any bacterial strain. The positive controls for each strain induced the expected increase in the number of revertants per plate, indicating that the test was valid. The authors concluded that unheated Diacylglycerol Oil and heated Diacylglycerol Oil showed no evidence of gene mutagenic potential in the presence or absence of metabolic activation.

In the chromosomal aberration test, unheated or heated Diacylglycerol Oil (suspended in a 0.5% carboxymethylcellulose sodium aqueous solution (0.5% CMC-Na) was incubated with CHL/IU cells for 6 h (short-term test) and 24 or 48 h (continuous treatment) either with or without metabolic activation. Diacylglycerol Oil (heated or unheated) was tested at doses up to 5000 μ g/ml. Colcemid was added to the medium at 2 hours prior to harvesting, and 100 metaphases per slide (200 metaphases per dose) were analyzed.

In both short-term and continuous treatment tests with heated or unheated Diacylglycerol Oil, the frequency of cells with structural or numerical aberrations was <5% (negative results) both with and without metabolic activation. Neither heated nor unheated Diacylglycerol was cytotoxic. Each positive control, mitomycin and benzo(a)pyrene caused an increase in the frequency of cells with >10% chromosomal aberration. The authors concluded that neither unheated nor heated Diacylglycerol were clastogenic.

In the micronucleus assay, either heated or unheated Diacylglycerol Oil (in olive oil) was administered at doses of 500, 1000, and 2000 mg/kg, via oral gavage, to three groups of 8-week-old, male ICR (Crj: CD-1) mice, respectively. Each animal (in each dose group) received two oral doses, separated by 24 h. The negative control group received olive oil, and positive control mice received a single intraperitoneal (i.p.) injection of cyclophosphamide (in water; dose = 40 mg/kg). Bone marrow smears were prepared at 24 h post administration of the last dose. The number of micronucleated polychromatic erythrocytes (MNPCEs) in 2000 PCEs was counted. A response was classified as positive if the test substance caused a significant, dose-dependent increase in the total number of MNPCEs. The PCE/NCE (nor-

mochromatic erythrocyte) ratio was determined by examining 1000 erythrocytes from each mouse.

A significant increase in the incidence of MNPCEs was not observed in any of the groups (3 per test substance) dosed with unheated or heated Diacylglycerol Oil. However, regarding the PCE/NCE ratio, a significant difference (p < 0.05) between the negative control group and the 2000 mg/kg unheated Diacylglycerol Oil dose group, but not the two lower dose groups, was noted.

The authors were unsure as to whether this finding was indicative of a toxic effect of unheated Diacylglycerol Oil, but noted that the finding resulted mainly from intraspecific variability (i.e., ratio of 39.3 reported for one mouse, compared to the group average value of 49.7 ± 5.5). For heated Diacylglycerol Oil, a significant difference in the PCE/NCE ratio (p < 0.05) was observed between the negative control group and the $1000 \, \mathrm{mg/kg}$ dose group, but not the other two dose groups. This finding was not considered to be a toxic action of heated Diacylglycerol Oil, because the effect was not dose-dependent and the value of the ratio was only marginally significant. The positive control, cyclophosphamide, induced a statistically significant (p < 0.01) increase in the incidence of MNPCEs.

Overall, the authors concluded that unheated Diacylglycerol Oil and heated Diacylglycerol Oil did not show any evidence of genotoxic potential (Kasamatsu et al. 2005).

CARCINOGENICITY AND TUMOR PROMOTION

Glyceryl Distearate

In a study reported by Setala et al. (1961), the character of ultrastructural changes in mouse epidermis caused by Glyceryl Distearate was evaluated with and without initiation. Randomly bred mice of a tumor-resistant strain were used. Glyceryl Distearate (0.0066 M in acetone) was administered six times per week (Sundays excluded). The test substance was dropped onto the skin and the theoretical single dose of Glyceryl Distearate at the cutaneous surface was 0.16 mg. In initiation experiments, 9,10-dimethyl-1,2-benzanthracene (DMBA) in light paraffin was used. No controls using acetone alone were performed. The solution was carefully dropped onto the center of the treatment area on the back and spread with a glass rod. Posttreatment with Glyceryl Distearate was started after 30 days. Hair within the treatment area was cut at 1-week intervals. In experiments without initiation, skin biopsies were obtained from five animals on days 2, 6, 10, 16, 30, and 60 after the beginning of the study. In experiments with initiation, the first skin biopsy was taken on day 30 and 2 days after initiation.

In the absence of initiation, Glyceryl Distearate (0.0066 M in acetone) induced a moderate hyperplastic response. The stratum corneum was thicker and coarser than normal (biopsy taken on day 30). Glyceryl Distearate did not cause cellular or nuclear atypia, and the pilosebaceous apparatus was intact. Epidermal alterations were accompanied by inflammation of the dermis. With initiation, there was an increase in the total cell count.

Ultrastructurally, Glyceryl Distearate, in the absence of initiation, induced hyperplasia of the interfollicular epidermis (IFE). In the presence of initiation, few and sparse alterations were observed in the early stages of the treatment period. Later, a definite hyperplasia of the interfollicular epidermis was noted. Whether or not Glyceryl Distearate increased the DMBA tumor incidence was not stated (Setala et al. 1961).

Glyceryl Dioleate

Semba and Inui (1991) evaluated the inhibitory effects of free radical scavengers (superoxide dismutase, catalase, and mannitol) on diacylglycerol-induced transformation. Glyceryl Dioleate promoted transformation in 3-methylcholanthrene (3-MC)-initiated BALB/3T3 A31-1-1 cloned cells in vitro. In cultures treated with Glyceryl Dioleate (10 μ g/ml), the mean number of transformed foci per dish (25 dishes total) was 1.0 \pm 0.2. In control cultures treated with DMSO (control), the mean number of transformed foci per dish (60 dishes total) was 0. The mean number of transformed foci per dish (40 dishes total) in 3-MC (0.5 μ g/ml)-treated cultures was 0.2 \pm 0.1. Superoxide dismutase, catalase, and mannitol had significant inhibitory effects (p < 0.01) on Glyceryl Dioleate-induced transformation. The results of this study suggest that activation of PKC alone is insufficient for transformation, and that the generation of reactive oxygen accompanied by activation of the enzyme is essential to the promotion process in BALB/3T3 cells.

Glyceryl Stearate

As noted earlier glyceryl triesters and glyceryl monoesters do contain glyceryl diesters. Therefore, data on the phototoxicity/photoallergenicity of a glyceryl monoester, Glyceryl Stearate, are included in this review.

Saffioti and Shubik (1963) tested the tumor promoting activity of Glyceryl Stearate on the clipped dorsal skin of Swiss mice. One week after a single application of 9,10-dimethylbenz(a)anthracene (DMBA) (1% to 1.5% in mineral oil), 5% Glyceryl Stearate (in acetone) was applied to the skin twice weekly. No tumors developed; slight epidermal hyperplasia at the site of application was noted.

Other Diacylglycerols

Smart et al. (1986) reported that sn-1,2-didecanoylglycerol (2.6 μ mol), sn-1,2-dioctanoylglycerol (2.6 μ M), and sn-1,2-dioleylglycerol (2.6 μ mol) stimulated epidermal DNA synthesis after topical application to CD-1 mice. However, 1,3-didecanoylglycerol (2.6 μ mol) had little or no effect. The increased epidermal DNA synthesis induced by sn-1,2-dioleoylglycerol or TPA was inhibited when either was administered simultaneously with fluocinolone acetonide, a potent inhibitor of tumor promotion.

In a later study, Smart et al. (1988) reported that the topical application of 2.5 μ M sn-1,2-didecanoylglycerol to female CD-1 mice and a tumor-promoting dose of 1 nmol TPA induced ornithine decarboxylase activity to the same extent. However,

topical application of 2.5 μ M sn-1,2-didecanoylglycerol did not cause morphological changes in the epidermis following a single application or repeated applications twice a week for 4 weeks. Topical application of 2.5 μ M sn-1,2-didecanoylglycerol also did not function as a complete tumor promoter when applied twice weekly for 28 weeks. More frequent applications of sn-1,2-didecanoylglycerol (5 μ mol twice a day for 5 days) induced epidermal hyperplasia that was comparable to that of 5 nmol TPA twice weekly for 4 weeks.

Smart et al. (1989) demonstrated that, after frequent, repetitive treatment, the synthetic lipid second messenger, sn-1,2-didecanoylglycerol is a complete tumor promoter in DMBA-initiated mouse skin. In this study, an application frequency and dose of sn-1,2-didecanoylglycerol that induced epidermal hyperplasia were used. One week after treatment with 200 nmol DMBA, the mice (CD1 females) were dosed with 2 or 5 μ mol sn-1,2-didecanoylglycerol for 5 days per week over a period of 20 or 24 weeks. sn-1,2-Didecanoylglycerol functioned as a complete tumor promoter in a dose-dependent manner. It was stated that it is likely that rapid metabolism of sn-1,2-diacylglycerols necessitates frequent application for their effectiveness as complete tumor promoters.

Mills et al. (1993), using transgenic TG.AC mice, found that diacylglycerols are implicated in the clonal expansion of mutated Ha-ras—containing cells. The authors concluded that these data indicate that an increase in endogenous Diacylglycerol could contribute to the clonal expansion of cells containing Ha-ras oncogene.

Smart (2001) noted, in studies on the tumor promoting activity of 1,2-diacylglycerols, that the structure of the particular diacylglycerol, its dose, and the exposure regimen by which the dose is delivered all play a role in tumor promotion. In particular, only a specific 1,2-diacylglycerol configuration was active and that the other configuration (also known as 2,3-diacylglycerol) was not active in tumor promotion.

Findings relating to 1,2-diacylglycerol-induced activation of PKC are summarized as follows: (1) If the fatty acid in the 1 or 2 position is unsaturated, then the diacylglycerol can activate PKC. 1,2-Diacylglycerols with two saturated fatty acids are less effective (Mori et al. 1982). (2) The stereospecificity of diacylglycerols is critical in terms of their biological activity. 1,2-sn-Diacylglycerols, but not 2,3-sn-diacylglycerols, activate PKC (Nomura et al. 1986). (3) The activities of different diacylglycerols (i.e., activation of PKC) have been compared. The most important observation was that the activity of saturated 1,2-diacylglycerols was lost when the fatty acid moiety in the structure was a long-chain fatty acid. This effect was not only observed in in vitro kinase assays, but also in a cellular model (platelets). When the length of the acyl chain was greater than 14 carbons, the activity was very low or absent. However, the issue of unsaturated 1,2-diacylglycerols potently activating PKC, even if the acyl chain is long, was raised by Go et al. (1987).

Given the relationship between chain length/structure of fatty acid moiety and PKC activation, Table 4 contains a list of the

TABLE 4
Diacylglycerol fatty acid moieties for cosmetic ingredients.

Ingredient	Fatty acid (FA) moiety saturation	Fatty acid chain length
Glyceryl Dilaurate	No unsaturated FA	12 carbons @
Glyceryl Diarachidate	No unsaturated FA	20 carbons @
Glyceryl Dibehenate	No unsaturated FA	22 carbons @
Glyceryl Dierucate	2 unsaturated FA	22 carbons @
Glyceryl Dihydroxystearate	No unsaturated FA	18 carbons @
Glyceryl Diisopalmitate	No unsaturated FA	16 carbons @
Glyceryl Diisostearate	No unsaturated FA	18 carbons @
Glyceryl Dilinoleate	2 unsaturated FA	18 carbons @
Glyceryl Dimyristate	No unsaturated FA	14 carbons @
Glyceryl Dioleate	2 unsaturated FA	18 carbons @
Glyceryl Diricinoleate	2 unsaturated FA	18 carbons @
Glyceryl Dipalmitate	No unsaturated FA	16 carbons @
Glyceryl Dipalmitoleate	2 unsaturated FA	16 carbons @
Glyceryl Palmitate Lactate	No unsaturated FA	16 carbons (palmitic) 3 carbons (lactic)
Glyceryl Distearate	No unsaturated FA	18 carbons @
Glyceryl Stearate Citrate	No unsaturated FA	18 carbons (stearic) 5 carbons (citric)
Glyceryl Stearate Lactate	No unsaturated FA	18 carbons (stearic) 3 carbons (lactic)
Glyceryl Stearate Succinate	No unsaturated FA	18 carbons (stearic) 4 carbons (succinic)

1,3-diacylglycerols included in this safety assessment, along with a description of the fatty acid moiety (unsaturated or not) and its chain length.

Diacylglycerol Oil

Soni et al. (2001) evaluated the chronic toxicity of diacylglycerol oil in a 2-year feeding study using four groups of 60, 4-week-old male and female SPF rats of the Crj:CD(SD) strain. A randomized diacylglycerol, manufactured through the esterification of fatty acids, is the main constituent of diacylglycerol oil. The primary components of this oil are 1,3- and 1,2-diacylglycerol (ratio of 7:3). Other components include 20% triacylglycerol and 5% monoacylglycerol.

The four groups used in the study were defined as follows:

- low-dose group receiving 2.65% diacylglycerol oil plus 2.65% edible oil composed of rapeseed oil, corn oil, high linoleic safflower oil, and high oleic safflower oil (a fatty acid composition equivalent to diacylglycerol oil). Dose estimated at 0.89 g/kg body weight/day—males and 1.18 g/kg body weight/day—females;
- high-dose group receiving 5.3% diacylglycerol oil.
 Dose estimated at 1.77 g/kg body weight/ day—males and 2.35 g/kg body weight/ day—females;
- control group 1 receiving 5.3% edible oil composed of rapeseed, corn, high linoleic safflower, and high oleic safflower oils; and
- control group 2 receiving 5.3% edible oil composed of rapeseed and soybean oils only.

Animals were observed for general signs and mortality throughout the study. At weeks 30 and 77, 10 rats per sex from each group were anesthetized i.p. with sodium phenobarbital, and blood was collected from the abdominal aorta for clinical hematological and chemical analysis; urinalyses were also performed on 10 rats per sex from each group. The remaining animals were killed at the end of week 105 and necropsied.

Mortalities for the four groups were as follows: low-dose group (54 rats: 24 males, 30 females), high-dose group (58 rats: 29 males, 29 females), control 1 (50 rats: 28 males, 22 females), and control 2 (58 rats: 27 males, 31 females).

Histopathological examination was performed on a variety of tissues from animals that were killed at the end of 30 and 77 weeks of feeding. For rats killed after 105 weeks of feeding, only the mammary gland was examined microscopically.

No statistically significant differences in cumulative survival rate or treatment-related clinical signs of toxicity were observed in animals killed at 30, 77, or 105 weeks.

A statistically significant difference in body weight (limited to days 148 to 162) occurred between low dose males and control group 2. Feed consumption also differed significantly from controls, on an intermittent basis, in high- and low-dose males. The authors did not consider these findings to be toxicologically significant.

Hematology, blood chemical analysis, or urinalysis findings, for rats in low- or high-dose groups indicated no consistent pattern that was indicative of a treatment-related effect. Statistically significant changes in several parameters were reported. For example, compared to control group 2, activated partial thromboplastin time (APTT) in high-dose male rats, dosed for 30 weeks,

was significantly prolonged. Additionally, compared to control groups 1 and 2, the platelet count in high- and low-dose females, dosed for 77 weeks, was significantly decreased. The authors considered these differences to be incidental because they were either limited to one sex or one collection interval and lacked a dose response, or were not supported by any other changes in related clinical parameters.

At gross examination, changes in organ weight were found to be significant when results for test and control animals were compared after 30 weeks of feeding. Compared to control group 2, absolute weights of the thyroid, heart, kidney, and liver and relative liver weight in the low-dose males were significantly lower. A comparison of control groups 1 and 2 with high-dose males revealed no significant differences in organ weight. No significant changes in weight of these organs (heart, thyroid, kidney, liver) were noted after 77 weeks of feeding. None of the observed changes in the weight of these organs were considered treatment-related by the authors.

There were statistically significant differences in organ weights in high-dose males and females or low-dose females after 77 weeks of feeding (e.g., absolute and relative brain weight, relative spleen weight in males, and absolute and relative heart weights in females) and at 105 weeks (absolute thyroid weight in high-dose males). However, these differences were not considered toxicologically significant or treatment related, in that they were either limited to one sex, one collection period, or were not associated with a dose-response.

Compared to controls, neither gross nor microscopic findings indicated an increase in the incidence of non-neoplastic changes in rats that received high or low doses of diacylglycerol oil. The incidence of either benign or malignant epithelial mammary gland tumors was not different from that noted in the control group. Benign mammary gland tumors in dead female rats of the high-dose group were more frequent. However, it was concluded that the increased incidence was not related to feeding with diacylglycerol oil for the following reasons: (1) no dose-related increase in the incidence of mammary tumors was observed; (2) compared to control group 1 or the low-dose exposure group, the mammary tumor incidence in the high-dose group was not significantly increased; (3) the number of rats with benign tumors in the high-dose group was within the range of historical incidence reported in the published literature.

The only statistically significant differences (compared to control group 2, but not control group 1) among the groups tested were the increased incidence of adenocarcinoma in rats that died early, and the significantly higher total incidence (compared to control group 2, but not control group 1) of benign and malignant tumors combined for high-dose rats that died early and those killed at 105 weeks. The neoplastic findings were not dose related.

Overall, the authors concluded that no treatment-related effects of diacylglycerol oil were observed in rats at dietary concentrations up to 5.3% in the diet over a 2-year period (Soni et al. 2001).

REPRODUCTIVE AND DEVELOPMENTAL TOXICITY

Studies on the reproductive and developmental toxicity of glyceryl diesters were not found in the published literature.

CLINICAL ASSESSMENT OF SAFETY

Skin Irritation

Glyceryl Dilaurate

CTFA (1977) reported that the skin irritation potential of an eye shadow containing 1.5% Glyceryl Dilaurate was evaluated in a single-insult (24 h) occlusive patch test using 20 subjects (ages not stated). The control group consisted of 20 subjects (ages not stated) tested with an eye shadow that did not contain Glyceryl Dilaurate. Volatiles on the patch were allowed to evaporate over a period of $\approx\!15$ to 30 min, and each patch was hydrated prior to application. Reactions were scored according to the following scale: 0 (no evidence of any effect) to 4 (severe: deep red erythema with vesiculation or weeping with or without edema). Skin irritation was not observed in the test or control group.

CTFA (1999b) reported the skin irritation potential of a foundation containing 1.5% Glyceryl Dilaurate evaluated using 20 subjects (ages not stated) according to the procedure in the preceding study. The control group consisted of 20 subjects (ages not stated) tested with a foundation that did not contain Glyceryl Dilaurate. Three of the 20 subjects tested with the Glyceryl Dilaurate foundation had a barely perceptible reaction (\pm , minimal faint uniform or spotty erythema). Reactions were not observed in the remaining 17 subjects in this group. The distribution of reactions in the control group was as follows: \pm reaction (4 subjects), 1 reaction (2 subjects), 1+ reaction (1 subject), and 0 reaction (13 subjects). A 1 reaction (mild) was defined as pink, uniform erythema covering most of the contact site, and a 2 reaction (moderate) was defined as pink-red erythema, visibly uniform in the entire contact area. The 1+ reaction was classified as an intermediate reaction. Primary irritation indexes for the test and control foundation were 0.08 and 0.28, respectively.

Spring Grove Laboratories (2004) conducted a histological evaluation of skin from 5 of 30 female volunteers (18 to 56 years old) who had applied a prototype lotion or placebo formulation, both containing 0.5% Glyceryl Dilaurate, consecutively for 16 weeks (first phase: 30 subjects) or 21 weeks (second phase: 5 of the 30 subjects) in a within-subject, blinded study. The lotion also contained botanical extract, whereas the placebo formulation did not. The subjects were instructed to apply the lotion to one leg and the placebo formulation to the other at least once daily. The use of any other moisturizer during the study was prohibited.

In the first phase of the study (30 subjects), digital photographs of the application sites were taken at baseline, week 8, and week 16. The subjects also completed a self-assessment questionnaire. In the second phase (5 of the 30 subjects), the application sites were evaluated for erythema or scaling at week

21 according to the following scale: 0 (no evidence of any effect) to 4 (severe, defined as deep red erythema with or without vesiculation or weeping).

Skin irritation was not observed in any of the 30 subjects tested during the first phase of the study. Similarly, neither localized erythema nor scaling was observed in any of the five subjects tested during the second phase. Biopsies (2 mm) taken from both legs of the five subjects indicated no recognizable abnormalities of the skin. The epidermis was described as normal in thickness, and there was no evidence of scaling, inflammation, or neoplasms in any of the tissues that were evaluated (Spring Grove Laboratories 2004).

Glyceryl Dibehenate

The Institut D'espertise Clinique (1996) evaluated a trade mixture (Compritol 888 ATO) containing an unspecified concentration of Glyceryl Dibehenate in a skin irritation test using ten volunteers (eight females, two males). The test substance (10% in kernel stone oil; volume = 0.02 ml) was applied to the back under an occlusive patch (Finn Chamber on Scanpor) for a period of 48 h. Kernel stone oil, patch tested according to the same procedure, served as the negative control. At \approx 30 min post removal, reactions (erythema and edema) were scored according to the following scales: 0 (no erythema) to 4 (purpuric erythema) and 0 (no edema) to 3 (severe edema [1 mm thick at least] on a surface greater than the application area). Additional numerical scales were used for the evaluation of dryness and desquamation, papules, vesicles, bullae, pustules, detergent effect, and reflectivity.

Slight erythema (hardly visible) was observed in 2 of the 10 subjects tested. An observation described as "a slightly worn aspect of the skin" was reported for two additional subjects. Compritol 888 ATO (10%) did not induce pathological irritation or significant cutaneous intolerance, and, therefore, the investigators concluded that the test substance was well tolerated (Institut D'espertise Clinique 1996).

Skin Irritation and Sensitization

Glyceryl Dilaurate

Ivy Laboratories (1997) evaluated the skin sensitization potential of an eye shadow containing 1.5% Glyceryl Dilaurate (see preceding section, Skin Irritation, for additional results) in a maximization test using 29 healthy adult volunteers (19 females, 10 males; 18 to 60 years old). Twenty-five subjects completed the study. Three subjects were released due to lack of compliance, and one was released due to intolerance to sodium lauryl sulfate. During induction and challenge phases, patches were applied to the upper outer arm, the volar forearm, or to the back of each subject.

According to the induction procedure, 0.25% aqueous sodium lauryl sulfate (SLS) (≈ 0.1 ml) was applied under an occlusive patch that was fastened to the skin with occlusive tape for 24 h. After patch removal, the eye shadow containing 1.5%

Glyceryl Dilaurate was applied (same site), and the site was covered with occlusive tape for 48 h (or 72 h: weekend application). If skin irritation was not observed at the time of patch removal, the SLS patch was reapplied (same site) for 24 h. Patch removal was followed by reapplication of another induction patch with the eye shadow. This sequence of SLS pretreatment, followed by application of the eye shadow, was repeated for a total of five induction exposures. If skin irritation was observed during induction, SLS pretreatment was eliminated and eye shadow only was applied.

The challenge phase was initiated after a 10-day nontreatment period. Following 1 h of pretreatment with 5.0% aqueous SLS (under occlusive patch, new site), the subjects were challenged with a single application of the eye shadow at the same site. The new site was on the opposite arm, forearm, or side of the back. The challenge patch remained in place for 48 h, and sites were graded according to the following scale at 1 and 24 h post removal: 0 (not sensitized) to 3 (strong sensitization: large vesiculobullous reaction). Neither contact allergy nor any other adverse reaction was observed in any of the 25 subjects who completed the study. It was concluded that the eye shadow containing 1.5% Glyceryl Dilaurate does not possess a detectable contact-sensitizing potential, and, thus, is not likely to cause contact sensitivity reactions under normal use conditions (Ivy Laboratories 1997).

Ivy Laboratories (1999) also evaluated the skin sensitization potential of a foundation containing 1.5% Glyceryl Dilaurate according to the procedure in the preceding maximization test. The study group consisted of 28 healthy adult volunteers (19 females, 9 males; 18 to 47 years old). Twenty-seven subjects completed the study. One subject was lost to follow-up. Neither contact allergy nor any other adverse reaction was observed in any of the 27 subjects who completed the study. It was concluded that the foundation containing 1.5% Glyceryl Dilaurate does not possess a detectable contact-sensitizing potential, and, thus, is not likely to cause contact sensitivity reactions under normal use conditions.

Glyceryl Dioleate

Inveresk Research International Limited (1991) evaluated the skin irritation and sensitization potential of Glyceryl Dioleate in a human repeat insult patch test using 86 volunteers. The test laboratory stated that 82 subjects completed the study and that the reasons for withdrawal were not test substance-related.

The test substance (50% w/w in liquid paraffin, 0.3 ml) was applied to Webril pads located down the center line of a piece of Blenderm adhesive tape, and the patch strip was applied to the lateral surface of the upper arm. A patch scheme comprised two patch strips, each with three Webril pads. The subjects were also patch tested with liquid paraffin (100%), which served as the negative control. During the 3-week induction phase, patches were applied to the same test site for 24 h on Mondays, Wednesdays, and Fridays. Reactions were scored prior to each subsequent patch application according to the following scale: 0 (no

visible reaction) to 5 (bullous reaction). At week 6, challenge patches were applied to both arms of each subject (original site and new test site, respectively). Challenge reactions were scored at 48 and 96 h post application according to the same grading scale.

Glyceryl Dioleate induced minimal irritation (mild erythematous reaction in seven subjects) during induction. However, the results of a statistical comparison indicated that reactions at test sites were not significantly more irritating when compared to control sites. No reactions indicative of sensitization were observed during the challenge phase (Inveresk Research International Limited 1991).

Glyceryl Palmitate Lactate

Danisco Ingredients (1996) evaluated the skin irritation and sensitization potential of Glyceryl Palmitate Lactate ($50\% \ w/v$ in liquid paraffin) using 91 healthy volunteers (males and females, 18 to 65 years old) in a modified Draize repeat insult patch test. The study was classified as single blind. Patches consisted of a 5 cm wide strip of Scanpor tape to which 10 Finn chambers were fixed in pairs. The test substance was heated to 60° C and 0.02 g was placed directly into individual Finn chambers; 0.02 ml liquid paraffin was dispensed onto a filter disc in the chamber. Liquid paraffin was the negative control.

The 91 volunteers were divided into two groups, group 1 (39 subjects) and group 2 (52 subjects). In Group 1, the first set of induction patches was applied to the upper back for 23 h and subsequent patch applications were 47 h in duration. The patch test schedule for these subjects was as follows: days 1, 2, 4, 7, 9, 11, 14, 16, and 18. In group 2, 47-h induction patch applications were made according to the following schedule: days 1, 3, 5, 8, 10, 12, 15, 17, and 19. In both groups, induction patches were applied to the same site, unless a reaction stronger than mild erythema was observed. Reactions were scored after patch removal according to the following scale: 0 (no visible reaction) to 5 (bullous reaction). During the challenge phase, initiated on day 35, patches were applied to new sites on the upper back for 47 h.

Challenge reactions were scored at 1 and 49 h after patch removal. Classification of either test substance as irritating was based on 10% of the induction scores defined as >1 (mild erythema). Sensitization was defined as a rapid response to challenge patch application, characterized by severe erythema and edema (usually with papules and/or vesicles).

According to the test laboratory, 17 of the 91 subjects withdrew for reasons unrelated to patch testing; 74 subjects (64 women, 10 men) completed the study. Glyceryl Palmitate Lactate (50% w/v in liquid paraffin) did not induce skin irritation or sensitization (Danisco Ingredients 1996).

PHOTOTOXICITY/PHOTOALLERGENICITY

Studies on the phototoxicity/photoallergenicity of the glyceryl diesters were not identified in the published literature. As noted earlier, however, glyceryl monoesters do contain glyceryl diesters. Therefore, data on the phototoxicity and photo-allergenicity of a glyceryl monoester, Glyceryl Rosinate are included.

Glyceryl Rosinate

Biosearch, Inc. (1992a) reported a study in which the phototoxicity of a lipstick containing 1.0% Glyceryl Rosinate, as supplied, was evaluated using 10 volunteers (17 to 55 years old). All subjects were in good health and free of any visible skin disease or anomaly in the area of skin designated for patch testing. Subjects on medication (especially medications suspected of causing photobiological reactions or medications with the potential for modifying the inflammatory response) were excluded. The subjects were classified as Fitzpatrick skin types I, II, and III. The degree of skin pigmentation did not significantly influence responses to UV light or interfere with the scoring of skin reactions. The test substance was applied (approximately 20 mg/site) to two sites on the back of each subject and spread to cover the areas uniformly.

One of the test sites was irradiated with 0.5 MED (minimal erythemal dose; in seconds) of UVA and UVB light (continuous spectrum in UVA and UVB regions, 290 to 400 nm) between 30 and 60 min after application of the test substance. The MED was defined as the shortest exposure time at which erythema was first observed 20 ± 4 h after exposure. Irradiation with UVA and UVB light was followed by exposure to a total of 14 J/cm^2 of UVA. A 2 mm thick WG-345 Schott filter was interposed to eliminate UVB (290 to 320 nm) radiation from the ultraviolet source. Reactions were scored at 24, 48, and 72 h post irradiation according to the following scale: 0 (no visible erythema) to 3 (severe erythema [very intense redness]).

The second site to which the test substance had been applied was not irradiated and served as an irritation control. A third site served as the untreated, irradiated control.

Skin irritation was not observed at control or irradiated sites in any of the 10 subjects tested. The lipstick containing 1.0% Glyceryl Rosinate did not elicit a phototoxicity response (Biosearch, Inc. 1992a).

Biosearch, Inc. (1992b) also reported a study in which the photoallergenicity of a lipstick containing 1% Glyceryl Rosinate, as supplied, was evaluated using 26 volunteers (17 to 55 years old). Four of the original 30 subjects withdrew for personal reasons. All subjects were in good health and free of any visible skin disease or anomaly in the area of skin designated for patch testing. Subjects on medication (especially medications suspected of causing photobiological reactions or medications with the potential for modifying the inflammatory response) were excluded. Skin types were variable and the degree of skin pigmentation did not significantly influence responses to UV light or interfere with the scoring of skin reactions. During the induction phase, each of the subjects received six applications of the test substance over a period of 3 weeks. For each application, approximately 0.15 g of the test substance was placed on

an occlusive patch that was applied to the back for 24 h. Patches were applied on Tuesdays and Thursdays.

After patch removal, each site was exposed to 2.0 MEDs of UVB radiation and 4 J/cm² of UVA radiation. The subjects were instructed to keep the back covered throughout the study to avoid exposure to natural or artificial sunlight. The challenge phase was initiated 18 days after the last induction exposure. Challenge patches containing the test substance were applied to two new, adjacent test sites for 24 h. After patch removal, reactions were scored according to the scale indicated in the preceding study.

One of the test sites was then exposed to a combination of 0.5 MED of UVB and 4 J/cm² of UVA light. The other site was not exposed to UVA light and served as the irritation control. The UV light control site was defined as an additional site that was not exposed to the test substance, but was irradiated with 0.5 MED of UVB and 4 J/cm² of UVA light. Challenge sites were scored at 24, 48, and 72 h post irradiation.

No reaction was observed at control or test sites on any of the 26 volunteers tested. The lipstick containing 1.0% Glyceryl Rosinate did not elicit a photoallergic response (Biosearch, Inc. 1992b).

Case Reports

Glyceryl Diisostearate

Hayakawa et al. (1987) reported chelitis in an 18-year-old girl with a history of what was described as lip cream dermatitis. The patient was patch tested with Glyceryl Diisostearate, an ingredient of one of the lipsticks used, as well as Glyceryl Diisostearate impurities. Both Glyceryl Diisostearate (in petrolatum) and purified Glyceryl Diisostearate (no glyceryl monoisostearate peak in the gas chromatograph) were tested in this study.

The results of an analysis of Glyceryl Diisostearate by gas chromatography were as follows: Glyceryl Diisostearate (65.89%), glyceryl triisostearate (29.05%), glyceryl monoisosterate (0.43%), and isostearic acid (0.21%). Patch tests were conducted using Finn chambers that were secured with Scanpor tape for 48 h.

Reactions were scored according to International Contact Dermatitis Research Group (ICDRG) recommendations. A strong positive reaction (++) to Glyceryl Diisostearate (35.5% in petrolatum) was observed at 48 and 72 h. Purified Glyceryl Diisostearate induced + and ++ reactions at 48 and 72 h, respectively. All three Glyceryl Diisostearate impurities induced ++ reactions (48 and 72 h) at the following test concentrations in petrolatum: Glyceryl Monoisostearate (0.01%), isostearic acid (10%), and Glyceryl Triisostearate (27%). In an analysis of Glyceryl Triisostearate by gas chromatography, a peak for glyceryl monoisostearate was not observed (Hayakawa et al. 1987).

Tanaka et al. (1993) observed itchy, facial erythema in a 35-year-old female, following application of a foundation containing 1.77% Glyceryl Diisostearate. The reaction cleared within

a few days after withdrawal of the product and application of a mild, topical corticosteroid. In patch tests with individual ingredients of the product, a positive (+) reaction to 1.77% Glyceryl Diisostearate in petrolatum was found. Reactions were scored according to ICDRG recommendations.

The investigators stated that allergic contact dermatitis to this ingredient was extremely rare, and also noted that the patient also had sensitization reactions to lipsticks containing 35.5% Glyceryl Diisostearate. The following three chemical impurities were detected when Glyceryl Diisostearate was analyzed by gas chromatography: glyceryl monoisostearate (0.43%), glyceryl triisostearate (29.05%), and isostearic acid (0.21%). A strong positive reaction was observed when the patient was tested with 0.01% glyceryl monoisostearate (Tanaka et al. 1993).

SUMMARY

The safety of Glyceryl Dilaurate, Glyceryl Diarachidate, Glyceryl Dibehenate, Glyceryl Dierucate, Glyceryl Dihydroxystearate, Glyceryl Diisopalmitate, Glyceryl Diisostearate, Glyceryl Dilinoleate, Glyceryl Dimyristate, Glyceryl Dioleate, Glyceryl Diricinoleate, Glyceryl Dipalmitate, Glyceryl Dipalmitoleate, Glyceryl Distearate, Glyceryl Palmitate Lactate, Glyceryl Stearate Citrate, Glyceryl Stearate Lactate, and Glyceryl Stearate Succinate are considered in this safety assessment. These chemicals are also called diacylglycerols, diglycerides, or glyceryl diesters.

For Glyceryl Dioleate production, raw materials are fully refined nonlauric vegetable oils. The oils are further processed using special hydrogenation and fractionation techniques. The end products are produced by reacting selected mixtures of the partly hydrogenated, partly fractionated oils, and fats with vegetable-derived glycerine to yield partial glycerides. In the final stage of the production process, the products are purified by deodorization. Deodorization effectively removes pesticide residues and lower boiling residues such as residues of halogenated solvents and aromatic solvents. No manufacturing data were available for other diglycerides.

1,2-Diglycerides and 1,3-diglycerides are not stable and are easily isomerized. Although there are various reports of the relative proportion of the 1,2, 2,3, and 1,3 forms, the most recent analytical data demonstrate that 10% to 15% 1,2 diester (optically and physiologically active isomer) is present in the product as sold, with 10% to 15% 2,3 diester (optically active, physiologically non-active isomer) and 70% to 80% 1,3 diester (not optically or physiologically active) comprising the remainder of the product.

Diglycerides are used as skin conditioning agents—emollients in cosmetics. Reports to FDA in 2001 indicate that of the 18 ingredients in this safety assessment, Glyceryl Dilaurate, Glyceryl Diisostearate, and Glyceryl Dioleate are used in cosmetics (133 uses total). Concentration of use data received from the cosmetics industry in 1999 confirm the use of the above three glyceryl diesters, but also indicate use of Glyceryl Stearate

Lactate and Glyceryl Distearate. Current concentrations of use in cosmetics were: Glyceryl Dilaurate (up to 5%), Glyceryl Diisostearate (up to 43%), Glyceryl Dioleate (up to 2%), Glyceryl Distearate (up to 7%), and Glyceryl Stearate Lactate (up to 5%).

Diglycerides have been approved by FDA for use as indirect food additives. Reportedly, Glyceryl Dilaurate has been detected in pharmaceutical excipients.

Diglycerides are intermediates in the metabolism of triglycerides to free fatty acids and glycerol.

Various studies indicate that the sustained activation or inhibition of PKC (diacylglycerol-activated isoenzyme) in vivo may play a critical role in the regulation of long-term cellular events such as proliferation, differentiation and tumorigenesis. In vitro results indicate that the capacity for activation of PKC α is considerably greater for 1,2-diacylglycerols than for 1,3-diacylglycerols. However, mixed results have been reported in other in vitro experiments. Practically the same degree of activation of PKC α was induced by 1,2- and 1,3-diacylglycerols in one experiment and a very low capacity for 1,3-diacylglycerol-induced activation of PKC α was demonstrated in another. The activation of PKC α was dependent on the components of the micelle in which PKC was incorporated.

Exposure of murine keratinocytes to UVB radiation increases cellular diacylglycerol concentrations via changes in de novo metabolism that involve a diacylglycerol kinase pathway. Elevated diacylglycerol may influence signal-transduction pathways that are mediated by cellular lipids, and contribute to keratinocyte responses to UV light.

The topical application of diacylglycerols increased pigmentation in skin of outbred pigmented guinea pigs and female Skh HR-2 pigmented hairless mice. The increase in epidermal pigmentation was a PKC-dependent process. Treatment of the hairless mice with a diacylglycerol or the diacylglycerol + UVA/UVB light revealed no skin irritation or mucopolysaccharide deposition in excess of that anticipated from exposure to UV light alone.

An acute oral LD_{50} of >5 g/kg (Glyceryl Dilaurate) was reported in rats. The oral administration of a trade mixture containing an unspecified concentration of Glyceryl Dibehenate did not cause death in rats.

A mono-, di-, and triglyceride mixture including 45% Glyceryl Dilaurate did not induce any deleterious effects when fed to male and female albino rats at a dietary concentration of 25% (effective concentration of Glyceryl Dilaurate = 11.25%) for 2 years.

Undiluted Glyceryl Dilaurate was non-irritating to the eyes of albino rabbits. A trade mixture containing an unspecified concentration of Glyceryl Dibehenate was classified as a slight ocular irritant in a test involving rabbits. Glyceryl Stearate Citrate did not induce ocular irritation in the Draize test.

Glyceryl Dilaurate (undiluted) was classified as a mild primary irritant in a study involving albino rabbits, in which the

test site was covered with occlusive patches for 24 h. In another 24-h occlusive patch test, Glyceryl Dilaurate (melted in a water bath) was nonirritating to abraded and intact skin sites of albino rabbits. In a Draize skin irritation test using albino rabbits, Glyceryl Stearate Citrate induced slight skin irritation. However, results were negative for this ingredient in the Magnusson-Kligman guinea pig maximization test. The animals were challenged with 25% Glyceryl Stearate Citrate in petroleum jelly for 48 h; sites were covered with patches made of filter paper. In another guinea pig maximization test, Glyceryl Diisostearate did not induce sensitization at concentrations ranging from 5% to 100% in ethanol.

Triisostearin (C18) did not induce significant cutaneous reactions in a study evaluating the phototoxicity and photoal-lergenicity potential of this ingredient in guinea pigs. No significant cutaneous reactions, with or without UV irradiation, were found when the phototoxicity and photoallergenicity potential of Glyceryl Isostearate was evaluated using guinea pigs.

Two commercial glyceryl diesters, Glyceryl Dilaurate and Glyceryl Distearate, and the two diacylglycerols, 1-oleoyl-2-acetoyl-sn-glycerol (OAG) and 1,2-dipalmitoyl-sn-glycerol (DPG), did not alter cell proliferation (as determined by DNA synthesis) in normal human dermal fibroblasts in vitro at doses up to $10 \mu g/ml$.

Diacylglycerol Oil was not genotoxic, in that neither unheated Diacylglycerol Oil nor heated Diacylglycerol Oil induced bacterial gene mutation in the Ames test and did not induce clastogenicity in mammalian Chinese hamster lung cells. Also, there was no evidence that either unheated Diacylglycerol Oil or heated Diacylglycerol Oil had genotoxic potential in a rodent bone marrow micronucleus assay.

In the absence of initiation, Glyceryl Distearate (in acetone) induced a moderate hyperplastic response in randomly bred mice of a tumor-resistant strain. With DMBA initiation, an increase in the total cell count was observed. Whether Glyceryl Distearate increased the DMBA tumor incidence was not stated. In a glyceryl monoester study, a single application of DMBA to the skin was followed by 5% Glyceryl Stearate (in acetone) twice weekly. No tumors developed and slight epidermal hyperplasia at the site of application was noted.

Glyceryl Dioleate induced transformation in 3-methylcholanthrene-initiated BALB/3T3 A31-1-1 cloned cells in vitro.

In a study involving female CD-1 mice, PKC was linked to epidermal hyperplasia and tumor promotion. A tumor-promoting dosing regimen that consisted of multiple applications of 5 or 10 μ mol sn-1,2-didecanoylglycerol, a model sn-1,2-diacylglycerol and complete tumor promoter, twice daily for 1 week, caused more than a 60% decrease in cytosolic and particulate PKC activity and marked epidermal hyperplasia. However, applications of 10 μ mol sn-1,2-didecanoylglycerol twice weekly for 1 week, a non-tumor-promoting dosing regimen, caused a decrease in cytosolic PKC activity, increased particulate PKC activity and did not induce epidermal hyperplasia.

A study indicated that the overexpression of PKC α in the epidermis of transgenic mice increased the expression of specific proinflammatory mediators and induced cutaneous inflammation, but had little to no effect on epidermal differentiation, proliferation, or TPA tumor promotion. Other studies have shown that Diacylglycerols, intracellular ligands for PKC, were elevated in psoriatic lesions, and that this was accompanied by alterations in PKC isoform levels and activity.

The increased epidermal DNA synthesis induced by sn-1,2-dioleylglycerol or TPA after topical application to CD-1 mice was inhibited when either was administered simultaneously with fluocinolone acetonide, a potent inhibitor of tumor promotion. A single application or applications twice a week for four weeks of sn-1,2-dioleoylglycerol did not cause morphological changes in the epidermis or function as a complete tumor promoter when applied twice weekly for 28 weeks. More frequent applications of sn-1,2-didecanoylglycerol (5 μ mol twice a day for 5 days) induced epidermal hyperplasia that was comparable to that resulting from application of 5 nmol TPA twice weekly for 4 weeks.

The synthetic lipid second messenger, sn-1,2-didecanoylglycerol has been demonstrated to be a complete tumor promoter in DMBA-initiated mouse skin.

In studies on the tumor promoting activity of 1,2-diacylglycerols, it was noted that the structure of the particular diacylglycerol, its dose, and the exposure regimen by which the dose is delivered all play a role in tumor promotion. In particular, only a specific 1,2-diacylglycerol configuration was active and that the other configuration (also known as 2,3-diacylglycerol) was not active in tumor promotion.

Compared to one of the two control groups in a chronic oral study using groups of 60 SPF rats, a statistically significant increase in the number of either benign or malignant epithelial mammary gland neoplasms was observed in female rats fed a basal diet containing 5.3% diacylglycerol oil, but not in the remaining dose group fed a basal diet containing 2.65% diacylglycerol oil. These changes were not considered biologically significant because of the absence of a dose response and because the tumor incidences in the two control groups were not similar. No treatment-related gross or microscopic changes were observed in other tissues that were evaluated.

Findings relating to 1,2-diacylglycerol-induced activation of PKC are summarized as follows: (1) If the fatty acid in the 1 or 2 position is unsaturated, then the diacylglycerol can activate PKC, but 1,2-Diacylglycerols with two saturated fatty acids are less effective; (2) the stereospecificity of diacylglycerols is critical in terms of their biological activity, e.g., 1,2-sn-diacylglycerols, but not 2,3-sn-diacylglycerols, activates PKC; (3) the activities of different diacylglycerols (i.e., activation of PKC) have been compared. The activity of saturated 1,2-diacylglycerols was lost when the fatty acid moiety in the structure was a long-chain fatty acid. This effect was not only observed in in vitro kinase assays, but also in a cellular model (platelets). When the length of the acyl chain was greater than 14 carbons, the activity was very low or absent. However, the issue of unsaturated 1,2-diacylgly-

cerols potently activating PKC, even if the acyl chain is long, was raised.

Studies on the reproductive and developmental toxicity of glyceryl diesters were not found in the published literature.

An eye shadow containing 1.5% Glyceryl Dilaurate did not induce skin irritation in a single-insult, 24-h patch test involving 20 subjects. Mild skin irritation reactions to a foundation containing 1.5% Glyceryl Dilaurate were observed in 7 of 20 subjects tested according to the same procedure. A trade mixture containing an unspecified concentration of Glyceryl Dibehenate did not induce pathological irritation or significant cutaneous intolerance in a 48-h occlusive patch test involving eight female and two male subjects.

A histological evaluation was performed on skin from five of 30 female volunteers (18 to 56 years old) who had applied a prototype lotion or placebo formulation, both containing 0.5% Glyceryl Dilaurate, consecutively for 16 weeks (first phase: 30 subjects) or 21 weeks (second phase: 5 of the 30 subjects) in a within-subject, blinded study. Skin irritation was not observed in any of the 30 subjects tested during the first phase of the study. Neither localized erythema nor scaling was observed in any of the five subjects tested during the second phase. Biopsies (2 mm) taken from both legs of the five subjects indicated no recognizable abnormalities of the skin. The epidermis was described as normal in thickness, and there was no evidence of scaling, inflammation, or neoplasms in any of the tissues that were evaluated.

In a maximization test involving 25 subjects, it was concluded that an eye shadow containing 1.5% Glyceryl Dilaurate did not possess a detectable contact-sensitizing potential, and, thus, is unlikely to cause contact sensitivity reactions under normal use conditions. The same conclusion was made for a foundation containing 1.5% Glyceryl Dilaurate in a maximization test involving 27 subjects. Sensitization was not induced in any of 82 subjects patch tested with 50% w/w Glyceryl Dioleate (in liquid paraffin) in a repeat-insult, occlusive patch test. Mild erythema was observed in seven subjects during induction. Glyceryl Palmitate Lactate (50% w/v in liquid paraffin) did not induce skin irritation or sensitization in 74 subjects patch tested (Finn chambers) in a modified Draize repeat-insult patch test.

Phototoxicity was not induced in a group of ten healthy volunteers tested with a lipstick containing 1.0% Glyceryl Rosinate. Similarly, photoallergenicity was not induced in a group of 26 healthy volunteers patch tested (occlusive patches) with the same product in a repeated insult patch test.

A 35-year-old female subject had sensitization reactions to a foundation containing 1.77% Glyceryl Diisostearate, 1.77% Glyceryl Diisostearate in petrolatum (+ reaction), and lipsticks containing 35.5% Glyceryl Diisostearate. In another case report, an 18-year-old girl with a history of lip cream dermatitis was patch tested (48-h test, Finn chambers) with 35.5% Glyceryl Diisostearate in petrolatum. A strong positive reaction was observed at 48 and 72 h.

DISCUSSION

The available safety test data indicate that diglycerides in 1,3-diester configuration do not present any significant acute toxicity risk, nor are these ingredients irritating, sensitizing, or photosensitizing. Whereas no data are available regarding reproductive or developmental toxicity, there is no reason to suspect any such toxicity because the dermal absorption of these chemicals is negligible. The CIR Expert Panel also noted that, in the relevant studies, the terminology is "1,2-diacylglycerol" and not glycerol-1,2-diester.

Data regarding the induction of PKC and the tumor promotion potential of 1,2-diacylglycerols increased the level of concern. The Panel noted that nominally glyceryl-1,3-diesters contain 1,2-diesters, raising the concern that 1,2-diesters could potentially induce hyperplasia. The Panel did note that these compounds are more likely to cause these effects when the fatty acid chain length is ≤ 14 carbons, when one fatty acid is saturated and one is not, and when given at high doses, repeatedly.

Most of the glyceryl diesters have fatty acid chains longer than 14 carbons and none have mixed saturated/unsaturated fatty acid moieties. In addition, a 21-week use study of a prototype lotion containing 0.5% Glyceryl Dilaurate (a 14-carbon chain fatty acid) indicated no evidence of scaling, inflammation, or neoplasms in biopsy specimens. Also, DNA synthesis assays on Glyceryl Dilaurate and Glyceryl Distearate indicated that neither chemical altered cell proliferation (as determined by DNA synthesis) in normal human dermal fibroblasts in vitro at doses up to $10~\mu g/ml$.

The Panel understands that use testing is a common practice in industry and, if histopathology data are collected, can demonstrate an absence of epidermal hyperplasia. Because the concentration and use of these ingredients can vary (may be as high as 43% for Glyceryl Diisostearate in lipstick, and the frequency of application of these products can be several times daily), and because the proportion of glyceryl diesters that are inactive 1,3 isomers versus potentially biologically active 1,2 isomers is unknown, the Panel believes that each use should be examined to ensure the absence of epidermal hyperplasia during product development and testing.

In the absence of inhalation toxicity data on the diglycerides in this safety assessment, the Panel determined that these ingredients can be used safely in aerosolized products when packaging and use ensure that particulates are not respirable (are greater than $10~\mu m$). The Panel reasoned that the particle size of anhydrous hair sprays (60 to $80~\mu m$) and pump hair sprays (>80 μm) is large compared to the median aerodynamic diameter of $4.25~\pm~1.5~\mu m$ for a respirable particulate mass.

The CIR Expert Panel recognizes that certain ingredients in this group are reportedly used in a given product category, but the concentration of use is not available. For other ingredients in this group, information regarding use concentration for specific product categories is provided, but the number of such products is not known. In still other cases, an ingredient is not in current use, but may be used in the future.

Although there are gaps in knowledge about product use, the overall information available on the types of products in which these ingredients are used and at what concentration indicate a pattern of use. Within this overall pattern of use, the Expert Panel considers all ingredients in this group to be safe.

CONCLUSION

The CIR Expert Panel concluded that Glyceryl Dilaurate, Glyceryl Diarachidate, Glyceryl Dibehenate, Glyceryl Dierucate, Glyceryl Dihydroxystearate, Glyceryl Diisopalmitate, Glyceryl Diisostearate, Glyceryl Dilinoleate, Glyceryl Dimyristate, Glyceryl Dioleate, Glyceryl Dipalmitate, Glyceryl Dipalmitoleate, Glyceryl Dipalmitate, Glyceryl Dipalmitoleate, Glyceryl Stearate Citrate, Glyceryl Stearate Lactate, and Glyceryl Stearate Succinate are safe as cosmetic ingredients in the practices of use and concentrations as described in this safety assessment provided that the content of 1,2-diesters is not high enough to induce epidermal hyperplasia.

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