

Safety Assessment of Silk Protein Ingredients as Used in Cosmetics

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Abstract

The Expert Panel for Cosmetic Ingredient Safety (Panel) reviewed the safety of Hydrolyzed Silk and 9 other silk protein ingredients, which function primarily as skin and hair conditioning agents and bulking agents in cosmetic products. The Panel reviewed relevant data relating to the safety of these ingredients and concluded that 8 ingredients are safe in the present practices of use and concentration in cosmetics, as described in this safety assessment, but that the available data are insufficient for determining the safety of 2 silk protein ingredients in cosmetic products, MEA-Hydrolyzed Silk and Silkworm Cocoon Extract.

Keywords

cosmetics, safety, silk proteins

Introduction

The safety of the following 10 Silk protein ingredients as used in cosmetics is reviewed in this safety assessment:

Fibroin
Hydrolyzed Fibroin
Hydrolyzed Sericin
Hydrolyzed Silk
MEA-Hydrolyzed Silk
Sericin
Silk
Silk Extract
Silk Powder
Silkworm Cocoon Extract

According to the International Cosmetic Ingredient Dictionary and Handbook (Dictionary), these ingredients are reported to function as skin and hair conditioning agents and bulking agents in cosmetic products.¹

Silk is a fibrous protein that is a product of the silkworm, *Bombyx mori*, and spiders (*Nephila clavipes* and *Araneus diadematus*). Silk proteins are usually produced within specialized glands. These proteins are biosynthesized in epithelial cells and secreted into the lumen of these glands, where the proteins are stored prior to being spun into silk fibers.² The silkworm (*B mori*) is the source of silk from which cosmetic ingredients are derived.¹

MEA-Hydrolyzed Silk is evaluated in this safety assessment, and it should be noted that the Expert Panel for Cosmetic

Ingredient Safety (Panel) has previously issued a final amended report on other MEA-ingredients with the conclusion that the following ingredients are safe in the present practices of use and concentration described in the report (rinse-off products only) when formulated to be nonirritating³: ethanolamine MEA, ethanolamine HCl, MEA-benzoate, MEA-cocoate, MEA-laureth-6 carboxylate, MEA-laureth sulfate, MEA-lauryl sulfate, MEA-PPG-6 laureth-7 carboxylate, MEA-PPG laureth-6 carboxylate, MEA-PPG-8 steareth-7 carboxylate, MEA-salicylate, MEA-sulfite, MEA-tallowate, and MEA-undecylenate. The Panel also cautioned that these ingredients should not be used in cosmetic products in which *N*-nitroso compounds may be formed.

Data on skin depigmentation effects are included in this safety assessment; however, skin depigmentation is considered a drug, and not a cosmetic, effect in the United States, and is not within the Panel's purview.

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Chemistry

Definition and Structure

The Silkworm, *B mori*, produces silk proteins during the final stage of larval development. Two silk proteins, Fibroin and Sericin, have been distinguished as major components of silk cocoons.⁴ All of the ingredients in this report are related because they are derived from the silk fibers produced by *B mori*. The definitions and functions of Fibroin, Sericin, and other silk protein ingredients reviewed in this safety assessment are presented in Table 1.¹

Fibroin. Bombyx mori silk Fibroin was determined via nuclear magnetic resonance to exist as a repeated type II β-turn structure, wherein the conformation of one chain enables the formation of intramolecular hydrogen bonds with 2 juxtaposed (adjacent) chains.⁵

Sericin. Circular dichroism and infrared absorption spectra show that the molecular configuration of Sericin is mainly random crimp. 6,7 X-ray diffraction analysis and differential thermal analysis indicate that the assembled structure or Sericin powder is an amorphous structure 7 ; however, the amorphous Sericin transforms into a β -structure in the presence of water. 8

Chemical and Physical Properties

Properties of Fibroin, Sericin, and other silk protein ingredients are summarized in Table 2. Polarization microscopy shows that, in Silk, Sericin forms 3 layers surrounding a Fibroin fiber.⁵

Method of Manufacture

Silk. Fibroin (main protein of Silk) and Sericin (another Silk protein) are secreted by insect silk glands. Fibroin, in aqueous solution, is converted into silk fibers by a process that is called spinning. ^{10,11} According to another source, in the process of manufacturing lustrous Silk from the dried cocoons of silkworms, Fibroin is separated from Sericin, the other major component of the cocoon, by a degumming process, and the Sericin is mostly discarded in the wastewater. ⁹

Several methods have been reported for removing Sericin in the degumming process of cocoons. Practically all industrial removal processes involve extraction with soaps and detergents. Heat and acid extraction are other methods. Sericin extracted by different methods can yield different amino acid compositions. Additional information indicates that some commercial Silk is prepared from natural Silk by removing Sericin and that the purified aqueous Fibroin is dried and pulverized into a powder. 12

Hydrolyzed Silk. Hydrolyzed Silk has been reported to be prepared from the cocoon of the silkworm moth (*B mori*). ¹³ The silk thread is isolated from the cocoon and the fibers are cleaned and degummed. The individual silk fiber is then wound with other silk fibers to create one long thread. The threads are then combed to remove noils, which are short fibers considered

to be by-products of the textile industry. The noils are used in the production of Hydrolyzed Silk proteins through carefully controlled hydrolysis. The resultant material is a 5% solution of a water-soluble silk protein.

It has been reported that Hydrolyzed Silk (average molecular weight[MW] = 300 Da) may be prepared by acid, alkaline, or enzyme-catalyzed hydrolysis; Hydrolyzed Silk protein (average MW = 650 Da) may be prepared by alkaline or enzymatic hydrolysis. 14,15 These processes occur for several hours until the desired MW is reached. The final product is a 20% water solution of Hydrolyzed Silk protein (MW = 300 Da) or a 6.5% water solution of Hydrolyzed Silk protein (MW = 650 Da). Furthermore, another supplier has reported that Hydrolyzed Silk is prepared by acid and enzyme hydrolysis until the MW reaches the target range. 16

According to other sources, Hydrolyzed Silk is produced according to the following procedures ¹⁷⁻¹⁹—Procedure 1: (1) hydrolysis, (2) inactivation of hydrolytic agent, (3) filtration, (4) treatment, (5) concentration, and (6) sterilization. Procedure 2: (1) proteins hydrolyzed in water at specific pH and temperature for specific duration, (2) filtration to isolate desired components, (3) addition of quaternium-15, EDTA, and methylparaben, or just EDTA and methylparaben, and (4) make batch adjustments if needed (refiltration).

Composition/Impurities

Silk. Silk contains nitrogen (13%-20%) and, for material from one supplier, the reported or specified maximum concentration of heavy metals is 20 ppm. ¹²

Hydrolyzed Silk. Data on the composition of Hydrolyzed Silk are presented in Table 3. 20,21 Hydrolyzed Silk is marketed as an amino peptide concentrate that is rich in the 2 proteins that comprise natural Silk, Sericin and Fibroin. 22 It consists of $\sim 19\%$ Hydrolyzed Silk and also contains the preservatives phenoxyethanol (0.4%) and potassium sorbate (0.2%). 23 Other preservatives in Hydrolyzed Silk include quaternium-15, EDTA, and methylparaben. 18,19

Another source indicates that Hydrolyzed Silk (MW = 300 Da) is marketed as a 20% water solution and that Hydrolyzed Silk protein (MW = 650 Da) is marketed as a 6.5% water solution. Hydrolyzed Silk protein (MW = 300 Da) from one source was reported to contain heavy metals and arsenic at levels of \leq 4 ppm and 0.4 ppm, respectively. Hydrolyzed Silk protein from another source (MW = 650 Da) was reported to contain heavy metals and arsenic at \leq 10 ppm and 1 ppm, respectively. Is

According to another supplier, their Hydrolyzed Silk ingredients are marketed as aqueous solutions, 2 of which are 20% to 30% Hydrolyzed Silk and 27% to 32% Hydrolyzed Silk.²⁴

Fibroin. Silk derived from the silkworm *B mori* contains 2 major proteins, Fibroin and Sericin. Fibroin is a fibrous protein, present as a delicate twin thread in which the 2 strands are linked by disulfide bonds and enveloped by successive sticky layers of Sericin. ⁹ Individual filaments are large molecules (3700 amino

acids).²⁵ Fibroin has also been described as a glycoprotein composed of 2 comparably composed protein subunits covalently linked by disulfide bonds. Fibroin filaments have both crystalline and amorphous domains.² The amorphous domains are characterized by the presence of amino acids with bulkier side chains,²⁶ whereas the crystalline domains are characterized by high percentages of alanine, glycine, and serine.² Fibroin is a highly insoluble protein containing, as a whole, up to 90% of the amino acids glycine, alanine, and serine.²⁷ According to another source, Fibroin contains 46% glycine, 29% alanine, and 12% serine.²⁵

More detailed information on the composition of Fibroin, from the cocoon of the *B mori* caterpillar, indicates that it consists of 2 polypeptide chains or, more specifically, heavy and light chains of 391 kDaa and 25 kDaa, respectively; a disulfide bridge links the heavy chain to the light chain. The heavy chains contain 5263 residues, composed of 45.9% glycine, 30.3% alanine, 12.1% serine, 5.3% tyrosine, 1.8% valine, and only 4.7% of the other 15 amino acid types.

Sericin. Sericin, also referred to as silk glue, is a globular protein that constitutes 25% to 30% of silk proteins. It contains 18 amino acids, most of which have highly polar side-chains, containing hydroxyl, carboxyl, or amino groups. The highly hydrophilic nature of Sericin is due to the high content of serine and aspartic acid, approximately 33.4% and 16.7% of Sericin, respectively. The predominant amino acids comprising Sericin are serine, glycine, and glutamic acid, and side-chain hydroxyl, carboxyl, and amino groups enable easy cross-linking, copolymerization and blending with other natural or synthetic polymers. According to another source, Sericin contains 37% serine, 17% glycine, and 16% aspartate. Sericin contains 37% serine, 17% glycine, and 16% aspartate.

Depending on the solubility, Sericin can be separated into 3 fractions²: A, B, and C. Sericin A (17.2% nitrogen) comprises the outermost layer and is insoluble in hot water. Sericin B (16.8% nitrogen) is the middle layer and, on acid hydrolysis, yields the same amino acids as Sericin A and, additionally, tryptophan. Sericin C is the innermost layer, positioned adjacent to the Fibroin strands.

Because the method of production of Sericin involves extraction from cocoons using soaps and detergents, alkali soaps and detergents are typically present as impurities.⁹

USE

Cosmetic

The safety of the silk protein ingredients included in this safety assessment is evaluated based on the expected use of these ingredients in cosmetics. The Panel uses data received from the US Food and Drug Administration (FDA) and the cosmetics industry to determine expected cosmetic use. Use frequencies of individual ingredients in cosmetics are collected from manufacturers and reported by cosmetic product category in FDA's Voluntary Cosmetic Registration Program (VCRP) database. Use concentration data are submitted by Industry in response to surveys of maximum reported use concentrations, by product category, which are

conducted by the Personal Care Products Council (Council). Collectively, the use frequency and use concentration data indicate that 7 of the 10 Silk protein ingredients are currently being used in cosmetic products. According to these data, the following 3 Silk protein ingredients are not being used in cosmetics:

Fibroin
MEA-Hydrolyzed Silk
Silkworm Cocoon Extract

According to 2015 VCRP data, the greatest reported use frequency is for Hydrolyzed Silk (675 formulations, mostly rinse-off), followed by Silk Powder (177 formulations, mostly leave-on) (Table 4).³⁰ Lower use frequencies are reported for the remaining silk ingredients. The results of a concentration-of-use survey conducted in 2014 indicated that Silk Powder had the highest maximum concentration of use; it was used at concentrations up to 1.4% in leave-on products (face powders; Table 4).³¹ In some cases, reported uses appear in the VCRP database, but concentrations-of-use data were not provided. For example, Hydrolyzed Sericin is reported as used in 4 cosmetic formulations, but use concentration data were not submitted.

Cosmetic products containing Silk proteins may be applied to the skin and hair or, incidentally, may come in contact with the eyes and mucous membranes. Products containing these ingredients may be applied as frequently as several times per day and may come in contact with the skin or hair for variable periods following application. Daily or occasional use may extend over many years.

Hydrolyzed Silk and Silk Extract are used in hairspray at maximum concentrations up to 0.024\% and 0.0036\%, respectively. Silk Powder is also used in hairspray (maximum concentration 0.02%). Hydrolyzed Fibroin and Silk Powder are used in perfume at maximum concentrations up to 0.000047\% and 0.1\%, respectively. Maximum use concentrations for the following ingredients in face powders are reported: Sericin (0.00047%), Silk (0.1%-0.2%), and Silk Powder (0.1%-1.4%). In practice, 95% to 99% of the droplets/particles released from cosmetic sprays have aerodynamic equivalent diameters >10 um, with propellant sprays yielding a greater fraction of droplets/particles below 10 μm, compared with pump sprays. 32-35 Therefore, most droplets/particles incidentally inhaled from cosmetic sprays would be deposited in the nasopharyngeal and bronchial regions and would not be respirable (ie, they would not enter the lungs) to any appreciable amount. 32,33

The silk proteins reviewed in this safety assessment do not appear on the list of ingredients prohibited from use in cosmetic products marketed within the European Union (Annex II of the Council of the European Communities Council Directive) or on the list of ingredients with use restrictions in products marketed within the European Union (Annex III).³⁶

Noncosmetic

Fibroin. Silk fibers made from ibroin have many uses in textiles (medical and industrial applications) mainly because of the

unique properties of Fibroin, such as water absorbency, dying affinity, thermo-tolerance, luster, and insulation properties. Fibroin is also a raw material for producing precious fabrics, parachutes, tire lining materials, artificial blood vessels, and surgical sutures. Natural, nonabsorbable Silk surgical suture containing the organic protein, Fibroin, is an FDA-approved medical device. 37

Toxicokinetics

Toxicokinetics studies of the silk proteins reviewed in this safety assessment were not found in the published literature, and unpublished data were not submitted.

Toxicology

Single Dose (Acute) Toxicity

Oral

Hydrolyzed Silk. The acute oral toxicity of Hydrolyzed Silk (MW ~ 300 Da; acid and enzyme hydrolysis product) was evaluated using rats (5 males, 5 females; strain not stated). A single dose of 10 g/kg was administered orally to each animal. No signs of toxicity were observed during the 14-day observation period after dosing.

In another study, the acute oral toxicity of Hydrolyzed Silk protein (MW ~ 1000 Da; produced via alkali hydrolysis) was evaluated using albino rats (5 males, 5 females). A single dose of the test material (5 g/kg body weight) was administered using an intragastric feeding needle. Signs of toxicity were not observed during the study and none of the animals died. The median lethal dose (LD₅₀) was >5 g/kg.

The acute oral toxicity of Hydrolyzed Silk protein (15%-25% in water; specific gravity = 1.10) was studied using 10 albino rats (5 males, 5 females).³⁹ The test substance was administered orally at a dose of 5 g/kg, and dosing was followed by a 14-day observation period. Gross necropsy was performed on all animals. Only 1 animal (male) died, and thoracic cavity filled with fibrous tissue was noted at necropsy. Whether or not this death was due to experimental error was not stated. Gross changes were not observed in the remaining animals. The test substance was classified as nontoxic.

Silk. Ten male Sprague-Dawley rats were dosed orally (16 g/kg) with Silk. ⁴⁰ The form of Silk administered, test concentration, vehicle, and dosing method were not stated. Dosing was followed by a 14-day observation period. None of the animals died, and except for slight lethargy, there were no signs of toxicity during the observation period. The oral LD₅₀ was >16 g/kg, and Silk was considered nontoxic in this study.

Silk Powder. A 30% solution of Silk Powder in distilled water was administered orally (feeding tube) to 12 female DD-strain mice. The animals received doses up to 12 g/kg body weight (dose volume = 40 mL/kg). Dosing was followed by a 7-day observation period. Toxic signs were not observed during the

study, and the test substance was classified as practically non-toxic ($LD_{50} > 12$ g/kg body weight).

Repeated Dose Toxicity

Dermal

Hydrolyzed Silk. In a cumulative skin irritation study involving 8 Hartley guinea pigs, a 6.5% aqueous solution of Hydrolyzed Silk protein (MW = 650 Da) was applied to the back once daily for 35 days. ⁴² The animals were killed and necropsied at the end of the dosing period. Body weight gain was normal, and no abnormalities were noted at necropsy. Other study results relating to skin irritation potential are included in Table 5.

Cytotoxicity

Sericin obtained via urea extraction was slightly toxic to mouse fibroblasts in vitro at concentrations as low as 60 μ g/mL, and toxicity was substantial (ie, severely harmful) at concentrations greater than 100 μ g/mL. When using other extraction methods (heat, acid, or alkaline), Sericin yielded less toxicity, as measured by the percentage of viable fibroblasts.⁴⁴

Skin Depigmentation

Sericin. Sericin was formulated as an 8% cream and applied to one side of the extremity (arm and leg) of renal patients who normally experienced dry and itchy skin. A cream base was applied to the other extremity and served as the control. From 47 subjects who completed the study, the skin hydration of the patients' extremities increased after receiving both Sericin cream and the cream base, but the changes in skin hydration were much greater on the side receiving the Sericin cream than on the side receiving the cream base. Additionally, at the end of the study, the skin pigmentation level was significantly reduced on both the arms (P = 0.032) and legs (P = 0.021) of the Sericin-treated side compared with the side treated with cream base.

The depigmentation effect was presumed to result from tyrosinase inhibition. The degree of inhibition of tyrosinase (the rate-limiting enzyme for melanin production) activity by Sericin is known to depend upon the extraction method and silk strain source. ⁴⁶ For example, colored silk cocoons, which contain flavonoids and carotenoids, exhibit higher anti-tyrosinase activity than white-shelled cocoons.

Reproductive and Developmental Toxicity

Reproductive and developmental toxicity studies of the silk proteins reviewed in this safety assessment were not found in the published literature, and unpublished data were not submitted.

Genotoxicity

Hydrolyzed Silk

The genotoxicity of Hydrolyzed Silk protein (10% aqueous) was evaluated in the Ames test using the following *Salmonella typhimurium* strains, with and without metabolic activation: TA98, TA100, TA1535, TA1537, and TA1538.⁴⁷ Results were negative in all bacterial strains. The positive and negative controls performed as expected in this assay.

Carcinogenicity

Cell Proliferation

Sericin. The effect of Sericin on the rat insulinoma cell line (seeded on ASF104 culture medium [serum-free medium containing insulin and transferrin]) was evaluated. Bovine serum albumin (BSA) served as the control protein. The RIN-5F cell cultures were identified as follows: ASF104 (1 mL), ASF104 with 0.1% Sericin, and ASF104 with 0.1% BSA. The cells were cultured for 22 hours. Viable and nonviable cell numbers were determined using the trypan blue exclusion method. The cells in the control culture failed to proliferate. However, RIN-5F cells propagated significantly in the presence of Sericin (P < 0.05) or BSA (P < 0.01). Therefore, Sericin and BSA were efficient inducers of RIN-5F cell proliferation.⁴⁸

Anticarcinogenicity

Sericin

A study was performed to evaluate the protective effect of Sericin on tumor promotion in the 7,12 dimethylbenz[a]anthracene (DMBA)-initiated and 12-O-tetradecanoylphorbol-13acetate (TPA)-promoted mouse skin tumorigenesis model.⁴⁹ In the first experiment, Sericin was applied topically to DMBA (190 nM)-initiated, female CD-1 (ICR): Cri mice (groups of 16) at doses of 2.5 mg and 5 mg twice weekly for 16 weeks. Sericin was applied 30 minutes prior to each promotion treatment with TPA (3.2 nM). The tumor necrosis factor α (TNF- α , pro-inflammatory cytokine) and 4-hydroxynonenal (4-HNE) staining method was used, whereby the affinity-purified goat polyclonal TNF-α antibody and monoclonal antibody were placed on the specimens. Sericin caused a significant reduction in both the tumor incidence and tumor multiplicity at doses of 2.5 mg and 5 mg per application, compared to the control group that was not treated with Sericin. The expression of TNF-α protein and the level of 4-HNE in normal epidermis were significantly reduced in both Sericin treatment groups.

In the second experiment (groups of 5 ICR mice), Sericin (5 mg) was applied topically to dorsal skin 30 minutes prior to application of TPA. The same doses of TPA and Sericin (used in first experiment) were applied twice at an interval of 24 hours. Treatment with Sericin inhibited double TPA treatment-induced morphological changes; these were indicative of an inflammatory response, that is leukocyte infiltration, hyperplasia, and cell proliferation. Treatment with Sericin also

significantly suppressed the elevation of 4-HNE levels and elevated expressions of c-fos, c-myc, and cyclooxygenase-2 (COX-2) in normal epidermis that were induced by double application of TPA. The results of this study suggest that Sericin has a protective effect against tumor promotion in mouse skin by suppressing oxidative stress, inflammatory responses, and TNF- α . ⁴⁹

In another study, male CD-1 (ICR): Crj mice (2 groups of 11 and 12, respectively) were fed diets supplemented with 1.5% Sericin (3 g/kg/d) or 3% Sericin (6 g/kg/d) for 5 weeks. ⁵⁰ The animals also received weekly injections of 1,2 dimethylhydrazine (DMH) during the initial 3 weeks of the study. The feeding of Sericin in the diet resulted in a dose-dependent decrease in the development of colonic aberrant crypt foci. In a second experiment, mice were fed a diet supplemented with 3% Sericin for 115 days. The animals were also injected with DMH weekly during the initial 10 weeks. Both the incidence and number of colon tumors were suppressed by Sericin consumption.

A study was performed to assess the protective effect of Sericin on mid-wavelength ultraviolet light (UVB)-induced acute damage and tumor promotion in HR-1 hairless mouse skin.⁵¹ Three groups of 10 mice were treated dermally with Sericin, BSA, and vehicle (ethanol), respectively, in the first experiment. One group of mice was treated with 180 mJ/cm² UVB light once daily for 7 days, after which red sunburn lesions of the skin were observed. Both the area and the intensity of the redness of these lesions were reduced by the topical application of 5 mg Sericin immediately after UVB treatment. The differences (area and intensity of the redness) between the vehicle and Sericin groups were statistically significant (P < 0.01). This was not true when the group treated with BSA (5 mg), rather than Sericin, was compared to the vehicle control. The results of immunohistochemical analyses indicated that the application of Sericin suppressed UVB-induced elevations in 4-HNE, expression of COX-2 protein, and proliferating cell nuclear antigen-labeling index in the UVB-exposed epidermis.

Three groups of 15 mice of the same strain were treated with Sericin, BSA, and vehicle (ethanol), respectively, in the second experiment. One group of mice was treated (dermal application) with 200 nmol DMBA, followed by a 1-week nontreatment period. 7,12 dimethylbenz[a]anthracene-treated skin was then irradiated with 180 mJ/cm² of UVB twice weekly, and each irradiation was followed by topical treatment with Sericin (5 mg). Another group of mice was treated similarly with BSA (5 mg), rather than Sericin. Treatments (UVB dosing, followed by topical treatment) were repeated for 22 weeks. A statistically significant reduction in both tumor incidence and multiplicity was noted at a dose of 5 mg, indicative of a suppressive effect of Sericin. When compared to all of the animals in the vehicle and BSA groups having skin tumors 22 weeks after the topical application of DMBA, only 6\% of the DMBA-exposed mice in the Sericin-treated group exhibited skin tumors, indicating 94% (P < 0.001) reduction in tumor incidence. Similarly, when the tumor data were evaluated for tumor multiplicity (ie, the number of tumors per mouse), from the first tumor appearance to the termination of the experiment, Sericin produced statistically significant (P < 0.05) protection against UVB-induced tumor promotion in DMBA-exposed mouse skin. The results of this study (including the first and second experiments) suggest that Sericin possesses a photoprotective effect against UVB-induced acute damage and tumor promotion by reducing oxidative stress, COX-2, and cell proliferation in mouse skin. ⁵¹

Irritation and Sensitization

Skin Irritation and Sensitization

Skin irritation and sensitization studies on silk protein ingredients are summarized in Table 5. These ingredients are, at most, mild skin irritants and lack skin sensitization potential. In the only available human skin irritation/sensitization tests in which test concentrations were reported, 20% aqueous Hydrolyzed Silk was a nonirritant and a non-sensitizer and 6.5% aqueous Hydrolyzed Silk was a nonirritant (skin irritation only evaluated).

Allergenicity

Human

Silk. The relationship between Silk sensitization and asthma incidence was evaluated in 871 children living in China. ⁵² Skin testing was performed using a slightly modified version of the semiquantitative puncture method. The results of multivariate analyses of asthma incidence and skin test reactivity to aero-allergens were presented. Individual skin test results were not provided. Children who were sensitized to Silk had 2.6 times higher odds of having asthma than did nonreactors, after adjustment for age, gender, familial correlations, and skin test reactivity to other aeroallergens using generalized estimating equations. This association between sensitization to Silk and asthma yielded lower statistical *P* values when the eosinophil counts of the participants were included as either a categorical variable or a linear term in the multivariate model.

Sixty-four (<15 years old; males and females) children with Silk-induced asthma in China were studied.⁵³ The diagnosis was based on a history of wheezing, positive skin tests to Silk, positive nasal or conjunctival provocation tests, or serum immunoglobulin E (IgE)-Silk waste (serum antibodies against Silk waste [severely broken Silk threads, used only as filling for bed quilts or clothes and mattresses]). The average age of asthma onset was 4 years 2 months. Conjunctival provocation tests were performed on 80% of the cases. The first symptom was observed an average of 10 months after initial exposure to Silk. Asthma was accompanied by allergic rhinitis in 61% of the patients and was accompanied by conjunctivitis in 14% of the cases. In most cases, asthma occurred during the winter, due to the seasonal use of bed quilts or clothes filled with Silk. The average mean wheal diameter elicited by Silk in prick tests was greater than the diameters measured from 2 histamine equivalent prick tests per Silk-sensitive subject.

In relation to the preceding study, it should be noted that allergenic proteins have been extracted from 1 Silk batch that was imported to be used as filling material for bed mattresses and rugs.⁵⁴ Immunoglobulin E and immunoglobulin G (IgG) antibodies to the extracted Silk proteins were measured by radioallergosorbent in sera of 9 Silk-sensitive subjects and in sera of healthy control subjects. Immunoglobulin E and IgG antibodies to the individual Silk polypeptides (40 kDa, 45 kDa, and 70 kDa) were detected using the immunoblot technique; the 6 kDa Silk polypeptide bound only IgG. The sera of Silksensitive subjects contained high titers of IgE and low titers of IgG antibodies to the separated Silk polypeptides. Low IgG antibody titers to a limited number of these polypeptides were detected in sera of control subjects. In another study involving 10 patients with a positive skin prick test to Silkworm crude extract, arginine kinase (42-kDa protein) was identified as a major allergen in this crude extract.55

Phototoxicity

Hydrolyzed Silk. The phototoxicity of 6.5% aqueous Hydrolyzed Silk protein (MW = 650 Da; produced by alkaline and enzyme hydrolysis) was evaluated using groups of 6 Hartley guinea pigs. The 3 groups were identified as test, positive control, and negative control groups. 8-Methoxypsoralen (1%) served as the positive control. The negative control was not stated. The test material was applied topically (dose = 0.05 mL/2 \times 2 cm) to 2 sites on dorsal skin. One site was irradiated once with a FL-40 S lamp and BLP lamp (wavelength range not stated), and the other site was covered. The sites were examined macroscopically at 24, 48, and 72 hours. Phototoxicity was evaluated based on the difference in severity of skin reactions between the irradiated and non-irradiated sites. Hydrolyzed Silk protein was classified as non-phototoxic. Positive responses were observed in all guinea pigs treated with 8-methoxypsoralen + light.

Silk Powder. Silk Powder (0.1 g) was applied to the back (2 sites (2 areas per site) of each of 6 female guinea pigs of the Hartley strain. ⁴¹ The test site (cm² area not stated) was covered with a patch plaster for 4 hours, after which the sites were irradiated with UV (minimal erythema dose, 15-minute exposure) from a 20-W lamp. The test sites were evaluated for erythema and eschar formation after 24 and 48 hours. Positive reactions were not observed in this study.

Photoallergenicity

Hydrolyzed Silk. The photoallergenicity of 6.5% aqueous Hydrolyzed Silk protein (MW = 650 Da; produced by alkaline and enzyme hydrolysis) was evaluated using groups of 6 Hartley guinea pigs. The 3 groups were identified as test, positive control, and negative control groups; the positive control was 3,5,4'-tribromosalicylanilide (2% in 85% dimethyl sulfoxide), but the negative control was not stated. The test material was applied transdermally (0.05 mL/2 \times 2 cm), with or without UV, 5 times per week (2 hours per day) for a total of 10 applications.

Applications were made on both sides of the dorsal area, symmetrically. One side was irradiated for 2 hours, and the other side was covered. The photo challenge phase was initiated after a 2-week nontreatment period. The test material was applied to 2 sites. One side was irradiated for 2 hours, and the other side was covered. The application site was examined macroscopically after the challenge and 24, 48, and 72 hours later. No effects were observed in negative controls or in guinea pigs treated with the test material (with or without exposure to light). Hydrolyzed Silk protein was considered non-photosensitizing in this study. The expected results were achieved with the positive control.

Case Reports

According to one case report, recurrent granulomas with remarkable infiltration of eosinophils may have resulted from an IgE-mediated hypersensitivity reaction to silk Fibroin, a component of the braided Silk suture used.⁵⁸ In this report, a lateral skin flap technique had been performed to correct tracheostomal stenosis, using Silk sutures, after a total laryngectomy.

Adverse reactions to virgin Silk sutures in 12 cataract surgery patients have also been reported.⁵⁹ Nodular episcleritis, peripheral corneal ulceration, and wound necrosis with dehiscence were observed, sometimes resulting in endophthalmus or epithelial downgrowth. Conjunctival and scleral histopathologic studies in 4 eyes showed acute and chronic inflammation with multinucleated giant cells. Type I allergic responses and upregulated levels of specific IgE were reported to occur in patients after repeated surgical procedures.^{58,60}

A female patient with a history of severe atopic dermatitis and various allergies, and a family history of atopic eczema and asthma, presented with exacerbation of eczema on her hands and wrists and urticarial papules on the flexor aspects of both forearms. The same types of lesions had been observed on both arms after wearing the same Silk shirt but quickly disappeared after the shirt was taken off. Radioallergosorbent tests yielded positive results for Silk waste (k73) of 2.08 universal arbitrary units (Ua)/mL and positive results for Silk (k74) of 3.62 Ua/mL. The total IgE count was >5000 kilo units (kU)/L. These results confirmed the diagnosis of immunological contact urticaria caused by Silk.

Ocular Irritation

Animal

Hydrolyzed Silk. The ocular irritation potential of Hydrolyzed Silk protein (15%-25% in water) was evaluated in the Draize test using 6 New Zealand white rabbits.⁶² The test substance (0.1 mL) was instilled into 1 eye of each animal, and eyes were not rinsed. Contralateral eyes served as controls. Observations for ocular reactions were made up to 72 hours post-instillation. The test substance was practically nonirritating to the eyes of rabbits.

The ocular irritation potential of 6.5% aqueous Hydrolyzed Silk (MW ~ 300 Da) was studied using 6 New Zealand white rabbits.⁶³ The test material (0.1 mL) was instilled into the right

eye of each animal, and the left eye served as the untreated control. Reactions were scored at 24, 48, and 72 hours post-instillation. Slight conjunctival redness, the only reaction reported, was observed in 1 rabbit. It was concluded that it is not likely that Hydrolyzed Silk would be classified as an ocular irritant, according to the definitions of the US Federal Hazardous Substances Act. The ocular irritation potential of a higher MW Hydrolyzed Silk (MW = 650 Da; test concentration not stated) was evaluated in New Zealand white rabbits according to the same test procedure, and the results were negative. 64

Hydrolyzed Silk (MW ~ 1000 Da; produced by alkali hydrolysis) was placed (0.1 mL) in the right eye of each of 6 New Zealand white rabbits.³⁹ Observations for any signs of corneal opacity, iritis, or conjunctivitis were made at 24, 48, and 72 hours post-instillation. The authors concluded that Hydrolyzed Silk was practically nonirritating to the eyes of rabbits.

In a cumulative ocular irritation test, 6.5% aqueous Hydrolyzed Silk protein (MW = 650 Da; 0.1 mL; produced by acid, alkaline, and enzyme hydrolysis), was instilled into the eyes of 6 New Zealand white rabbits 3 times per day for 4 days continuously. Conjunctival redness was observed in 5 of 6 animals at 3 or 4 days. Reactions in the cornea or iris were not observed. The authors concluded that Hydrolyzed Silk protein was practically nonirritating to the eyes of rabbits.

Silk. Silk (0.1 g) was instilled into 1 eye of each of 9 adult albino rabbits. The eyes of 3 rabbits were rinsed immediately after instillation. Untreated eyes served as controls. The eyes were examined at 24, 48, and 72 hours post-instillation. Transient conjunctival redness (unrinsed eyes) was observed in 5 of 6 rabbits. However, no effects on the cornea or iris were observed. Ocular irritation was not observed in the 3 rabbits subjected to ocular rinsing. Silk was classified as a nonirritant in this study.

Silk Powder. The ocular irritation potential of a Silk Powder solution (10% in saline) and the supernatant fluid from this solution (filtered after 24 hours) was evaluated using a total of 6 white rabbits. Four rabbits received the 10% in saline solution and 6 rabbits received the supernatant. Either test substance (0.1 mL) was instilled into the right eye, and eyes were rinsed. Untreated left eyes served as controls. The eyes were examined for reactions for up to 168 hours post-instillation. The Silk Powder solution (10% in saline) was classified as slightly positive. The supernatant from this solution was not an ocular irritant.

In Vitro

Hydrolyzed Silk. The ocular irritation potential of Hydrolyzed Silk protein (MW = 300 Da; 2% active solution) was evaluated in the in vitro hen's egg test on the chorioallantoic membrane (HET-CAM). The material (0.3 mL) was tested on the chorioallantoic membrane of fertilized Leghorn hens' eggs that had been incubated for 10 days. Results were negative (score = 0.3).

Hydrolyzed Silk protein (MW = 300 Da; 10% active solution) was tested for ocular irritation potential in the in vitro red

blood cell aggregation test, which evaluates effects on the cytoplasmic membrane. A total irritation classification was obtained by determining the hemolysis/denaturation (L/D) ratio. A substance with an L/D of >100 was classified as a nonirritating. Hydrolyzed Silk caused neither hemolysis nor denaturation and was classified as nonirritating.

The Irritection assay was used to evaluate the ocular irritation potential of Hydrolyzed Silk. The test material was applied to the Irritection system at dose volumes of 25, 50, 75, 100, and 125 μ L. The samples remained at room temperature for 24 hours and were then analyzed by spectrophotometry. Over the range of dose volumes tested, ocular Irritection scores for Hydrolyzed Silk ranged from 2.5 to 3.5. Scores in this range corresponded to a classification of minimally irritating.

The ocular irritation potential of Hydrolyzed Silk was studied using the EpiOcular model assay. ⁶⁹ The test material was applied to a reconstructed human corneal epithelial model for 30 minutes, and cell viability was measured by dehydrogenase, present in the cell mitochondria, reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), into blue, MTT formazan salt. The irritation potential of the test material is dictated by the reduction in tissue viability of exposed tissues, compared to the negative control (sterile deionized water). Methyl acetate served as the positive control. Hydrolyzed Silk was classified as a nonirritant. The negative and positive controls were nonirritating and irritating, respectively.

Other Effects

Immunological Responses

Sericin and fibroin

In Vitro. Soluble Sericin proteins (2 μg of Sericin per well tissue culture plate) extracted from native Silk fibers did not induce significant macrophage activation. ⁷⁰ Macrophages exposed in vitro to the Silk preparations failed to respond with consistently elevated levels of TNF in either short- or long-term cultures. However, the suspension of the crystalline particles prepared by enzymatic digestion of Silk Fibroin was the only Silk preparation that yielded significant TNF release, which was probably a nonspecific response to insoluble physical particulates, rather than a specific, chemically induced response to Silk. Whether or not the statistical significance of this finding was determined was not stated. However, the average TNF release (corrected for volume and expressed as total release from specified cell count) and standard error of the mean were determined.

A study was designed to investigate the inflammatory mediators induced by Sericin. The Sericin was analyzed in an in vitro macrophage (mouse alveolar macrophage cell line) and monocyte (mouse monocyte cell line) assay to assess cytokine activation. These cell lines were used for monitoring levels of interleukin (IL)-1 β and TNF- α generated after activation by Sericin at concentrations of 0.2 to 1.0 mg/mL. Inflammatory

mediators activated by Sericin were also investigated in vivo, using a rat wound-healing model. At the concentrations tested, silk Sericin increased the amounts of inflammatory mediators and pro-inflammatory cytokines, TNF- α and IL-1 β (involved in the modulation of skin growth, repair, and scarring during inflammation). However, the maximum levels of TNF- α and IL-1 β released from monocytes and macrophage cells after silk Sericin exposure were 500 and 350 pg/mL, respectively. It was noted that these levels of cytokines would not be sufficient to cause an inflammatory response or prevent cellular proliferation.

The suppression of inflammation by Sericin has been reported. ⁷² Sericin solution (0.004-0.080 mg/mL) applied topically to the top of the hind paw of rats prior to a carrageenan subcutaneous injection under the plantar surface of the hind paw exhibited anti-inflammatory activity, similar to the effect of indomethacin (a nonsteroidal anti-inflammatory drug used as a control). The amount of mast cells in rat tissue treated with Sericin or indomethacin was much lower compared to the number of cells found in tissue treated with water (control). Further investigation indicated that Sericin did not cause a hypersensitivity reaction. On the contrary, it inhibited COX-2 and inducible nitric oxide synthase (iNOS) production (monitored by total RNA and real-time polymerase chain reaction) in fibroblast cell culture, resulting in lowering the inflammation of the carrageenan induction.

Wound Healing

Nonhuman

Sericin. The effect of a Sericin cream on wound healing was evaluated using 18 male Sprague-Dawley rats (8 weeks old). The composition of the cream was described as follows: 8% Sericin, white petrolatum, mineral oil, lanolin, glycerin, bisabolol, propylparaben, and methylparaben. Except for Sericin, the concentration of each cream component was not stated. The cream was applied topically to full-thickness skin wounds on the dorsum of each animal, and wound surfaces were observed for 15 days post-application. Cream base without Sericin served as the control. Histological examination of wounds after 15 days of treatment with 8% Sericin cream revealed complete healing, no ulceration, and an increase in collagen, as compared to treatment with the control cream. Wounds treated with the control cream had some ulceration and acute inflammatory exudative materials.

In a similar study involving 45 Sprague-Dawley rats (8 weeks old), 8% Sericin cream was applied to full-thickness wounds on the dorsum of each animal. Cream base without Sericin served as the control. Excised rat tissue was prepared for cytokine determination. Interleukin-1 β and TNF- α are pro-inflammatory cytokines that are involved in a variety of immunological functions. Wounds treated with Sericin cream did not yield significantly high levels of IL-1 β and TNF- α on day 7, which suggests that the cream did not induce an inflammatory or immunological response.

In Vitro

Sericin. Human skin fibroblasts were incubated with Sericin in vitro for 72 hours. The cell count in treated cultures after 72 hours was enhanced to 250% of the untreated (ie, no-Sericin) control cultures. In another study, replacing the culture medium with Sericin solution in the mouse L929 fibroblastic cell line in culture increased the percentage of cell proliferation significantly, especially at a high Sericin concentration (1.0 mg/mL). The amount of NF-1 α and IL-1 β released from alveolar macrophage NR8383 and mouse J774.2 monocyte cell lines after the addition of Sericin (0.2-1.0 mg/mL) to the culture media was negligible, indicating that Sericin did not cause severe damage to the cells.

Summary

The safety of the following 10 silk protein ingredients in cosmetics is reviewed in this safety assessment: Fibroin, Hydrolyzed Fibroin, Hydrolyzed Sericin, Hydrolyzed Silk, MEA-Hydrolyzed Silk, Sericin, Silk, Silk Extract, Silk Powder, and Silkworm Cocoon Extract. These ingredients are reported to function as skin and hair conditioning agents and bulking agents in cosmetic products. Frequency of use data from the FDA's VCRP and the results of an industry survey indicate that 7 of the 10 Silk protein ingredients are being used in cosmetic products. Silk Powder has the highest reported maximum concentration of use; it is used at concentrations up to 1.4% in leave-on products (face powders).

The silkworm, *B mori*, produces Silk proteins during the final stage of larval development, and 2 Silk proteins, Fibroin and Sericin, have been distinguished as major components of Silk cocoons. In the process of manufacturing Silk, Fibroin is separated from Sericin by a degumming process. There are several methods for removing Sericin in the degumming process of cocoons. Because the method of production of Sericin involves extraction from cocoons using soaps and detergents, detergents and alkali soaps and are typically present as impurities in Sericin.

Hydrolyzed Silk is prepared from the cocoon of B mori. Hydrolyzed Silk protein (MW = 300 Da) may be prepared by acid, alkaline, or enzyme hydrolysis; Hydrolyzed Silk protein (650 Da) may be prepared by alkaline and enzyme hydrolysis.

In acute oral toxicity studies involving rats, LD $_{50}$ values of >10 g/kg body weight (Hydrolyzed Silk protein, MW ~300 Da), >5 g/kg (Hydrolyzed Silk protein, MW ~1000 Da), >16 g/kg (Silk), and >12 g/kg (30% aqueous Silk Powder) were reported. Signs of toxicity were not observed.

In a repeated insult patch test (RIPT, occlusive patches) involving dermal applications of Silk Powder (50% in sterile water) to 20 guinea pigs over a 3-week period, 2 animals died. However, necropsy results were not indicative of a test substance–related effect. Similarly, no abnormalities were noted at necropsy in a cumulative skin irritation in which 6.5% aqueous Hydrolyzed Silk (MW = 650 Da) was applied to 8 Hartley guinea pigs once daily for 35 days.

Sericin obtained via urea extraction was toxic to mouse fibroblasts in vitro at concentrations as low as $60 \mu g/mL$. When

compared to urea extraction, the extraction of Sericin by other methods resulted in reduced toxicity to mouse fibroblasts in vitro.

In the Ames test, results for 10% aqueous Hydrolyzed Silk protein were negative. Sericin and BSA were efficient inducers of rat RIN-5F cell proliferation in a rat insulinoma cell line. The results of another study involving CD-1 (ICR): Crj mice suggested that Sericin had a protective effect against tumor promotion in mouse skin by suppressing oxidative stress, inflammatory responses, and TNF- α .

The results of a study in mice suggested that Sericin possesses a photoprotective effect against UVB-induced damage and tumor promotion by reducing oxidative stress and cell proliferation in mouse skin. In another study involving mice, the feeding of diets supplemented with 1.5% or 3% Sericin for 5 weeks resulted in a dose-dependent decrease in the development of colonic aberrant crypt foci.

Undiluted Hydrolyzed Silk protein (MW ~300 Da; dose = $0.5 \text{ mL/}2.5 \text{ cm}^2$) caused reactions ranging from very slight to well-defined erythema on intact and abraded skin of rabbits. However, a primary irritation index (PII) of 1.1 was reported, and the test material was not classified as a primary skin irritant. Hydrolyzed Silk protein (MW ~1000 Da; dose = $0.5 \text{ mL}/2.5 \text{ cm}^2$; PII = 0.65) and Hydrolyzed Silk protein (MW = 650 Da; dose = $0.5 \text{ mL}/2.5 \text{ cm}^2$; PII = 0.05) were also classified as nonirritating to the skin of rabbits. Hydrolyzed Silk protein (15%-25% aqueous) was nonirritating to the skin of rabbits. In a cumulative skin irritation study involving guinea pigs, Hydrolyzed Silk protein (MW = 650 Da) was considered nonirritating to the skin. Hydrolyzed Silk protein (MW = 650) did not induce skin sensitization in a study involving guinea pigs. Silk Powder (0.5 g) was nonirritating to abraded or intact skin of rabbits, and Silk Powder (0.1 g) was nonirritating and non-sensitizing to the skin of guinea pigs.

There was no evidence of skin irritation in 6 rabbits after the application of Silk (0.5 g) under a 2 cm² patch for 24 hours. Silk Powder (0.5% wt/vol in distilled water) did not induce sensitization in an RIPT involving 20 guinea pigs. Also, the results of a preliminary skin irritation test at concentrations up to 10% in distilled water were negative. Similarly, Silk Powder (50% in sterile water) did not induce sensitization in an RIPT involving 20 guinea pigs. Furthermore, the results of a preliminary skin irritation test indicated that 75% Silk Powder was reasonably tolerated by the 5 guinea pigs tested.

Negative results were reported for Hydrolyzed Silk protein (MW = 300 Da) in a skin irritation study involving 20 subjects and for Hydrolyzed Silk protein (MW = 650) in a study involving 24 subjects. In 2 RIPTs involving 57 subjects and 49 subjects, respectively, Hydrolyzed Silk protein (MW $\sim\!1000$ Da; dose $\sim\!0.2$ mL/1" \times 3/4" area and dose $\sim\!2$ mL/4 cm² area, respectively) was not classified as a skin irritant or sensitizer. In a human repeated insult patch test involving 48 subjects, the results relating to the skin irritation and sensitization potential of Hydrolyzed Silk protein (MW $\sim\!300$ Da; dose = 20 μ L/40 mm² Finn chamber) were negative. Silk Powder (0.05 g) was nonirritating to the skin of 30 subjects.

Results for Hydrolyzed Silk were negative for skin irritation potential in the Irritection (dose volumes up to 125 μ L) and Epiderm (aqueous solution containing 27%-32% Hydrolyzed Silk) in vitro assays. The mouse local lymph node assay yielded negative results relating to the sensitization potential of 20% aqueous Hydrolyzed Silk protein (MW = 300 Da).

A 6.5% aqueous solution of Hydrolyzed Silk protein (MW = 650 Da; dose = 0.05 mL/2 cm2) was neither phototoxic nor photoallergenic to the skin of guinea pigs. Silk Powder (0.1 g) also was not phototoxic when applied to the skin of guinea pigs.

Hydrolyzed Silk protein of an average MW of ~ 300 Da, 650 Da, or ~ 1000 Da did not induce ocular irritation when instilled into the eyes of rabbits. The tests involving Hydrolyzed Silk protein (MW ~ 300 Da or ~ 1000 Da) were single-instillation tests, whereas Hydrolyzed Silk protein (MW = 650 Da) was instilled 3 times per day for 4 days. In another test, Hydrolyzed Silk protein (15%-25% aqueous, single instillation) was nonirritating to the eyes of rabbits. Hydrolyzed Silk protein (MW = 300 Da; 2% active solution) was also negative for ocular irritation potential in the HeT-CAM, Irritection, and Epiderm in vitro assays. Silk (0.1 g) was also classified as a nonirritant when instilled into the eyes of rabbits, and the results for 10% aqueous Silk Powder were slightly positive when instilled into the eyes of rabbits.

The suppression of inflammation by Sericin applied topically was reported in a study on rats. A hypersensitivity reaction was not observed.

An association between sensitization to Silk and asthma incidence was found in a study of 871 children. In another study of 64 children, the average mean wheal diameter elicited by ilk in prick tests was greater than 2 histamine equivalent prick tests. Allergenic proteins have been extracted from a Silk batch imported for use as filling material for bed mattresses and rugs. Immunoglobulin E and IgG antibodies to the extracted Silk proteins were measured in the sera of 9 Silk-sensitive subjects, and these sera were found to contain high titers of IgE and low titers of IgG antibodies to the separated silk polypeptides. In a study involving 10 patients with a positive skin prick test to silkworm crude extract, arginine kinase was identified as a major allergen in this crude extract.

A female patient with a history of severe atopic dermatitis and various allergies, and a family history of atopic eczema and asthma, presented with exacerbation of eczema on her hands and wrists and urticarial papules on the flexor aspects of both forearms. Radioallergosorbent tests yielded positive results for silk waste (k73) of 2.08 universal arbitrary units (Ua)/mL and positive results for Silk (k74) of 3.62 Ua/mL. The total IgE count was >5000 kilo units (kU)/L. These results confirmed the diagnosis of immunological contact urticarial caused by Silk.

In a case report, recurrent granulomas with remarkable infiltration of eosinophils may have resulted from an IgE-mediated hypersensitivity reaction to silk Fibroin. Additionally, type I allergic responses and upregulated levels of specific IgE have been reported in patients after repeated surgical procedures that involved the use of Silk sutures. In another case report, the

diagnosis of immunological contact urticarial caused by Silk was confirmed.

Skin depigmentation has been observed in renal patients after application of an 8% Sericin cream for treatment of dry and itchy skin. Histological examination of wounds in rats after 15 days of treatment with 8% Sericin cream revealed complete healing, no ulceration, and an increase in collagen, as compared to treatment with the control cream.

Discussion

After reviewing the available data, concerns pertaining to skin depigmentation, allergenicity, inhalation exposure, and the insufficiency of data on certain silk protein ingredients were discussed by the Panel. The Panel reviewed a study in which an 8% Sericin cream applied to the skin of renal patients caused depigmentation, but agreed that a review of ingredients for drug effects (ie, noncosmetic effects, such as depigmentation) is not within the Panel's purview. However, the Panel noted that an effect on cutaneous pigmentation would not be expected at the use concentrations of Silk protein ingredients in cosmetic products. Sericin is used at a maximum concentration of 0.00047% in cosmetic products.

Studies on patients inhalationally exposed to Silk waste via inhalation, which is distinct from the Silk protein ingredients that are being evaluated in this safety assessment, showed an association between asthma and dermal allergies to Silk in children in China. The Panel determined that the results of these studies do not support a cause-and-effect relationship of Silk protein ingredients in this safety assessment and the development of asthma.

Concern about pesticide residues and heavy metals that may be present in ingredients was expressed by the Panel. They stressed that the cosmetics industry should continue to use current good manufacturing practices (cGMPs) to limit impurities.

Some of the Silk protein ingredients are used in products that could be incidentally inhaled. For example, Hydrolyzed Silk and Silk Extract are used in hairspray at maximum concentrations up to 0.024% and 0.0036%, respectively, and the highest maximum use concentration for ingredient use in face powders is being reported for Silk Powder (1.4%). However, the Panel did not express concern over the use of these ingredients in formulations that might be inhaled. The Panel also noted that in aerosol products, 95% to 99% of droplets/particles would not be respirable to any appreciable amount. Furthermore, droplets/particles deposited in the nasopharyngeal or bronchial regions of the respiratory tract present no toxicological concerns based on the chemical and biological properties of these ingredients and their low use concentrations. Coupled with the small actual exposure in the breathing zone and the concentrations at which the ingredients are used, the available information indicates that incidental inhalation would not be a significant route of exposure that might lead to local respiratory or systemic effects. A detailed discussion and summary of the Panel's approach to evaluating incidental inhalation exposures to ingredients in cosmetic products is available at https://www. cir-safety.org/cir-findings.

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The Panel determined that the available data are insufficient for evaluating the safety of MEA-Hydrolyzed Silk and Silkworm Cocoon Extract in cosmetic products and that the following data on these ingredients are needed:

- Method of manufacture and impurities
- Concentration of use
- 28-day dermal toxicity study; if absorbed, genotoxicity and reproductive and developmental toxicity data may be needed
- Skin irritation and sensitization data

Conclusion

The Panel concluded that the following 8 ingredients are safe in the present practices of use and concentration in cosmetics, as described in this safety assessment.

Fibroin*
Hydrolyzed Fibroin
Hydrolyzed Sericin
Hydrolyzed Silk
Sericin
Silk
Silk Extract
Silk Powder

*Not reported to be in current use. Were this ingredient not in current use to be used in the future, the expectation is that it would be used in product categories and at concentrations comparable to others in this group.

The Panel also concluded that the available data are insufficient for to determine the safety of 2 silk protein ingredients in cosmetic products, MEA-Hydrolyzed Silk and Silkworm Cocoon Extract. Neither of these ingredients is reported to be in use.

Author's Note

Unpublished sources cited in this report are available from the Executive Director, Cosmetic Ingredient Review, 1620 L Street, NW, Suite 1200, Washington, DC 20036, USA.

Author Contributions

Johnson contributed to conception and design; acquisition, analysis, and interpretation; and drafted manuscript. Heldreth contributed to conception and design; acquisition, analysis, and interpretation; drafted manuscript; and critically revised manuscript. Bergfeld, Belsito, Hill, Klaassen, Liebler, Marks, Shank, Slaga, Snyder, and Gill contributed to conception and design; acquisition and interpretation; and critically revised the manuscript. All authors gave final approval and agree to be accountable for all aspects of work ensuring integrity and accuracy.

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Table 1. Definitions and Reported Functions of the Ingredients in This Safety Assessment. 1

Ingredient/CAS No.	Definition	Function
Fibroin 9007-76-5	Fibroin is a protein filament produced by the silkworm, Bombyx mori which together with Sericin composes Silk.	Bulking Agents
Hydrolyzed Fibroin	Hydrolyzed Fibroin is the hydrolysate of Fibroin derived by acid, enzyme, or other method of hydrolysis.	Hair Conditioning Agents; Skin Conditioning Agents-Miscellaneous
Hydrolyzed Sericin 870616-36-7 73049-73-7	Hydrolyzed Sericin is the hydrolysate of Sericin derived by acid, enzyme, or other method of hydrolysis.	Hair Conditioning Agents; Skin Conditioning Agents-Miscellaneous
Hydrolyzed Silk 73049-73-7 96690-41-4	Hydrolyzed Silk is the hydrolysate of Silk protein derived by acid, enzyme, or other method of hydrolysis.	Hair Conditioning Agents; Skin Conditioning Agents-Miscellaneous
MEA-Hydrolyzed Silk	MEA-Hydrolyzed Silk is the monoethanolamine salt of Hydrolyzed Silk	Hair Conditioning Agents; Skin Conditioning Agents-Miscellaneous
Sericin 60650-88-6 60650-89-7	Sericin is a protein isolated from the Silk produced by the silkworm, <i>B mori</i> .	Hair Conditioning Agents; Skin Conditioning Agents-Miscellaneous
Silk	Silk is the fibrous protein obtained from cocoons of the silkworm.	Bulking Agents
Silk Extract 91079-16-2	Silk Extract is the extract of silk fiber.	Skin Conditioning Agents-Miscellaneous
Silk Powder 9009-99-8	Silk Powder is finely pulverized Silk.	Bulking Agents; Skin Conditioning Agents– Miscellaneous; Slip Modifiers
Silkworm Cocoon Extract 91079-16-2	Silkworm Cocoon Extract is the extract of the cocoon of the silkworm, <i>B mori</i> .	Skin Conditioning Agents-Humectant

 Table 2. Chemical Properties of Silk Proteins.

Property	Value	Background Information
Sericin		
Form	Powder; amorphous structure ⁹	Transforms into a β -sheet structure in presence of water. ⁹ Easily dissolves in water at 50 °C to 60 °C; returns to gel form on cooling. ² Gelation is rapid at 108 °C and pH \approx 6 to 7. ⁹
Molecular weight	10 to >400 kDa ⁹ 35 to 150 kDa ⁹ 15 to 75 kDa ⁹ 10 to >225 kDa ⁹ <20 kDa ⁹ > 20 kDa ⁹	Depending on extraction methods, temperature, pH, and processing time. Heat and acid extraction. Alkaline solution extraction. Urea extraction. Recovered during early stages of raw Silk production. Obtained from later stages of raw Silk production.
Solubility	Highly soluble in water ²⁹	Decreases when molecules are transformed from random coil into the β -sheet structure. Isoelectric point \sim 4, because there are more acidic than basic amino acids in Sericin. 26
Fibroin		
Form	Pale yellow mass 10	
Molecular weight	300 to 420 kDa ⁷⁶	
Solubility	Soluble in concentrated alkalies, concentrated mineral acids, and in ammoniacal nickel oxide solution. Insoluble in water, alcohol, ether, and dilute alkalies. 10	
Hydrolyzed Fibroin		
Form	Yellow solution ⁷⁷	Acid hydrolysis usually causes Fibroin solution to turn yellow, and chemical changes in amino acids such as tryptophan and tyrosine are generally considered as the main reason for yellowing. Tryptophan and tyrosine become yellow upon hydrolysis, and the same is true for serine and glycine. Serine and threonine beak down easily during hydrolysis, and other amino acids in Fibroin decompose in the following order: tyrosine, methionine, cysteine, phenylalanine, and tryptophan. ⁷⁷
Hydrolyzed Silk		
Form	Amber liquid ²²	
Odor	Characteristic ²²	
Molecular weight Solubility	<10 kDa ²³ ; 2-4 kDa ²⁴ ; 300 Da and 650 Da ^{14,15} Soluble in water ²³	
Density (at room temperature)	1.05 to 1.11 g/mL ²²	
Silk		
Appearance Particle Size	White to slightly gray powder 12 5 to $15~\mu m^{12}$	

Table 3. Composition Data on Hydrolyzed Silk. 20,21

	Silk Hydrolysate (g/100 g protein)
Cysteic Acid	0
Hydroxyproline	0
Aspartic Acid	7.4
Threonine	3.9
Serine	17.4
Glutamic Acid	3.6
Proline	0.9
Glycine	20.6
Alanine	21
Cystine	0
Valine	3.7
Methionine	0.4
Isoleucine	0.8
Leucine	1.2
Tyrosine	12.2
Phenylalanine	2
Lysine	1.6
Histidine	0.9
Arginine	2.4
Tryptophan	NR
Lysinoalanine	NR

Abbreviation: NR, not reported.

Table 4. Frequency and Concentration of Use According to Duration and Type of Exposure. ^{30,31}

	Hydrolyzed Silk		Hydrolyzed Fibroin		Sericin	
	# of Uses	Conc. (%)	# of Uses	Conc. (%)	# of Uses	Conc. (%)
Totals/Conc. Range	675	0.0000007-0.5	ı	0.001-0.045	44 ().00047
Duration of Use						
Leave-On	348	0.0000007-0.23	I	0.001		0.00047
Rinse off	322	0.0000007-0.5	NR	0.045	14	0.00047
Diluted for (bath) Use	5	0.0001-0.0003	NR	NR	1	NR NR
Exposure Type						
Eye Area	36	0.0000007-0.23	NR	NR	10	0.00047
Incidental Ingestion	3	NR	NR	NR	NR	NR
Incidental Inhalation- Sprays	NR	0.0005-0.024	1***	NR	1	0.000047
Incidental Inhalation- Powders	NR	0.00023-0.11**	1***	0.001**	1	0.000047
Dermal Contact	213	0.0000007-0.11	I	NR	12	0.000047
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair–Non-Coloring	189	0.00002-0.5	NR	NR	NR	0.000047
Hair–Coloring	6	0.00022-0.008	NR	0.045	12	NR
Nail	I	0.01-0.03	NR	NR	NR	NR
Mucous Membrane	126	0.0001-0.035	NR	NR	I	NR
Baby Products	4	0.0003	NR	NR	NR	NR
		Silk	Silk	Extract	Silk	Powder
	# of Uses	Conc. (%)	# of Uses	Conc. (%)	# of Uses	Conc. (%)
Totals/Conc. Range	27	0.000005-0.2	13	0.0013-0.015	177	0.0001-1.4
Duration of Use						
Leave-On	23	0.000005-0.2	7	0.0036-0.015	163	0.0001-1.4
Rinse off	4	NR	6	0.0013-0.015	14	0.0001-0.1
Diluted for (bath) Use	NR	NR	NR	NR	NR	NR
Exposure Type						
Eye Area	I	0.000005	I	NR	33	0.05-1.1
Incidental Ingestion	NR	NR	NR	NR	16	0.05-1.1
Incidental Inhalation- Sprays	NR	NR	3*	0.0036	1	0.005-0.1
Incidental Inhalation- Powders	4	0.1-0.2	NR	0.1-0.2	60	0.1-1.4
Dermal Contact	17	0.000005-0.2	I	0.0013	67	0.0001-1.4
Deodorant (underarm)	NR	NR	NR	NR	NR	0.005
Hair—Non-Coloring (NR	NR	7	0.0036-0.015	6	0.01-0.048
Hair-Coloring	NR	NR	NR	NR	NR	NR
Nail	5	0.001-0.05	NR	NR	7	0.0001-0.025
Mucous Membrane	3	NR	NR	NR	19	0.01-1.1
Baby Products	NR	NR	NR	NR	NR	NR

Table 4. Current Frequency and Concentration of Use According to Duration and Type of Exposure. 30,31

	Hydrolyzed Sericin	
	# of Uses	Conc. (%)
Fotals/Conc. Range	4	NR
Duration of Use		
Leave-On	4	NR
Rinse off	NR	NR
Diluted for (bath) Use	NR	NR
Exposure Type		
Eye Area	I	NR
Incidental Ingestion	NR	NR
Incidental Inhalation- Sprays	 *	NR
Incidental Inhalation- Powders	NR	NR
Dermal Contact	I	NR
Deodorant (underarm)	NR	NR
Hair-Non-Coloring	2	NR
Hair-Coloring	NR	NR
Nail	NR	NR
Mucous Membrane	NR	NR
Baby Products	NR	NR

Abbreviation: NR, not reported.

Note: Because each ingredient may be used in cosmetics with multiple exposure types, the sum of all exposure type uses may not equal the sum total uses.

Table 5. Skin Irritation/Sensitization Potential of Silk Ingredients.

Skin Irritation and Sensitization—Non-Human

Hydrolyzed Silk

6 female New Zealand albino rabbits. Undiluted test material applied (0.5 ml) for 24 h to abraded or intact skin using 2.5 cm² occlusive patch. Reactions (scored I h after patch removal) ranged from very slight to well-defined erythema at both intact and abraded test sites. Primary irritation score (6 rabbits) = I.I. According to Draize system, combined averages (primary irritation scores) of 2 or less classified as mildly irritating. Mild skin irritant.⁷⁸

6 New Zealand white rabbits. 15% to 25% aqueous solution (0.5 ml) applied for 24 h to abraded and intact sites on trunk (2.5 cm² area per site) using occlusive patch. The test sites examined at 24 h and 72 h post-application. Nonirritant (PII = 0.65).³⁹

6 New Zealand white rabbits. 6.5% aqueous solution applied to back (2.5 \times 2.5 cm area) according to preceding test procedure. Very slight erythema in one rabbit. Nonirritant (PII = 0.05).⁷⁹

8 Hartley guinea pigs. 6.5% aqueous solution applied to back once daily for 35 days. After 31 days, the only reaction observed was very slight erythema in 3 animals. Nonirritant.⁴²

Groups of 6 Hartley guinea pigs. 6.5% aqueous solution. Maximization test. Subcutaneous injection and dermal application during induction. Dose per cm² not stated. 24-h occlusive challenge patch (0.2 ml test material) applied to dorsal skin on day 14. Reactions scored at 24 h and 48 h after challenge patch removal. I of 6 animals had moderate erythema and 2 of 6 animals had slight erythema at 24 h after patch removal. Two of 6 animals had slight erythema at 48 h after patch removal. Non-sensitizer. Positive control (0.1% 4-dinitrochlorobenzene) induced sensitization.⁸⁰

Mouse local lymph node assay (OECD Guideline No. 429). 20% aqueous Hydrolyzed Silk (MW = 300 Da) was non-sensitizer. ^{14,81} Silk

6 adult New Zealand albino rabbits. Test material (0.5 g in solvent (unnamed)) applied to intact area and abraded area on the back for 24 h; each site covered with 2 cm² occlusive patch. Reactions scored at 24 h and 72 h. No evidence of erythema, eschar, or edema at abraded or intact sites. Silk classified as nonirritant.⁴⁰

(continued)

Totals = Rinse-off + Leave-on + Diluted for (Bath) Use Product Uses.

^{*}It is possible that these products may be sprays, but it is not specified whether the reported uses are sprays.

^{**}It is possible that these products may be powders, but it is not specified whether the reported uses are powders.

^{***}Not specified whether a powder or spray, so this information is captured for both categories of incidental inhalation.

Table 5. (continued)

Skin Irritation and Sensitization—Non-Human

Silk Powder

6 female albino rabbits. Test material (0.5 g, under an occlusive patch) was applied for 4 h to abraded and intact sites (1 inch² area) on the back. Sites examined at 24 h. No skin irritation reactions at intact or abraded skin sites.⁴¹

Female guinea pigs (number of animals and strain not stated). Test material (0.1 g) applied to back (cm² area not stated) 5 times per week for 13 weeks. No irritation or sensitization reactions.⁴¹

20 Hartley albino guinea pigs (10 males, 10 females). Test material (5% w/v in distilled water; 0.5 ml on occlusive patch (Hill Top Chamber)) applied for 6 h to left shoulder once per week for a total of three 6-h applications. Dose per cm² not stated. After 2-week nontreatment period, challenge patch applied for 24 h to new site. After patch removal, depilatory applied to challenge site for 30 minutes. Reactions scored at ~ 2 h after depilation. Ten guinea pigs (controls) tested with distilled water. Nonirritant and non-sensitizer (test and controls). 82

20 female Dunkin-Hartley guinea pigs. Test material (50% in sterile water; 0.5 ml on 20 x 20 mm occlusive patch) applied repeatedly for 6 h to left flank (induction). Reactions were scored at 24 h after patch removal. This procedure repeated at weekly intervals (Days 8 to 9 and 15 to 16 of study). At day 29, challenge patch applied to right flank for 6 h. Reactions scored 24 h and 48 h after patch removal. Sterile water applied to 10 control guinea pigs. Non-sensitizer (test and control animals). Silk Powder (75%) reasonably tolerated in preliminary skin irritation test involving 5 guinea pigs. ⁴³

Skin Irritation and Sensitization—Human

Hydrolyzed Silk

20 subjects (2 men, 18 women). 20% aqueous Hydrolyzed Silk (\sim 3 mg on occlusive patch [Finn chamber]) applied to back for 48 h. Dose per cm² not stated. At 30 minutes after patch removal, mild erythema observed in 3 subjects. Mild skin irritation in only 1 of the 3 subjects at 24 h post-removal. No skin irritation in remaining 17 subjects. ^{14,83}

24 subjects (10 men, 14 women). 6.5% aqueous solution (0.2 ml on occlusive patch) applied to upper back for 24 h. Dose per cm² not stated. Classified as a nonirritant.⁸⁴

57 male and female subjects. HRIPT. During induction, Hydrolyzed Silk (~ 0.2 ml; concentration not stated) applied to upper back repeatedly using a 1" x 3/4" semi-occlusive patch. Challenge patch applied to original site and to new site on forearm. Application sites evaluated at 24 h and 48 h post-application. Nonirritant and non-sensitizer. 62

49 male and female subjects. HRIPT. Semi-occlusive patch (2 cm x 2 cm) containing ~ 2 ml Hydrolyzed Silk (average MW = 1000 Da; concentration not stated) applied to back repeatedly during induction. Challenge patch applied for 24 h to new site. Application sites evaluated at 24 h and 48 h post-application. Transient erythema (nonirritant and non-allergic in nature) observed during induction; I subject with cumulative skin irritation reaction after removal of 9th induction patch. Subject also had barely perceptible erythema at challenge site (48-h reading). No clinically significant irritation or evidence of allergic contact dermatitis.

48 subjects. HRIPT. During induction, 20% aqueous Hydrolyzed Silk (20 μ l on occlusive patch (8 mm diameter Finn chamber; 40 mm² surface)) applied for 48 h to the back repeatedly. 48-h challenge patch applied to new site on back. Nonirritant and non-sensitizer.

Silk Powder

30 male and female subjects. Silk Powder (0.05 g) applied for 48 h, under a closed patch, to the arm of each subject. Examinations for dermal reactions after 1 h and 24 h. No skin irritation.⁴¹

Skin Irritation and Sensitization—In Vitro

Hydrolyzed Silk

The Irritection assay. In vitro system that involves use of a proprietary solution containing both proteins and macromolecules in a well that is covered by a membrane. Hydrolyzed Silk applied to the Irritection system at dose volumes of 25 μ l, 50 μ l, 75 μ l, 100 μ l, and 125 μ l. Irritation measured quantitatively using a spectrophotometer. Nonirritant (Ocular Irritection scores: 0.25 to 0.40).

EpiDerm model assay. Test material (30 μl of aqueous solution containing 27% to 32% Hydrolyzed Silk) applied to human epidermal-derived keratinocytes (cultured to form a multilayer). Nonirritant.⁶⁹

References

- 1. Nikitakis J, Breslawec HP. *International Cosmetic Ingredient Dictionary and Handbook*. 14 ed. Personal Care Products Council; 2014.
- 2. Joseph J, Raj SJ. Therapeutic applications and properties of Silk proteins from *Bombyx mori. Front Life Sci.* 2012;6(3-4): 55-60.
- Fiume MM, Heldreth B, Bergfeld WF, et al. Final amended report on the safety assessment of ethanolamine and ethanolamine salts as used in cosmetics (Unpublished). Available from the Cosmetic Ingredient Review, 1620 L Street, N.W., Suite 1200, Washington, D.C. 20036. 2012:1-17.
- Mondal M, Trivedy K, Kumar SN. The Silk proteins, Sericin and Fibroin in Silkworms, Bombyx mori Linn – a review. Caspian J Env Sci. 2007;5(2):63-76.
- Asakura T, Suzuki Y, Nakazawa Y, Yazawam K, Holland GP, Yarger JL. Silk structure studied with nuclear magnetic resonance. Prog Nucl Magn Reson Spectrosc. 2013;69:23-68.
- Kweon HY, Yeo JH, Lee KG, et al. Effects of poloxamer on the gelation of Silk Sericin. *Macromol Rapid Commun*. 2000;21: 1302-1305.
- Sheng HY, Hong L, Lei W, Shun YY, Hoo A. Studies on the moicrostructure and physico-chemical properties of diffluent Sericin powder. Silk. 2000;2:238-243.
- Takasu Y, Yamada H, Tsubouchi K.Extraction and chromatographic analysis of cocoon Sericin of the Silkworm, Bombyx mori. *J Insect Biotechnol Sericol*. 2002;71(3):151-156.
- 9. Aramwit P, Siritientong T, Srichana T. Potential applications of Silk Sericin, a natural protein from textile industry by-products. *Waste Manag Res.* 2012;30(3):217-224.
- 10. O'Neal MJ. *The Merck Index: An Encyclopedia of Chemicals, Drugs, and Biologicals.* 15th ed. Merck & Co; 2013.
- 11. Altman GH, Diaz F, Jakuba C, et al. Silk-based biomaterials. *Biomaterials*. 2003;24:401-416.
- Personal Care Products Council. Method of production and properties of Silk. Unpublished data submitted by the Personal Care Products Council on March 13, 2015. 2015:1.
- Arch Personal Care Products LP. Solu-Silk Protein SF (Hydrolyzed Silk) Manufacturing Process. Unpublished data submitted by the Personal Care Products Council on May 14, 2012. 2012:1-8.
- Anonymous. Information of Hydrolyzed Silk Protein-1 (method of manufacture; molecular weight; impurities; summary of safety data). Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 2012:1-2.
- 15. Anonymous. Information of Hydrolyzed Silk Protein-2 (method of manufacture, molecular weight, impurities, summary of safety data). Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 2012:1-2.
- 16. Personal Care Products Council. Method of production and composition data. Hydrolyzed Silk, Hydrolyzed Silk protein-1, and Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on April 3, 2015. 2015:1.
- 17. Proalan s/a. NorSilk (INCI: Hydrolyzed Silk): manufacturing flow diagram. Unpublished data submitted by the Personal Care Products Council on March 13, 2015. 2014:1.

- 18. Active Concepts. 20621-AC Silk hydrolysate—manufacturing flow chart. Unpublished data submitted by the Personal Care Products Council on MArch 25, 2015. 2015:1.
- Active Concepts. 20625-AC Silk hydrolysate H—manufacturing flow chart. Unpublished data submitted by the Personal Care Products Council on March 25, 2015. 2015:1.
- Arnaud JC, Boré P.Ion-exchange chromatography for the analysis of protein derivatives and specific amino acids. Chapter: 8. In: Boré P, ed. Cosmetic Analysis Selective Methods and Techniques. 1980:223-224.
- 21. De Groot AP, Slump P. Effects of severe alkali treatment of proteins on amino acid composition and nutritive value. *J Nutr*. 1969;98(1):45-56.
- 22. Proalan s/a. NorSilk (INCI: Hydrolyzed Silk): technical information. Unpublished data submitted by the Personal Care Products Council on March 13, 2015. 2014:1.
- Proalan s/a. NorSilk (INCI: Hydrolyzed Silk): material safety data sheet. Unpublished data submitted by the Personal Care Products Council on March 13, 2015. 2014:1-7.
- Personal Care Products Council. Molecular weight and composition data on Hydrolyzed Silk ingredients. Unpublished data submitted by the Personal Care Products Council on March 25, 2015. 2015:1.
- Heslot H.Artificial fibrous proteins: a review. *Biochimie*. 1998; 80:19-31.
- 26. Padamwar MN, Pawar AP. Silk Sericin and its applications: a review. *J Sci Ind Res.* 2004;63(4):323-329.
- 27. Forlani G, Seves AM, Ciferri O. A bacterial extracellular proteinase degrading Silk Fibroin. *Int Biodeter Biodegr*. 2000;46(4): 271-275.
- 28. Kim E, Bayaraa T, Shin E, Hyun C. Fibroin-derived peptides stimulate glucose transport in normal and insulin-resistant 3T3-L1 adipocytes. *Biol Pharm Bull*. 2009;32(3):427-433.
- 29. Aramwit P.Bio-response to Silk Sericin. Chapter: 11. In: Kundu S, ed. *Silk Biomaterials for Tissue Engineering and Regenerative Medicine*. Woodhead Publishing: 2014:299-329:74.
- 30. Food and Drug Administration. *Information Supplied to FDA by Industry as Part of the VCRP FDA Database*. FDA; 2015.
- 31. Personal Care Products Council. Concentration of use by FDA product category: Silk. Unpublished data submitted by the Personal Care Products Council on January 6, 2015. 2015:1.
- 32. Rothe H, Fautz R, Gerber E, et al. Special aspects of cosmetic spray safety evaluations: principles on inhalation risk assessment. *Toxicol Lett.* 2011;205(2):97-104. PM:21669261.
- Bremmer HJ, Prud'homme de Lodder LCH, van Engelen JGM.
 Cosmetics fact sheet: to assess the risks for the consumer.
 Updated version for ConsExpo 4. 2020:1-77. Accessed August 24, 2011. http://www.rivm.nl/bibliotheek/rapporten/320104001.
 pdf.ReportNo.RIVM320104001/2006
- Rothe H. Special aspects of cosmetic spray evaluation. Unpublished information presented to the 26 September CIR Expert Panel. Washington, DC. 2011.
- 35. Johnsen MA. The influence of particle size. *Spray Technol Mark*. 2004;14(11):24-27.
- 36. European Union. Regulation (EC) No. 1223/2009 of the European Parliament and of the Council, of November 30, 2009, on

Cosmetic Products. Amended by Commission regulation (EU) No. 1004/2014 of September 18, 2014. 2009.

- Food and Drug Administration. General and plastic surgery devices. Natural nonabsorbable Silk surgical suture. 21CFR: 878.5030. 2014.
- Toxicol Laboratories Limited. Single dose oral toxicity study in the rat. Hydrolyzed Silk (MW ~ 300 Da). Study Ref. No. 108/ 8412. Unpublished data submitted by the Personal Care Products Council on July 3, 2012. 1985:1-6.
- 39. Consumer Product Testing Co. Primary dermal irritation in rabbits; primary ocular irritation in rabbits; acute oral toxicity in rats: Hydrolyzed Silk (MW ~ 1,000 Da). Experiment Reference No.: 85084-3. Unpublished data submitted by the Personal Care Products Council on July 3, 2012. 1985:1-12.
- Applied Biological Sciences Laboratory. Summary information on Silk: Skin irritation, eye irritation, and acute oral toxicity. Unpublished data submitted by the Personal Care products Council on March 13, 2015. 1980:1.
- 41. Anonymous. Toxicological data (primary irritation test on rabbit skin; rabbit eye Draize test; accumulative skin irritation test; phototoxicity test; acute oral toxicity test; human closed patch test. Unpublished data submitted by the Personal Care Products Council on May 1, 2015. 1978:1-16.
- Faculty Medicine, Hiroshima University. Cumulative application (skin) test. Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1985:1.
- Research Toxicological Centre S.p.A. Silk powder: delayed dermal sensitization study in the guinea pig. Unpublished data submitted by the Personal Care Products Council on March 11, 2015. 1995:1-20.
- 44. Aramwit P, Kanokpanont S, Nakpheng T, Srichana T. The effect of Sericin from various extraction methods on cell viability and collagen production. *Int J Mol Sci.* 2010;11(5):2200-2211.
- Aramwit P, Keongamaroon O, Siritientong T, Bang N, Supasyndh
 Sericin cream reduces pruritis in hemodialysis patients. *BMC Nephrol*. 2012;13:119.
- Aramwit P, Damrongsakkul S, Kanokpanont S, Srichana T. Properties and anti-tyrosinase activity of Sericin from various extraction methods. *Biotechnol Appl Biochem*. 2010;55(2):91-98.
- NAmSA. Hydrolyzed Silk protein: Genotoxicity; Salmonella typhimurium reverse mutation study. Unpublished data submitted by the Personal Care Products Council on May 1, 2015. 1997: 1-9.
- 48. Ogawa A, Terada S, Kanayama T, et al. Improvement of islet culture with Sericin. *J Biosci Bioeng*. 2004;98(3):217-219.
- 49. Zhaorigetu S, Yanaka N, Sasaki M, Watanabe H, Kato N.Silk protein, Sericin, suppresses DMBA-TPA-induced moue skin tumorigenesis by reducing oxidative stress, inflammatory responses and endogenous tumor promoter TNF-alpha. *Oncol Rep.* 2003;10(3):537-543.
- Sasaki M, Kato N, Watanabe H, Yamada H. Silk protein Sericin suppresses colon carcinogenesis induced by 1,2-dimethylhydrazine in mice. *Oncol.Rep.* 2000;7(5):1049-1052.
- 51. Zhaorigetu S, Yanaka N, Sasaki M, Watanabe H, Kato N. Inhibitory effects of Silk protein, sericin on UVB-induced acute damage

- and tumor promotion by reducing oxidative stress in the skin or hairless mouse. *J Photochem Photobiol*. 2003;71(1-3):11-17.
- 52. Caledon JC, Palmer LJ, Xu X, Wang B, Fang Z, Weiss ST. Sensitization to Silk and childhood asthma in rural China. *Pediatrics*. 2001;107(5):E80.
- 53. Wen CM, Ye ST, Zhou LX, Yu Y. Silk-induced asthma in children: a report of 64 cases. *Ann.Allergy*. 1990;65(5):375-378.
- 54. Dewair M, Baur X, Ziegler K. Use of immunoblot technique for detection of human IgE and IgG antibodies to individual Silk proteins. *J Allergy Clin Immunol*. 1985;76(4):537-542.
- Liu Z, Xia L, Wu Y, Xia Q, Chen J, Roux KH. Identification and characterization of an arginine kinase as a major allergen from Silkworm (*Bombyx mori*) larvae. *Int Arch Allergy Immunol*. 2009; 150(1):8-14.
- Faculty Medicine, Hiroshima University. Phototoxicity test.
 Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1985:1-2.
- 57. Faculty Medicine, Hiroshima University. Photoallergenicity test. Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1985:1-2.
- Kurosaki S, Otsuka H, Kunitomo M, Koyama M, Pawankar R, Matumoto K.Fibroin allergy. IgE mediated hypersensitivity to Silk suture materials. *Nihon Ika Daigaku Zasshi*. 1999;66(1): 41-44.
- Soong HK, Kenyon KR. Adverse reactions to virgin Silk sutures in cataract surgery. *Ophthalmology*. 1984;91(5):479-483.
- Rossitch D Jr, Bullard DE, Oakes WJ. Delayed foreign-body reaction to Silk sutures in pediatric neurosurgical patients. *Childs Nerv Syst.* 1987;3:375-378.
- 61. Vandevenne A, Morren M, Goossens A. Immunological contact urticaria caused by a Silk shirt in an atopic patient. *Contact Dermatitis*. 2015;72(4):240-241.
- 62. Consumer Product Testing Co. Repeated insult [patch test: Hydrolyzed Silk (MW ~ 1,000 Da). Experiment Reference Number: C97-0170. Unpublished data submitted by the Personal Care Products Council on July 3, 2012. 1997:1-10.
- Toxicol Laboratories Limited. Eye irritation study on Hydrolyzed Silk. Unpublished data submitted by the Personal Care Products Council on July 3, 2012. 1985:1-6.
- 64. Faculty Medicine, Hiroshima University. Primary eye irritation test. Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1985:
- Faculty Medicine, Hiroshima University. Cumulative application (eye) irritation test. Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 12, 2012. 1985:1-2.
- Anonymous. Het-Cam test. Hydrolyzed Silk protein-1. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 2003:1-3.
- Anonymous. RBCA test. Hydrolyzed Silk protein-1. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 2003:1-4.
- Active Concepts. AC Silk hydrolysate irritation analysis. Unpublished data submitted by the Personal Care Products Council on March 25, 2015. 2009:1-2.

- Active Concepts. Dermal and ocular irritation tests (AC Silk hydrolysate H). Unpublished data submitted by the Personal Care Products Council on March 25, 2015. 2015:1-4.
- Panilaitis B, Altman GH, Chen J, Jin HJ, Karageorgiou V, Kaplan DL. Macrophage responses to Silk. *Biomaterials*. 2003;24(18): 3079-3085.
- Aramwit P, Kanokpanont S, De-Eknamkul W, Srichana T. Monitoring of inflammatory mediators induced by Silk Sericin. *J Biosci Bioeng*. 2009;107(5):556-561.
- Aramwit P, Towiwat P, Srichana T. Anti-inflammatory potential of Silk Sericin. Nat Prod Commun. 2013;8:501-504.
- Aramwit P, Sangcakul A. The effects of Sericin cream on wound healing in rats. Biosci Biotechnol Biochem. 2007;71(10):2473-2477.
- Aramwit P, Kanokpanont S, Punyarit P, Srichana T. Effectiveness and inflammatory cytokines induced by Sericin compared to Sericin in combination with silver sulfadiazine cream on wound healing. Wounds. 2009;21(8):198-206.
- Tsubouchi K, Igarashi Y, Takasu Y, Yamada H. Sericin enhances attachment of cultured human skin fibroblasts. *Biosci Biotechnol Biochem*. 2005;69(2):403-405.
- 76. Guhrsa K, Weisshart K, Grosse F. Lessons from nature—protein fibers. *Rev Mol Biotechnol*. 2000;74(2):121-134.
- Kaili C, Ayub ZR, Hirabayashi K. Possible involvement of serine and glycine in the yellowing of acid Hydrolyzed Silk. *J Ser Sci*. 1996;65(2):109-113.
- Toxicol Laboratories Limited. Primary skin irritation study: Hydrolyzed Silk (MW ~ 300 Da). Study Ref. No. 109/8412. Unpublished data submitted by the Personal Care Products Council on July 3, 2012. 1985.

- Faculty Medicine, Hiroshima University. Primary skin irritation test. Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1985:1-2.
- Faculty Medicine, Hiroshima University. Skin sensitization test.
 Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1985:1-2.
- SafePharm Laboratories. Local lymph node assay in the mouse.
 Hydrolyzed Silk protein-1. SPL Project Number: 1268/096.
 Unpublished data submitted by the personal Care Products Council on July 11, 2012. 2003:1-11.
- Springborn Institute for Bioresearch, Inc. Guinea pig sensitization screen: Silk Powder. Unpublished data submitted by the Personal Care Products Council on July 11, 2015. 1985:1-18.
- 83. Dermis Research Center Co., Ltd. Human patch test under occlusive patch for 48 hours. Hydrolyzed Silk protein-1. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 2003:1-6.
- 84. Osaka City Institute of Public Health Sciences. Human patch test. Hydrolyzed Silk protein-2. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 1984:1.
- 85. Essex Testing Clinic. Repeated insult patch test: Hydrolyzed Silk (MW ~ 1,000 Da). ETC Entry No. 0194a. Unpublished data submitted by the Personal Care Products Council on July 3, 2012. 1985:1-5.
- 86. Aster Cosmétologie. Study of a cosmetic product, Hydrolyzed Silk protein-1, according to the repeated patch test method. Ref. Aster PC3354. Unpublished data submitted by the Personal Care Products Council on July 11, 2012. 2004:1-13.